
Secondary Metabolites from Microorganisms Isolated by the HPFiedler Group



AG Mikrobiologie/Antibiotika

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by

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Dedicated to late Professor Hans Zähner

I. Introduction

A more than 20-years period in the search for secondary metabolites from microorganisms provides the opportunity of looking back to the beginning of the research and summarizing an overview on the discovered and published novel natural compounds. First of all, I wish to acknowledge my appreciation of the outstanding work of my doctoral father and mentor Professor Hans Zähner, who founded in 1964 the Institute of Microbiology at the University of Tübingen. Both my diploma thesis in 1972 and my doctoral thesis in 1974 were performed under his supervision. The stimulating atmosphere in his huge team and the excellent technical facilities contributed mainly to the success of my own research. Hans Zähner's retirement in 1994 was an encouragement to continue his fascinating scientific work in screening and discovery of novel biologically active secondary metabolites produced by new isolated members of the bacterial order *Actinomycetales*.

Looking back to the late 1980s, it was a promising decision to establish a natural product library based on high-performance liquid chromatography and diode-array monitoring (HPLC-DAD) with the aim to identify or to characterize secondary metabolite profiles of freshly isolated actinomycetes strains in culture filtrates and crude extracts. In 1984 the first fully automated HPLC systems equipped with a diode array monitoring system was introduced to the market. Hewlett-Packard (Waldbronn, Germany) was our main partner in optimizing hardware and software suitable for routine HPLC analysis. At the late 1980s, a first fast computing Pascal Workstation and hard disks having capacities of 10-20 MB (!) were available, even at an exorbitant price compared to the current market situation. This 'high-tech' equipment permitted us to create an efficient data base on natural products which was built up in my group starting in 1988 (Fiedler, 1993). At this time no further coupling techniques with more efficient monitoring devices, e.g. mass spectrometry, were imaginable. Therefore, *ortho*-phosphoric acid was chosen as the most suitable acidic modifier added to the mobile phase, having optimal properties in separation selectivity and UV transmission. The separation was performed by a standardized linear gradient system from 0.1% aqueous *ortho*-phosphoric acid to 100% acetonitrile within 15 minutes. Nucleosil-100 C₁₈ reversed-phase material (5-μm particles; Macherey & Nagel, Germany) was selected as stationary phase because of its less adsorptive properties, high resolution and stability of retention times. This system continued to work excellently today.

New secondary metabolites from *Streptomyces* Tü strains

During the years 1988-1998 the research of my group was mainly sponsored by the Ministry of Research BMBF, who provided most of the fermentation and HPLC equipment. This period was the beginning of establishing my own group for isolating new actinomycetes strains from soil samples which were collected at various sites

throughout the world, and resulted in the discovery of following new secondary metabolites in strains of the genus *Streptomyces*:

- **Arylomycin A and B series:** lipopeptide antibiotics produced by *Streptomyces* sp. Tü 6075, antibacterial active, novel inhibitors of bacterial signal peptidase (Schimana *et al.*, 2004).
- **Bagremycins A and B:** produced by *Streptomyces* sp. Tü 4128, antibacterial and antifungal active (Bertasso *et al.*, 2001).
- **Decadienoic acid and decatrienoic acid:** produced by *Streptomyces viridochromogenes* Tü 6105, herbicidal active (Maier *et al.*, 1999).
- **Dioxolides A, B and D:** produced by *Streptomyces tendae* Tü 4042; no biological activity was found (Blum *et al.*, 1995).
- **Echinoserine:** a quinoxaline antibiotic produced by *Streptomyces tendae* Tü 4031, antibacterial active (Blum *et al.*, 1995).
- **Kanchanamycins A, C and D:** 36-membered polyene macrolide antibiotics produced by *Streptomyces olivaceus* Tü 4018 with antibacterial and antifungal activities (Fiedler *et al.*, 1996).
- **Naphthgeranine F:** a naphthoquinone antibiotic produced by *Streptomyces violaceus* Tü 3556 with antibacterial and antifungal activities (Hartjen *et al.*, 1995).
- **Nataxazole:** a benzoxazole antibiotic produced by *Streptomyces* sp. Tü 6176, antitumor active (Sommer *et al.*, 2008)
- **Phenalinolactones A–D:** terpenoglycoside antibiotics produced by *Streptomyces* sp. Tü 6071, active against Gram-positive bacteria (Gebhardt *et al.*, 2011).
- **Ripromycin:** a macrolactame antibiotic produced by *Streptomyces* sp. Tü 6239, antibacterial and antitumor active (Bertasso *et al.*, 2003).
- **Simocyclinones A–D:** angucyclinone antibiotics produced by *Streptomyces antibioticus* Tü 6040, antibacterial and antitumor active, novel potent inhibitors of bacterial gyrase (Schimana *et al.*, 2000, 2001; Holzenkämpfer *et al.*, 2002).
- **Spiridionic Acid:** produced by *Streptomyces* sp. Tü 6077, no biological activity was found (Textor *et al.*, 2007).
- **Spirofungin:** a polyketide-spiroketal antibiotic produced by *Streptomyces violaceus-niger* Tü 4113, antifungal active (Höltzel *et al.*, 1998).
- **Streptocidins A–D:** cyclic decapeptide antibiotics produced by *Streptomyces* sp. Tü 6071, antibacterial active (Gebhardt *et al.*, 2001; Höltzel *et al.*, 2001).

New secondary metabolites from *Micromonospora* strain Tü 6368

From a novel species of the genus *Micromonospora*, strain Tü 6368, the following new secondary metabolites were isolated and their structures elucidated (Antal *et al.*, 2005; Ströch *et al.*, 2005):

- **Galtamycin B:** an anthracycline antibiotic with antitumor activity;
- **Retymicin:** a xanthone antibiotic with antitumor activity;
- **Ribofuranosyl-lumichrome:** a new lumichrome derivative; no biological activity was found;
- **Saquayamycin Z:** an angucycline antibiotic, antibacterial and antitumor active.

New secondary metabolites from strains of the families *Streptosporangiaceae* and *Micromonosporaceae*

Within a scientific project funded by Novo Nordisk A/S, Denmark, specific attention was given to the actinomycete families *Streptosporangiaceae* and *Micromonosporaceae*. It was the beginning of a new area in the isolation and screening of less investigated actinomycete genera in my group with the aim to detect novel secondary metabolites. This work resulted in the discovery of:

- **(E)-4-Oxonon-2-enoic acid:** a fatty acid produced by *Streptomyces olivaceus* Tü 4018, antibacterial active (Pfefferle *et al.*, 1996).
- **1-Hydroxy-4-methoxy-2-naphthoic acid:** a naphthalene antibiotic produced by *Streptosporangium cinnabarinum* NNO 29536 and herbicidal active (Pfefferle *et al.*, 1997).
- **Kyanomycin:** an anthracycline antibiotic produced by a novel species of *Nonomuria*, strain NNO 22303. A blue-coloured compound with an unusual anthracycline-phosphatidylethanolamine hybrid structure exhibiting an antibacterial activity (Pfefferle *et al.*, 2000).
- **Tigloside:** a tigloylated tetrasaccharide produced by a novel species of *Amycolatopsis*, strain NNO 21702; no biological activity was found (Breinholt *et al.*, 1998).

New secondary metabolites from symbiotic microorganisms associated with arthropods

BMBF and BASF AG (Ludwigshafen, Germany) funded a 6-year research project to investigate symbiotic microorganisms from arthropods as a source of novel bioactive secondary metabolites. The results were not very promising, but led to the discovery of the following novel antibiotics:

- **Aspochalamins A-D and aspochalasin Z:** cytochalasan antibiotics produced by *Aspergillus niveus*, antibacterial and antitumor active (Gebhardt *et al.*, 2004, Höltzel *et al.*, 2004).

- **Endophenazines A–D:** phenazine antibiotics produced by various endosymbiotic isolates of *Streptomyces anulatus* with antibacterial, antifungal and herbicidal activities (Gebhardt *et al.*, 2002; Krastel *et al.*, 2002).

In addition, a multitude of partial novel natural products deriving from the primary metabolism were isolated and characterized, such as **3-hydroxyphenylacetic acid methylester**, **malonic acid phenylester**, **N-acetyl-phenylalanine** and **N-acetyltryptophan**.

Screening with actinomycetes from extreme terrestrial and marine habitats

A very stimulating and successful collaboration was started in 2000 together with the groups of Professor Michael Goodfellow from the University of Newcastle upon Tyne and Professor Alan T. Bull from the University of Kent (UK) that continued till my retirement in 2013. Both groups provided me with a huge series of dereplicated actinomycetes strains which were isolated from unique terrestrial and marine habitats including sediments of deep-sea trenches located in the Pacific and Atlantic Oceans. All strains were taxonomically characterised to the genus level that permitted an individual submerged cultivation in the screening stage, applying genus-adapted cultivation media. This collaboration resulted in the discovery of following metabolites:

- **Abyssomicins B, C, D, G and H, and atrop-Abyssomicin C:** polycyclic polyketide antibiotics with an unusual pharmacophore produced by the marine strain *Verrucosispora maris* AB-18-032 with antibacterial activity against Gram-positive bacteria including MRSA and vancomycin-resistant strains. Abyssomicin C and atrop-Abyssomicin C are novel inhibitors of *para*-aminobenzoic acid biosynthesis (Riedlinger *et al.*, 2004; Bister *et al.*, 2004; Keller *et al.*, 2007).
- **Atacamycins A–C:** 22-membered macrolactone antibiotics produced by *Streptomyces* sp. C38 which was isolated from a hyper-arid soil from the Atacama Desert, North Chile, showing an antitumor and phosphodiesterase inhibiting activity (Nachtigall *et al.*, 2011).
- **Bhimamycins F, H and I:** aromatic octaketide antibiotics produced by alkaliphilic strain *Streptomyces* sp. AK 671 with antibacterial and enzyme inhibitory activities (Jetter *et al.*, 2013).
- **Dermacozines A–L:** phenazine antibiotics produced by the deep-sea strains *Dermacoccus abyssi* MT1.1 and MT1.2, investigated in close collaboration with Professor Marcel Jaspars from the Aberdeen University. The novel phenazine-type compounds exhibit a free radical scavenging activity, antitumor and antiparasitic activities (Abdel-Mageed *et al.*, 2010).
- **Elaiomycins B and C:** alkylhydrazide antibiotics produced by *Streptomyces* sp. BK 190 with antibacterial, phosphodiesterase and acetylcholinesterase inhibiting activity (Kim *et al.*, 2011; Helaly *et al.*, 2011).

- **Genoketides A1 and A2, prechrysophanol glucuronide and chrysophanol glucuronide:** aromatic octaketides produced by alkaliphilic strain *Streptomyces* sp. AK 671, antitumor active (Fiedler *et al.*, 2008).
- **Lactonamycin Z:** produced by *Streptomyces sanglieri* AK 623, antibacterial and antitumor active (Höltzel *et al.*, 2003).
- **Proximicins A–C:** aminofuran antibiotics produced by the marine strain *Verrucosispora fiedleri* MG-37. Proximycin A was also isolated from the abyssomicin producer *Verrucosispora maris* AB-18-032. The metabolites show a weak antibacterial but a strong antitumor activity (Fiedler *et al.*, 2008; Keller *et al.*, 2008).
- **Pyrocoll:** a diketopiperazine antibiotic produced by alkaliphilic strain *Streptomyces* sp. AK 409, antibacterial, anti-parasitic and antitumor active (Dieter *et al.*, 2003).

NTK screening project with marine and extremophilic terrestrial actinomycetes

The promising results coming out of the collaboration with Professor Goodfellow and Professor Bull were the fundamentals that our consortium was funded by Boehringer Ingelheim Pharma (Biberach, Germany) in a 3-years scientific project, starting from 2002 till 2004. The goal of the collaboration was to screen extracts from unique marine and some extremophilic terrestrial members of the families *Streptomycetaceae*, *Micromonosporaceae* and *Pseudonocardiaceae*, and from selected members of mycolata as *Nocardia*, *Rhodococcus*, *Gordonia* and *Tsukamurella* for activities in various HTS assays. The first secondary metabolites coming out of this collaboration were:

- **Frigocyclinone:** an angucyclinone antibiotic produced by *Streptomyces griseus* NTK 97 from a soil sample collected at Antarctica, antibacterial active (Bruntner *et al.*, 2005).
- **Gephyromycin:** an angucyclinone antibiotic produced by *Streptomyces griseus* NTK 14, glutaminergic active towards neuronal cells (Bringmann *et al.*, 2005).

Unfortunately, the Boehringer-Ingelheim Company decided in 2004 to drop their activities in natural products completely, and therefore, the planned second phase of the proposed project in metabolite isolation and structure elucidation of the numerous hit strains was not possible. Only a few of these strains could be investigated later on in my group by undergraduate students, resulting in following new metabolites:

- **Albidopyrone:** a pyrone antibiotic produced by *Streptomyces* strain NTK 227, isolated from a sediment which was collected in the North-Atlantic Ocean. Albidopyrone showed an inhibitory activity against protein-tyrosin phosphokinase 1B (Hohmann *et al.*, 2009).

- **Benzoxacystol:** a 1,4-benzoxazine-type antibiotic produced by the marine strain *Streptomyces* sp. NTK 935, isolated from the same Canary Basin sediment as caboxamycin producer NTK 937. The metabolite has inhibitory activity against the enzymes glycogen synthase kinase 3 β and acetylcholinesterase (Nachtigall *et al.*, 2011)
- **Caboxamycin:** a benzoxazol antibiotic produced by *Streptomyces* strain NTK 937, isolated from a sediment which was collected in the Canary Basin at -3814 m. The metabolite showed antibacterial, antitumor and phosphodiesterase inhibitory activities (Hohmann *et al.*, 2009).

Project ACTAPHARM

Parallel to the NTK project sponsored by a pharmaceutical company, a further project was established with the aim to discover novel bioactive secondary metabolites from actinomycetes for clinical applications. Project ACTAPHARM was funded by the European Commission within the 5th framework program from 2001 till 2004, for a collaboration of the groups of Professor Liz Wellington (University of Warwick, UK), Professor Michael Goodfellow and Dr. Alan Ward (University of Newcastle, UK), Professor Lubbert Duijkuizen (University of Groningen, The Netherlands), PD Dr. Joachim Vater (Technische Universität Berlin, Germany), Professor Amalia Karagouni (University of Athens, Greece), Dr. Flavia Marinelli (Biosearch Italia, Gerenzano, Italy), and Dr. Fotinos (Lavipharm Group, Athens, Greece) and my own group. Until today, the following new secondary metabolites have been described:

- **Aurachins Q and R:** new members of the aurachin family produced by *Rhodococcus* sp. Acta 2259 with inhibitory activity against glycogen synthase kinase 3 β (Nachtigall *et al.*, 2010).
- **Bendigoles A–C:** new steroid metabolites produced by *Gordonia australis* Acta 2299 (Schneider *et al.*, 2008).
- **Elloxazinones A and B:** aminophenoxyazinone antibiotics produced by *Streptomyces* sp. Acta 2871 showing potent and selective antitumor activities (Graf *et al.*, 2006).
- **Fluostatins C, D and E:** fluorenone antibiotics produced by *Streptomyces* sp. Acta 1383 exhibiting an antitumor activity (fluostatin C) (Baur *et al.*, 2006; Schneider *et al.*, 2006).
- **Gombapyrones A–D:** α -pyrone antibiotics produced by *Streptomyces griseoruber* Acta 3662 with inhibitory activity against protein tyrosine phosphatase 1B and glycogen synthase kinase 3 β (Helaly *et al.*, 2009)
- **Grecocyclines A and B:** angucycline antibiotics from *Streptomyces* sp Acta 1362 with cytotoxic activity and inhibitory activity against protein tyrosine phosphatase 1B (Paululat *et al.*, 2010).

- **Grecoketides A and B:** glycosilated naphthoquinones produced by *Streptomyces* sp. Acta 1362 (Paululat *et al.*, 2008).
- **Nocardichelins A and B:** new type of siderophores produced by *Nocardia* sp. Acta 3026 with iron-chelating properties (Schneider *et al.*, 2006).
- **Langkocyclines A1, A2, A3, B1 and B2:** angucycline antibiotics produced by *Streptomyces* sp. Acta 3034 with antibacterial and antitumor activities (Kalyon *et al.*, 2013).
- **Langkolide:** a 32-membered macrolactone antibiotic produced by *Streptomyces* sp. Acta 3062 with antibacterial and antiproliferative activities (Helaly *et al.*, 2012).
- **Lipocarbazoles A1–A4:** carbazole-type metabolites produced by *Tsukamurella pseudospumae* Acta 1857 acting as free radical scavengers and showing antioxidative activity (Schneider *et al.*, 2009).
- **Phenelfamycins G and H:** antibiotics belonging to the elfamycin family related to phenelfamycins E and F, produced by *Streptomyces albospinus* Acta 3619 (Brötz *et al.*, 2011).
- **Skyllamycins A and B:** depsipeptide antibiotics produced by *Streptomyces* sp. Acta 2897. Skyllamycin A is identical with Antibiotic RP-1776, skyllamycin B is a new demethyl derivative of A. The compounds show an antibacterial activity (Pohle *et al.*, 2011).
- **Warkmycin:** an angucycline antibiotic produced by *Streptomyces* sp. Acta 2930 with antibacterial and antitumor activity (Helaly *et al.*, 2013).

Interactions of streptomycetes in the rhizosphere

A new research field was started in 2002 to investigate the interaction of soil streptomycetes with symbiotic fungi, pathogenic fungi and plants in the rhizosphere. The project was funded till 2010 by the Deutsche Forschungsgemeinschaft within the Graduate College ‘Infectious Biology’ at the University of Tübingen and was treated in close collaboration with the group of Professor Hampp, Institute of Physiological Ecology of Plants. The aim of our research was to characterize and identify the metabolites which act as signals in the communication between the different organisms. New metabolites coming out of this research were:

- **Aranciamycin Anhydride:** an anthracycline antibiotic from *Streptomyces* sp. Tü 6384 with antitumor activity (Nachtigall *et al.*, 2010).
- **Auxofuran:** a furan-type metabolite produced by the mycorrhiza helper bacterium *Streptomyces* sp. AcH 505 which act as a growth-stimulating factor for *Amanita muscaria* (fly acaric), and induces resistance against plant pathogenic fungi in plants (Riedlinger *et al.*, 2006).
- **Elaiomycins K and L:** azoxy-type antibiotics produced by *Streptomyces* sp. Tü 6399, exhibiting an antibacterial activity (Manderscheid *et al.*, 2013).

- **Fomannoxin Acids DFA, MFA-1 and MFA-2:** inactive conversion products of the fungal phytotoxin fomannoxin produced by *Heterobasidion* sp., converted by the rhizospheric strain *Streptomyces* sp. AcH 505.
- **Piceamycin:** a 23-membered macrolactam polyketide antibiotic produced by *Streptomyces* sp. GB 4-2 with inhibitory activity against Gram-positive bacteria, human tumor cell lines, and protein tyrosine phosphatase 1B (Schulz *et al.*, 2009).
- **Silvalactam:** a 24-membered macrolactam antibiotic produced by *Streptomyces* sp. Tü 6392 with antitumor activity (Schulz *et al.*, 2012).

National and international activities with strains and secondary metabolites discovered by the HP Fiedler Group

Some of the discovered new secondary metabolites which showed an unusual structure or structural elements were attractive for various groups involved in the genetics of the biosynthetic pathways. The following metabolites were investigated regarding their biosynthetic gene cluster and mode of action:

Abyssomicins: The characterization of the biosynthetic gene cluster in *Verrucosispora* sp. AB-18-032 is still in progress by the groups of Professor Roderich Süßmuth, Technische Universität Berlin, and Dr. JEM Stach, University of Newcastle, UK.

Gottardi, E.M.; J.M. Krawczyk, H. von Suchodoletz, S. Schadt, A. Mühlenweg, G.C. Uguru, S. Pelzer, H.-P. Fiedler, M.J. Bibb, J.E.M. Stach & R.D. Süßmuth: Abyssomicin biosynthesis: formation of an unusual polyketide, antibiotic-feeding studies and genetic analysis. *ChemBioChem* 12: 1401-1410, 2011.

Arylomycins: The biosynthetic gene cluster of these lipopeptide antibiotics produced by *Streptomyces* sp. Tü 6075 is under investigation in the group of Professor Stephanie Grond, Universität Tübingen. Studies for organic syntheses with biaryl-coupling enzymes are topic in the group of Dr. Tobias Gulder, Universität Bonn.

Caboxamycin and Nataxazole: The biosynthetic gene clusters of these carboxazole antibiotics produced by *Streptomyces* sp. NTK 937 and *Streptomyces* sp. Tü 6176, respectively, are investigated in the group of Professor José Salas, Universidad de Oviedo, Spain.

Endophenazines: The characterization of the biosynthetic gene cluster in *Streptomyces anulatus* 9958 with specific consideration of the origin of the prenyl side-chain was investigated in the group of Professor Lutz Heide, Universität Tübingen.

Saleh, O.; B. Gust, B. Boll, H.-P. Fiedler & L. Heide: Aromatic prenylation in the phenazine biosynthesis: dihydrophenazine-1-carboxylate dimethylallyltransferase from *Streptomyces anulatus*. *J. Biol. Chem.* 284: 14439-14447, 2009.

Frigocyclinone: The biosynthetic gene cluster of the angucyclinone antibiotic frigocyclinone produced by *Streptomyces griseus* NTK 97 is under investigation in the group of Professor Huizhan Zhang, East China University of Science and Technology, Shanghai, P.R. China.

Genoketides A1and A2: The biosynthetic gene cluster of chrysophanol glucuronide and its biosynthetic intermediates genoketide A1 and A2 and pre-chrysophanol glucuronide is under investigation in the group of Professor Tom Simpson, University of Bristol, UK.

Grecocyclines and Grecoketides: The biosynthetic gene cluster of both novel angucycline and naphthoquinone antibiotics, respectively, is under investigation by Dr. Andriy Luzhetskyy, Universität des Saarlandes.

Lactonamycin Z: The characterization of the biosynthetic gene cluster in *Streptomyces sanglieri* AK 623 was described by the group of Professor Ron Parry, Rice University, Houston, Texas, USA.

Zhang, X.; L.B. Alemany, H.-P. Fiedler, M. Goodfellow & R.P. Parry: Biosynthetic investigations of lactonamycin and lactonamycin Z: cloning of the biosynthetic gene clusters and discovery of an unusual starter unit. *Antimicrob. Agents Chemother.* 52: 574-585, 2008.

Kanchanamycins: The producing strain *Streptomyces olivaceus* Tü 4018 is used as a host for heterologous expression experiments in the group of Dr. Andriy Luzhetskyy, Universität des Saarlandes.

Phenalinolactones: The biosynthetic gene cluster in *Streptomyces* sp. Tü 6071 was described by the group of Professor Andreas Bechthold, Universität Freiburg. The strain is under investigation in the group of Professor Pieter Dorrestein, University of California, San Diego, USA.

Dürr, C.; H.-J. Schnell, A. Luzhetskyy, R. Murillo, M. Weber, K. Welzel, A. Vente & A. Bechthold: Biosynthesis of terpene phenalinolactone in *Streptomyces* sp. Tü6071: analysis of the gene cluster and generation of derivatives. *Chem. Biol.* 13: 365-377, 2006.

Polyketomycin: The characterization of the biosynthetic gene cluster in *Streptomyces diastatochromogenes* Tü 6028 was done in the group of Professor Andreas Bechthold, Universität Freiburg.

Daum, M.; I. Peintner, A. Frerich, M. Weber, T. Paululat & A. Bechthold: Organisation of the biosynthetic gene cluster and tailoring enzymes in the biosynthesis of the tetracyclic quinone glycoside antibiotic polyketomycin. *ChemBioChem* 10: 1073-1083, 2009.

Saquayamycin Z: The characterization of the biosynthetic gene cluster in *Micro-monospora* sp. Tü 6368 is still in work by the group of Professor Andreas Bechthold, University of Freiburg, and Professor Rongson Pongdee, University of Sewanee, Sewanee, TN, USA.

Erb, A.; A. Luzhetskyy, U. Hardter & A. Bechthold: Cloning and sequencing of biosynthetic gene cluster for saquayamycin Z and galtamycin B and the elucidation of the assembly of their saccharide chains. *ChemBioChem* 10: 1392-1401, 2009.

Simocyclinones: The complete biosynthetic gene cluster in *Streptomyces antibioticus* Tü 6040 was described by the groups of Professor Lutz Heide, Universität Tübingen, and Professor Andreas Bechthold, Universität Freiburg.

Galm, U.; J. Schimana, H.-P. Fiedler, J. Schmidt, S.-M. Li & L. Heide: Cloning and analysis of the simocyclinone biosynthetic gene cluster of *Streptomyces antibioticus* Tü 6040. *Arch. Microbiol.* 178: 102-114, 2002.

Trefzer, A.; S. Pelzer, J. Schimana, S. Stockert, C. Bihlmaier, H.-P. Fiedler, K. Welzel, A. Vente & A. Bechthold: Biosynthetic gene cluster of simocyclinone, a natural multihybrid antibiotic. *Antimicrob. Agents Chemother.* 46: 1174-1182, 2002.

The mode of action of simocyclinones is extensively studied by the groups of Professor Tony Maxwell and Professor Mark J. Buttner (John Innes Centre, Norwich, UK) and Dr. Keith C. Ellis (University of Kansas, USA).

Flatman, R.H.; A.J. Howells, L. Heide, H.-P. Fiedler & A. Maxwell: Simocyclinone D8: an inhibitor of DNA gyrase with a novel mode of action. *Antimicrob. Agents Chemother.* 49: 1093-1100, 2005.

Le, T.B.K.; H.-P. Fiedler, C.D. den Hengst, S.K. Ahn, A. Maxwell & M.J. Buttner: Coupling of the biosynthesis and export of the DNA gyrase inhibitor simocyclinone in *Streptomyces antibioticus*. *Molec. Microbiol.* 72: 1462-1474, 2009.

Oppegard, L.M.; B.L. Hamann, K.R. Streck, K.C. Ellis, H.-P. Fiedler, A.B. Khodursky & H. Hiasa: *In vivo* and *in vitro* patterns of the activity of simocyclinone D8, an angucyclinone antibiotic from *Streptomyces antibioticus*. *Antimicrob. Agents Chemother.* 53:2110-2119, 2009.

Edwards, M.J.; R.H. Flatman, L.A. Mitchenall, C.E.M. Stevenson, T.B.K. Le, T.A. Clarke, A.R. McKay, H.-P. Fiedler, M.J. Buttner, D.M. Lawson & A. Maxwell: A crystal structure of the bifunctional antibiotic, simocyclinone D8, bound to DNA gyrase. *Science* 326: 1415-1417, 2009.

Le, T.B.K.; C.E.M. Stevenson, H.-P. Fiedler, A. Maxwell, D.M. Lawson & M.J. Buttner: Structures of the TetR-like simocyclinone efflux pump repressor, SimR, and the mechanism of ligand-mediated derepression. *J. Molec. Biol.* 408: 40-56, 2011.

The simocyclinone producing strain *Streptomyces antibioticus* Tü 6040 is investigated in the group of Professor Justin Nodwell, McMaster University, Hamilton, Canada, regarding TetR-like proteins.

Skyllamycin: The biosynthetic gene cluster of the NRPS cyclodepsipeptide is investigated in the group of Professor Roderich Süssmuth, Technische Universität Berlin.

Pohle, S.; C. Appelt, M. Roux, H.-P. Fiedler & R.D. Süssmuth: Biosynthetic gene cluster of the non-ribosomally synthesized cyclodepsipeptide skyllamycin: deciphering unprecedented ways of unusual hydroxylation reactions. *J. Am. Chem. Soc.* 133: 6194-6205, 2011.

Spirodionic Acid: The characterization of the biosynthetic gene cluster in *Streptomyces* sp. Tü 6077 is under investigation in the group of Professor Stephanie Grond, Universität Tübingen.

Spirofungin: Investigations on the organization of PKS genes of *Streptomyces violaceusniger* Tü 4113 are in progress in the group of Dr. Leonard Katz, SynBERC, University of California, Emeryville, USA.

The spirofungin producing strain *Streptomyces violaceusniger* Tü 4113 is investigared regarding volatiles in the group of Dr. Jeroen Dickschat, TU Braunschweig.

Some of our secondary metabolites with interesting and unusual structural elements were subject for chemical total synthesis:

Abyssomicins

- Rath, J.-P.; S. Kinast & M.E. Maier: Synthesis of the fully functionalized core structure of the antibiotic abyssomicin C. *Org. Lett.* 7: 3089-3092, 2005.
- Zografos, A.L.; A. Yiotakis & D. Georgiadis: Rapid access to the tricyclic spirotetronic core of abyssomicins. *Org. Lett.* 7: 4515-4518, 2005
- Snider, B.B. & Y. Zou: Synthesis of the carbocyclic skeletonof abyssomicins C and D. *Org. Lett.* 7, 4939-4941, 2005
- Zapf, C.W.; B.A. Harrison, C. Drahla & E.J. Sorensen: A Diels-Alder macrocyclization enables an efficient asymmetric synthesis of the antibacterial natural product abyssomicin C. *Angew. Chem. Int. Ed.* 44: 6533-6537, 2005.
- Nicolaou, K.C. & S.T. Harrison: Total synthesis of abyssomicin and atrop-abysomicin C. *Angew. Chem. Int. Ed.* 45: 3256-3260, 2006.
- Couladouros, E.A.; E.A. Bouzas & A.D. Magos: Formal synthesis of abyssomicin C. *Tetrahedron* 62: 5272-5279, 2006.

Arylomycins

- Roberts, T.C.; P.A. Smith, R.T. Cirz & F.E. Romersberg: Structural and initial biological analysis of synthetic arylomycin A2. *J. Am. Chem. Soc.* 129: 15830-15838, 2007.

Fluostatin

- Yu, M. & S.J. Danishefsky: A direct route to fluostatin C by a fascinating Diels-Alder reaction. *J. Am. Chem. Soc.* 130: 2783-2785, 2008.

Lactonamycin Z

Adachi, S.; K. Watanabe, Y. Iwata, S. Kameda, Y. Miyaoka, M. Onozuka, R. Mitusi, Y. Saikawa & M. Nakata: Total synthesis of lactonamycin and lactonamycin Z with late-stage A-ring formation and glycosylation. *Angew. Chem. Int. Ed.* 52: 2087-2091, 2013.

Spirofungin

Shimizu, Y.; H. Kiyota & T. Oritani: Synthesis of the spiroacetal parts of spirofungin A and B. *Tetrahedr. Lett.* 41: 3141-3144, 2000.

Zanatta, S.D.; J.M. White & M.A. Rizzacasa: Total synthesis of the proposed structure for spirofungin B: a reassignment of the stereochemistry. *Org. Lett.* 6: 1041-1044, 2004.

Shimizu, T.; T. Satoh, K. Murakoshi & M. Sodeoka: Asymmetric total synthesis of (-)-spirofungin A and (+)-spirofungin B. *Org. Lett.* 7: 5573-5576, 2005.

Marjanovic, J. & S.A. Kozmin: Spirofungin A; stereoselective synthesis and inhibition of isoleucyl-tRNA synthetase. *Angew. Chem. Int. Ed.* 46: 8854-8857, 2007.

Crimmins, M.T. & E.A. O'Bryan: Enantioselective total synthesis of spirofungins A and B. *Org. Lett.* 12: 4416-4419, 2010.

Neumaier, J. & M.E. Maier: Synthesis of the spirofungin A core via a domino strategy consisting of olefinic ester ring-closing metathesis and iodospiroacetalization. *Synlett* 2011, 2, 187-190.

II. Isolated Metabolites

Abyssomicins B, C, atrop C, D, G and H

Producing organism: *Verrucospora maris* AB-18-032



Habitat:

marine; Sea of Japan, sediment –289 m

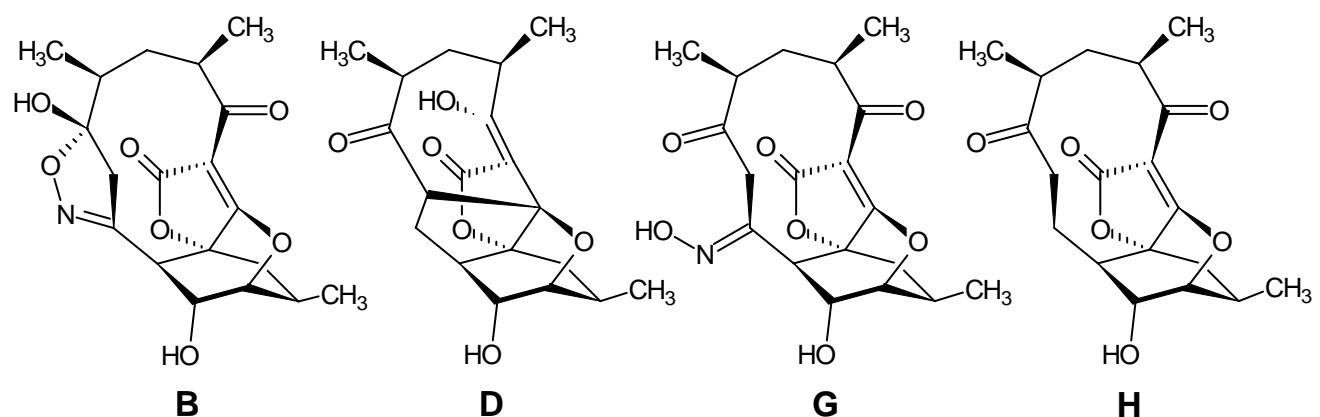
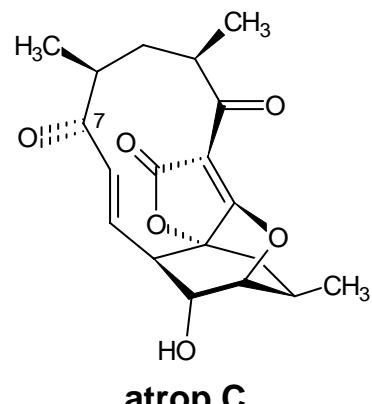
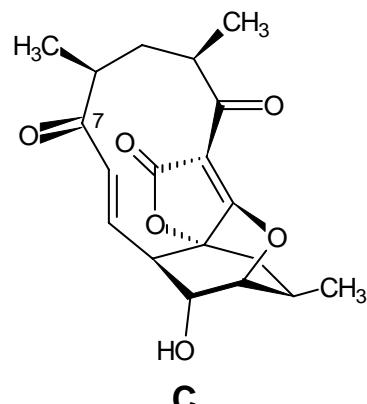
Screening method:

inhibition of *para*-aminobenzoic acid biosynthesis

Biological activity:

antibacterial, inhibitor of *para*-aminobenzoic acid and folic acid biosynthesis (abyssomicin C, atrop-abyssomicin C)

Structures:



Abyssomicins B, C, atrop C, D, G and H

References:

Riedlinger, J.; A. Reicke, H. Zähner, B. Krismer, A.T. Bull, L.A. Maldonado, A.C. Ward, M. Goodfellow, B. Bister, D. Bischoff, R. Süssmuth & H.-P. Fiedler. *J. Antibiotics* 57: 271-279, 2004

Bister, B.; D. Bischoff, M. Ströbele, J. Riedlinger, A. Reicke, F. Wolter, A.T. Bull, H. Zähner, H.-P. Fiedler & R.D. Süssmuth. *Angew. Chem. Int. Ed.* 43: 2574-2576, 2004

Keller, S.; G. Nicholson, C. Drahl, E. Sorensen, H.-P. Fiedler & R.D. Süssmuth. *J. Antibiotics* 60: 391-394, 2007

Gottardi, E.M.; J.M. Krawczyk, H. von Suchodoletz, S. Schadt, A. Mühlenweg, G.C. Uguru, S. Pelzer, H.-P. Fiedler, M.J. Bibb, J.E.M. Stach & R.D. Süssmuth: *ChemBioChem* 12: 1401-1410, 2011

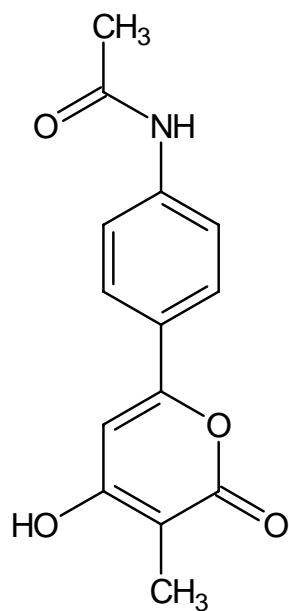
Goodfellow, M.; J.E.M. Stach, R. Brown, A.N.V. Bonda, A.L. Jones, J. Mexson, H.-P. Fiedler, T.D. Zucchi & A.T. Bull. *Antonie van Leeuwenhoeck* 101: 185-193, 2012.

Albidopyrone

Producing organism: *Streptomyces* sp. NTK 227



Habitat: marine; sediment –158 m, North Atlantic Ocean
Screening method: HPLC-DAD
Biological activity: inhibitor of protein-tyrosin phosphokinase 1B
Structure:



Reference:

Hohmann, C.; K. Schneider, C. Bruntner, R. Brown, A.L. Jones, M. Goodfellow, M. Krämer, J.F. Imhoff, G. Nicholson, H.-P. Fiedler & R.D. Süssmuth. *J. Antibiotics* 62: 75-79, 2009

Aranciamycin Anhydride

Producing organism: *Streptomyces* sp. Tü 6384

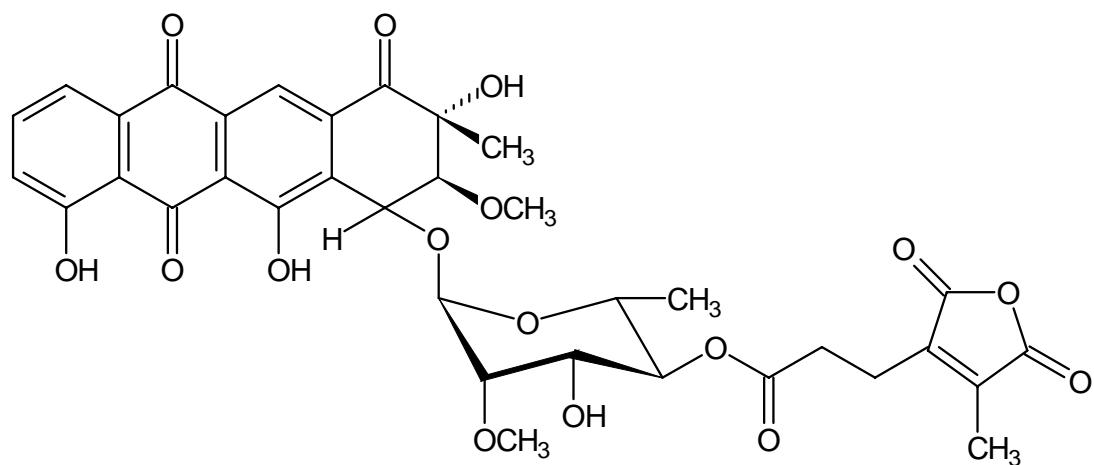


Habitat: terrestrial; rhizosphere of Norway spruce, Rammert Forest, Tübingen, Germany

Screening method: HPLC-DAD

Biological activity: antitumor

Structure:

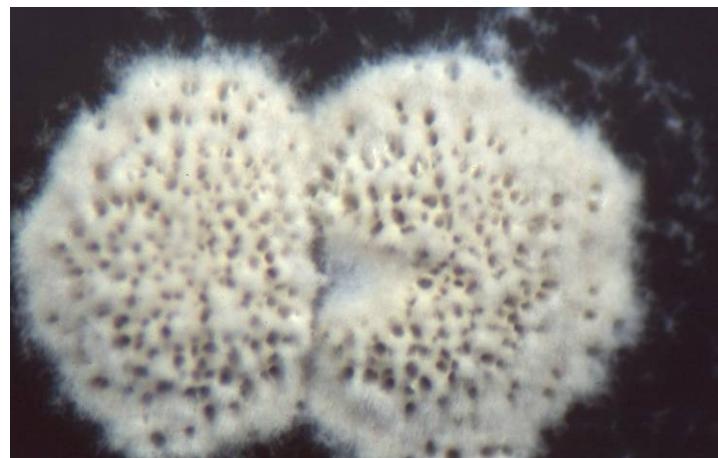


Reference:

Nachtigall, J.; D. Schulz, W. Beil, R.D. Süßmuth & H.-P. Fiedler. J. Antibiotics 63: 397-399, 2010

Arylomycins A and B

Producing organism: *Streptomyces* sp. Tü 6075



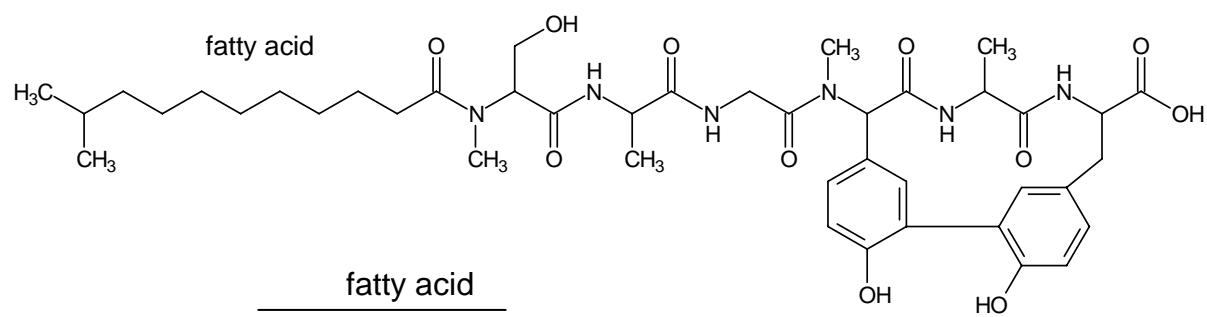
Habitat: terrestrial; Cape Coast, Ghana

Screening method: HPLC-DAD

Biological activity: antibacterial, inhibitors of bacterial signal peptidase

Structures:

Arylomycin A series



A1: *iso-C₁₁*

A2: *iso-C₁₂*

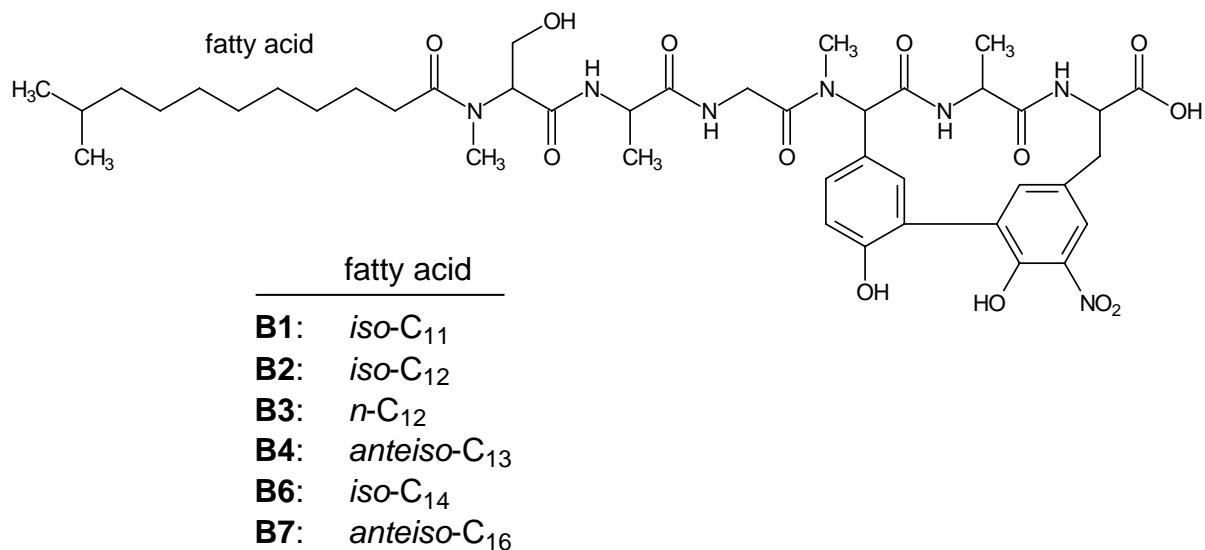
A3: *n-C₁₂*

A4: *anteiso-C₁₃*

A6: *iso-C₁₄*

Arylomycins A and B

Arylomycin B series

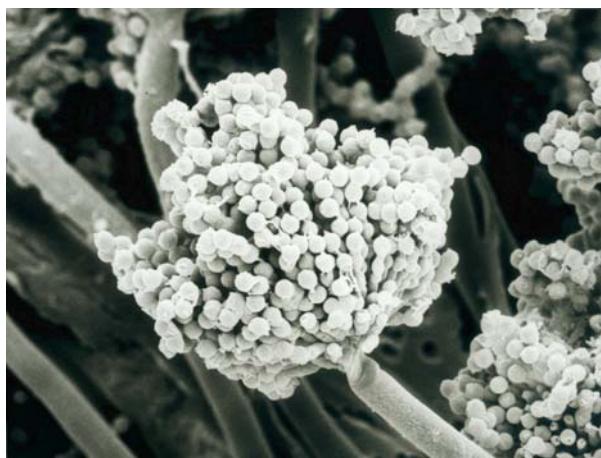


References:

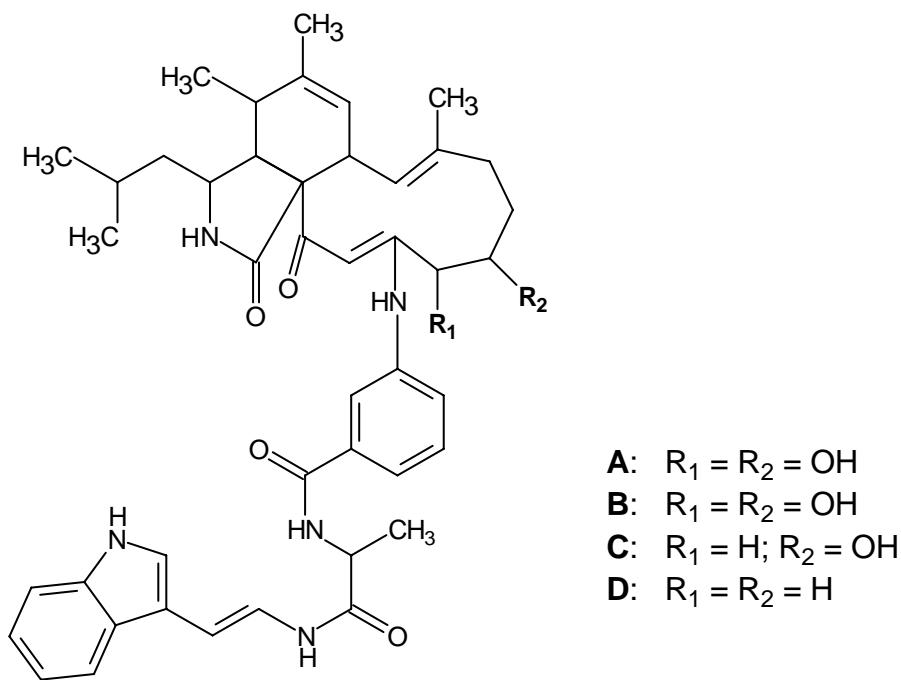
- Schimana, J.; K. Gebhardt, J. Müller, A. Höltzel, D.G. Schmid, R. Süssmuth, R. Pukall & H.-P. Fiedler. *J. Antibiotics* 55: 565-570, 2002
- Höltzel, A.; D.G. Schmid, G.J. Nicholson, S. Stevanovic, J. Schimana, K. Gebhardt, H.-P. Fiedler & G. Jung. *J. Antibiotics* 55: 571-577, 2002

Aspochalamins A–D

Producing organism: *Aspergillus niveus* LU 9575



Habitat: endosymbiotic; gut from a *Trichoniscidae*
Screening method: HPLC-DAD
Biological activity: antitumor, antibacterial
Structures:

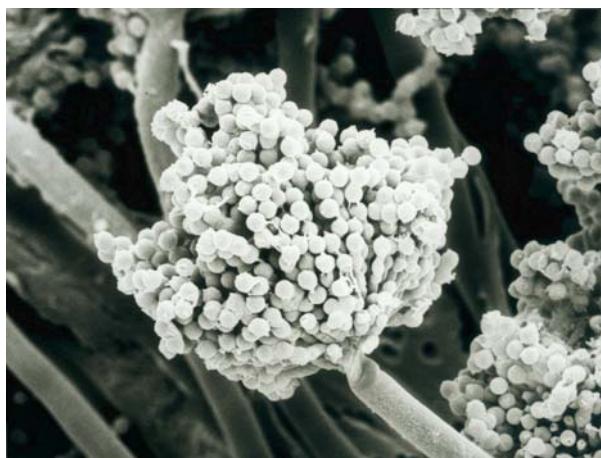


References:

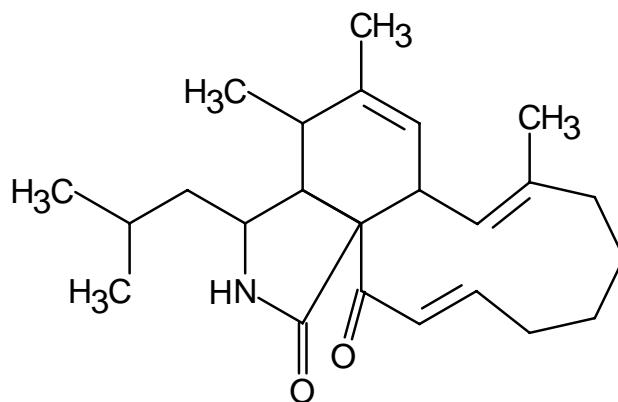
- Gebhardt, K.; J. Schimana, A. Höltzel, K. Dettner, S. Draeger, W. Beil, J. Rheinheimer & H.-P. Fiedler. *J. Antibiotics* 57: 707-714, 2004
- Höltzel, A.; D.G. Schmid, G.J. Nicholson, P. Krastel, A. Zeeck, K. Gebhardt, H.-P. Fiedler & G. Jung. *J. Antibiotics* 57: 715-720, 2004

Aspochalasin Z

Producing organism: *Aspergillus niveus* LU 9575



Habitat: endosymbiotic; gut from a *Trichoniscidae*
Screening method: HPLC-DAD
Biological activity: antitumor
Structure:



References:

- Gebhardt, K.; J. Schimana, A. Höltzel, K. Dettner, S. Draeger, W. Beil, J. Rheinheimer & H.-P. Fiedler. *J. Antibiotics* 57: 707-714, 2004
- Höltzel, A.; D.G. Schmid, G.J. Nicholson, P. Krastel, A. Zeeck, K. Gebhardt, H.-P. Fiedler & G. Jung. *J. Antibiotics* 57: 715-720, 2004

Atacamycins A–C

Producing organism: *Streptomyces* sp. C38

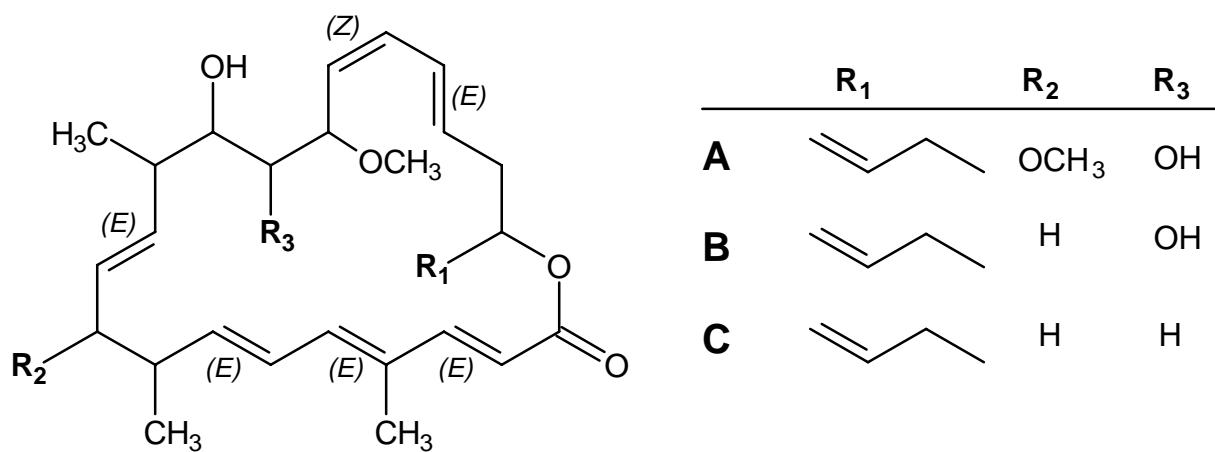


Habitat: terrestrial; hyper-arid soil from the Atacama Desert, Salar de Atacama, Chile

Screening method: HPLC-DAD

Biological activity: antitumor, inhibitors of phosphodiesterase

Structures:



Reference:

Nachtigall, J.; A. Kulik, S. Helaly, A.T. Bull, M. Goodfellow, J.A. Asenjo, A. Maier, J. Wiese, J.F. Imhoff, R.D. Süßmuth & H.-P. Fiedler. *J. Antibiotics* 64: 775-780, 2011

Aurachines Q and R

Producing organism: *Rhodococcus* sp. Acta 2259

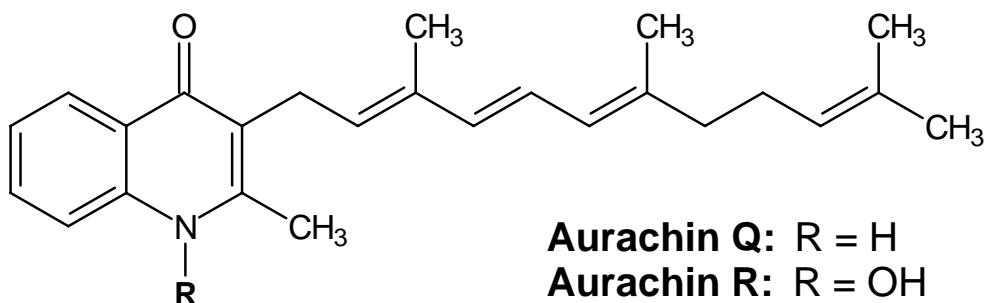


Habitat: activated sludge; Milcote Pilot Sewage Treatment Plant, Stratford-upon-Avon, UK

Screening method: HPLC-DAD

Biological activity: inhibitor of glycogen synthase kinase 3-beta

Structures:

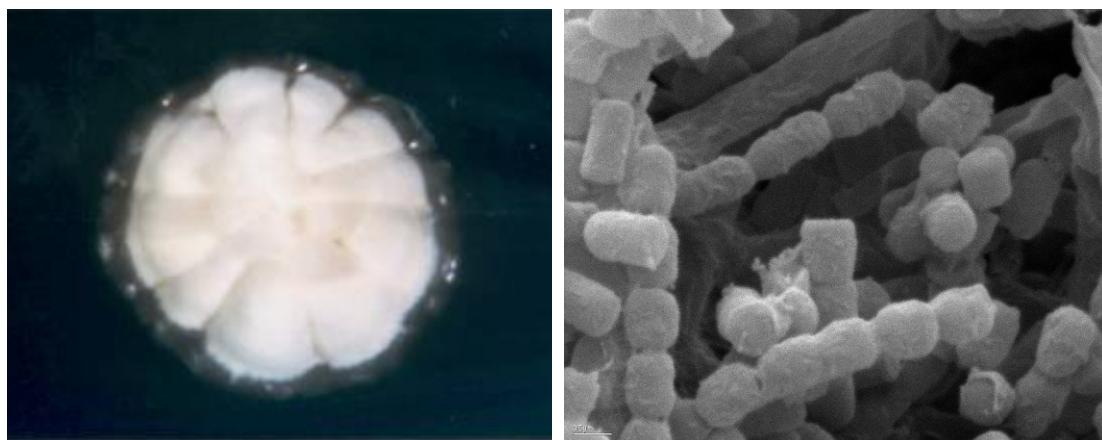


Reference:

Nachtigall, J.; K. Schneider, G. Nicholson, M. Goodfellow, H. Zinecker, J.F. Imhoff, R.D. Süßmuth & H.-P. Fiedler. *J. Antibiotics* 63: 567-569, 2010

Auxofuran

Producing organism: *Streptomyces* sp. AcH 505

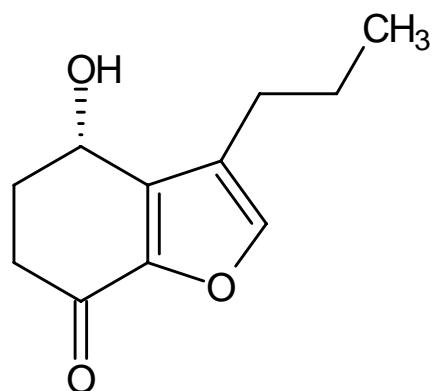


Habitat: terrestrial; rhizosphere of Norway spruce, Haigerloch, Germany

Screening method: HPLC-DAD

Biological activity: growth stimulating factor for symbiotic fungi

Structure:

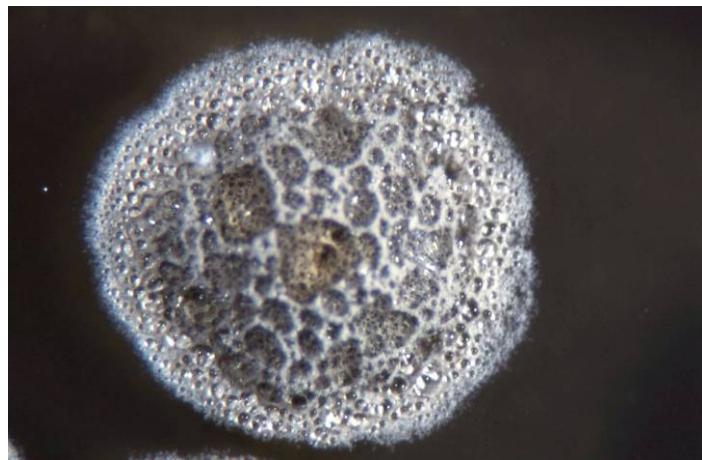


Reference:

Riedlinger, J.; S.D. Schrey, M.T. Tarkka, R. Hampp, M. Kapur & H.-P. Fiedler. Appl. Environ. Microbiol. 72: 3550-3557, 2006

Bagremycins A and B

Producing organism: *Streptomyces* sp. Tü 4128

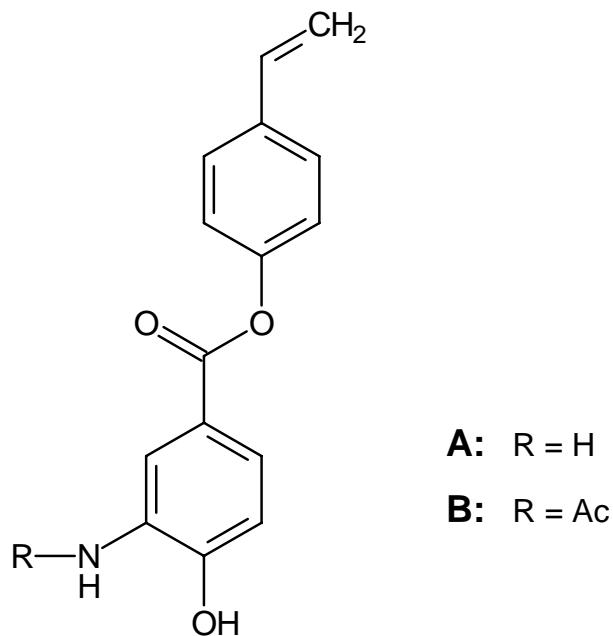


Habitat: terrestrial; Java, Indonesia

Screening method: HPLC-DAD

Biological activity: antibacterial, antifungal

Structures:



Reference:

Bertasso, M.; M. Holzenkämpfer, A. Zeeck, F. Dall'Antonia & H.-P. Fiedler. J. Antibiotics 54: 730-736, 2001

Bendigoles A–C

Producing organism: *Gordonia australis* Acta 2299

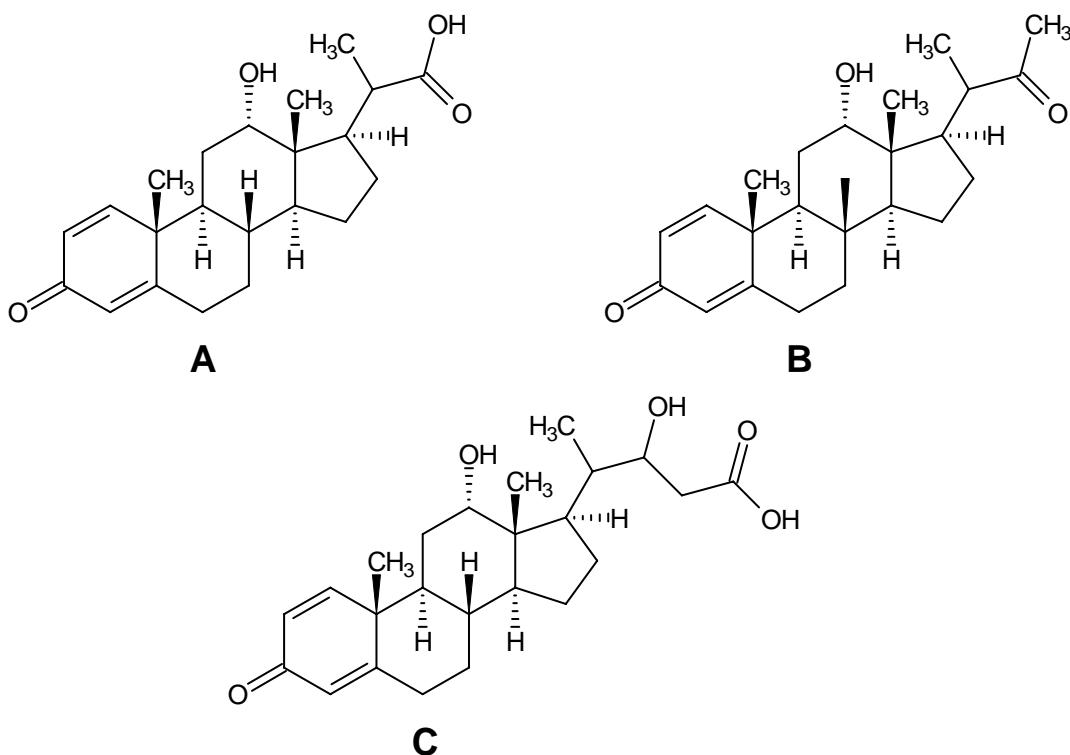


Habitat: activated sludge; Bendigo, Australia

Screening method: HPLC-DAD

Biological activity: binding activity to human progesterone and androgen receptor

Structures:



Reference:

Schneider, K.; E. Graf, E. Irran, G. Nicholson, F.M. Stainsby, M. Goodfellow, S.A. Borden, S. Keller, R.D. Süßmuth & H.-P. Fiedler. J. Antibiotics 61: 356-364, 2008

Benzoxacystol

Producing organism: *Streptomyces* sp. NTK 935

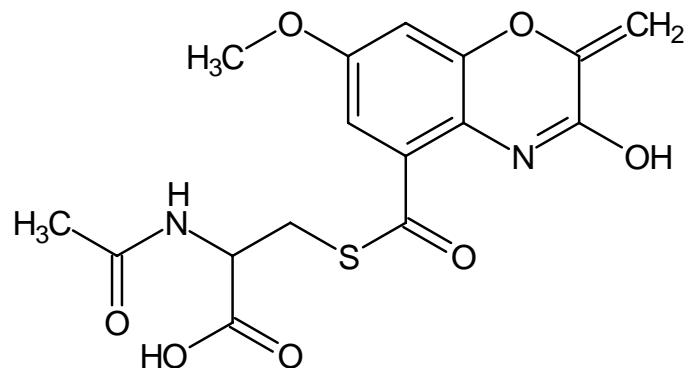


Habitat: marine; sediment –3814 m, Canary Basin, Atlantic Ocean

Screening method: HPLC-DAD

Biological activity: inhibitor of glycogen synthase kinase 3 β and acetylcholin esterase

Structure:



Reference:

Nachtigall, J.; K. Schneider, C. Bruntner, A.T. Bull, M. Goodfellow, H. Zinecker, J.F. Imhoff, G. Nicholson, E. Irran, R.D. Süßmuth & H.-P. Fiedler. *J. Antibiotics* 64: 453-457, 2011

Bhimamycins F, H and I

Producing organism: *Streptomyces* sp. AK 671

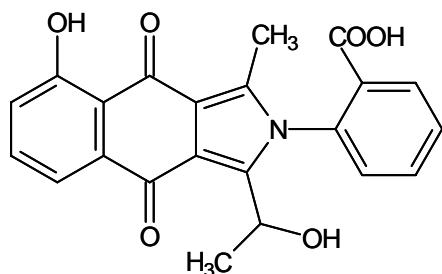


Habitat: terrestrial; Hamsterley Forest, Northumberland, UK

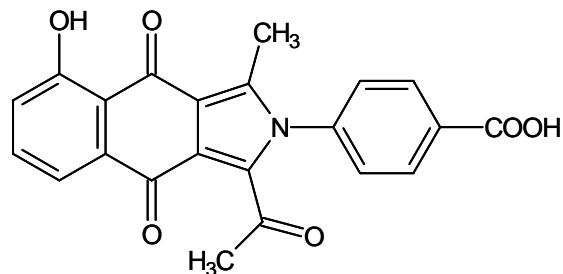
Screening method: HPLC-DAD

Biological activity: antibacterial (H, I), inhibitors of phosphodiesterase 4 (H, I) and glycogen synthase kinase-3 β (H)

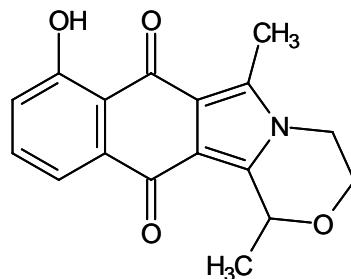
Structures:



Bhimamycin F



Bhimamycin H



Bhimamycin I

Reference:

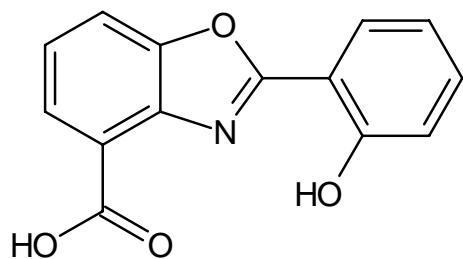
Jetter, P.; C. Steinert, M. Knauer, G. Zhang, T. Bruhn, J. Wiese, J.F. Imhoff, H.-P. Fiedler & G. Bringmann. *J. Antibiotics* in press, 2013

Caboxamycin

Producing organism: *Streptomyces* sp. NTK 937



Habitat: marine; sediment –3814 m, Canary Basin, Atlantic Ocean
Screening method: HPLC-DAD
Biological activity: antibacterial, antitumor, phosphodiesterase inhibitor
Structure:

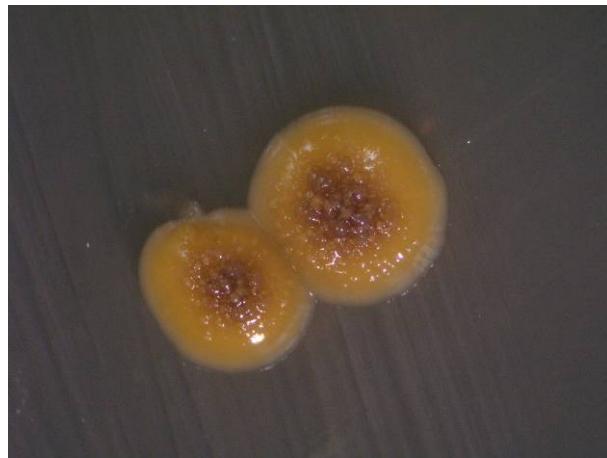


Reference:

Hohmann, C.; K. Schneider, C. Bruntner, E. Irran, G. Nicholson, A.T. Bull, A.L. Jones, R. Brown, J.E.M. Stach, M. Goodfellow, W. Beil, M. Krämer, J.F. Imhoff, R.D. Süßmuth & H.-P. Fiedler. *J. Antibiotics* 62: 99-104, 2009

Dermacozines A–L

Producing organism: *Dermacoccus abyssi* MT1.1, *Dermacoccus* sp. MT1.2

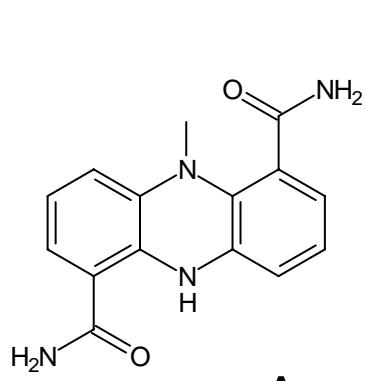


Habitat: marine; sediment –11,898 m, Challenger Deep, Mariana Trench, Pacific Ocean

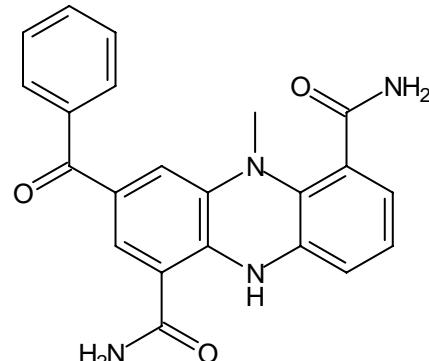
Screening method: HPLC-DAD

Biological activity: antioxidant

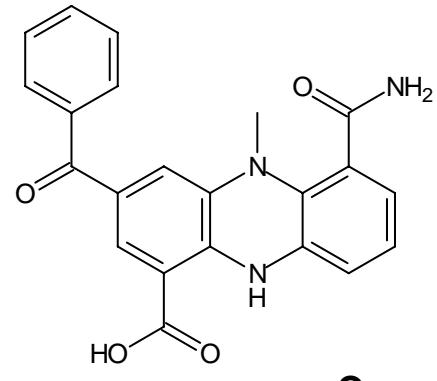
Structures:



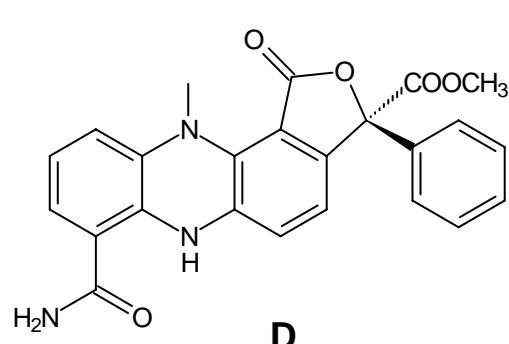
A



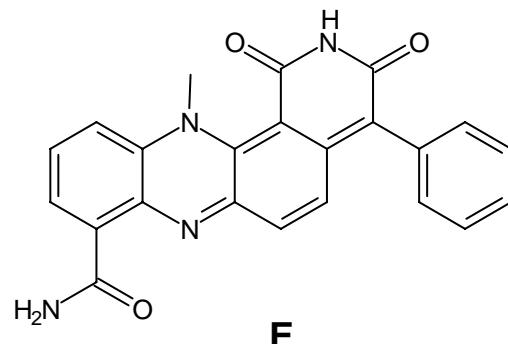
B



C

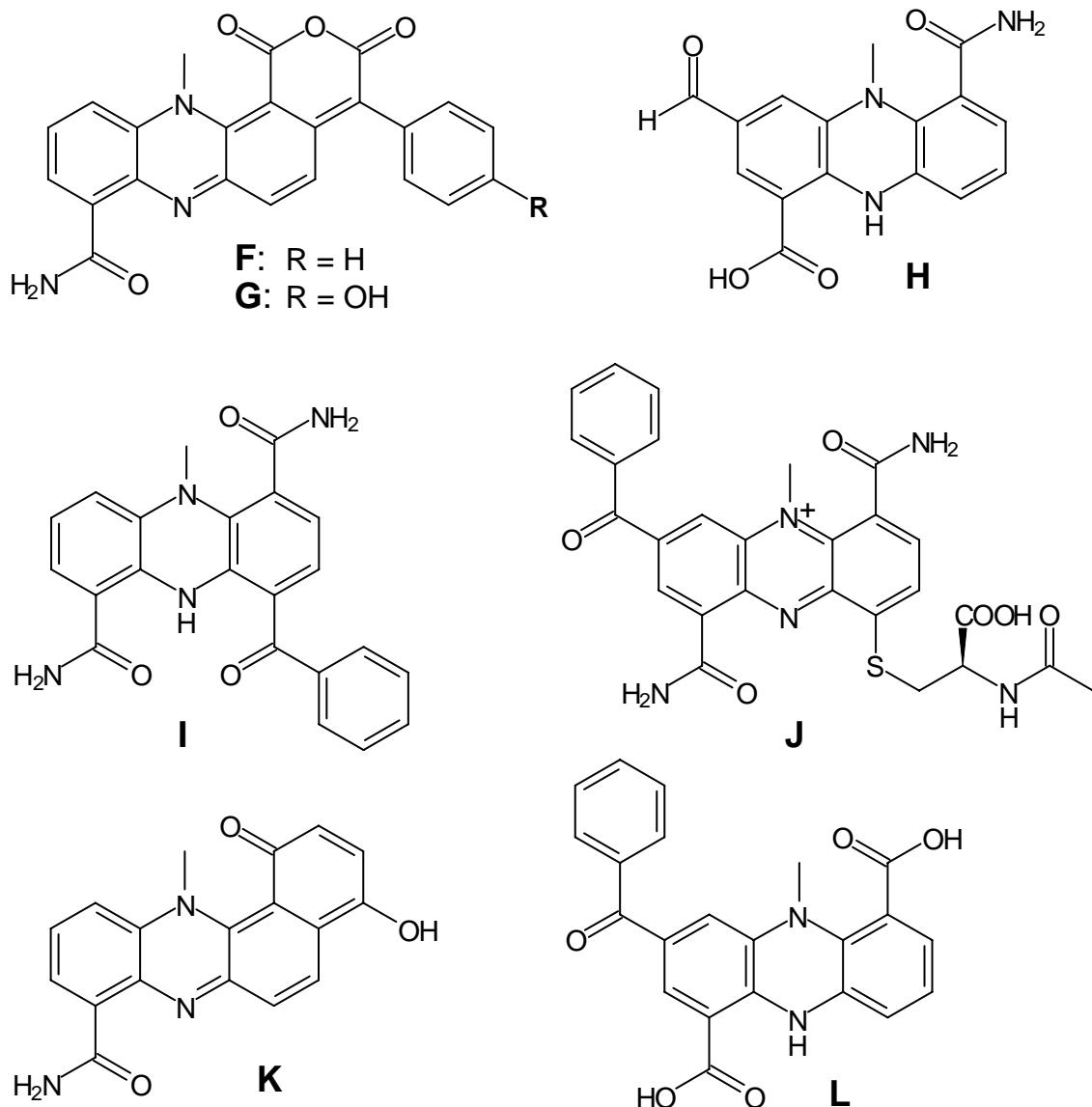


D



E

Dermacozines A–L

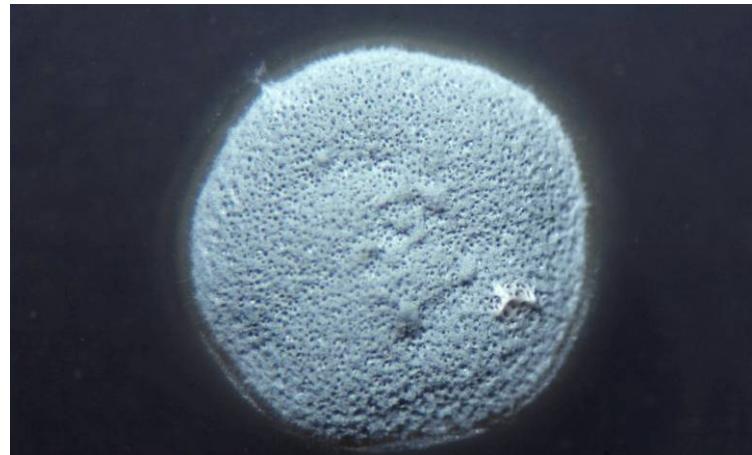


Reference:

Abdel-Mageed, W.M.; B.F. Milne, M. Wagner, M. Schumacher, P. Sandor, W. Pathom-aree, M. Goodfellow, A.T. Bull, K. Horikoshi, R. Ebel, M. Diedrich, H.-P. Fiedler & M. Jaspars. Org. Biomol. Chem. 8: 2352-2362, 2010

(2E,4Z)-Decadienoic Acid (2E,4Z,7Z)-Decatrienoic Acid

Producing organism: *Streptomyces viridochromogenes* Tü 6105

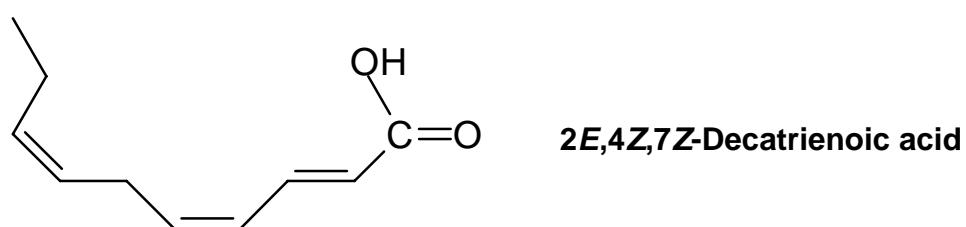
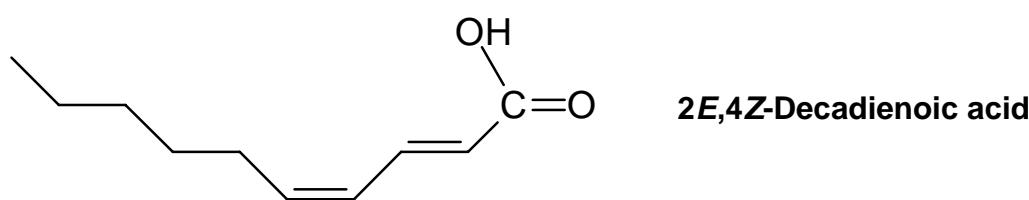


Habitat: terrestrial; Cape Coast, Ghana

Screening method: HPLC-DAD

Biological activity: herbicidal

Structures:



Reference:

Maier, A.; J. Müller, P. Schneider, H.-P. Fiedler, I. Groth, F.S.K. Tayman, F. Teltschik, C. Günther & G. Bringmann. Pestic. Science 55: 733-739, 1999

Dioxolides

Producing organism: *Streptomyces tendae* Tü 4042

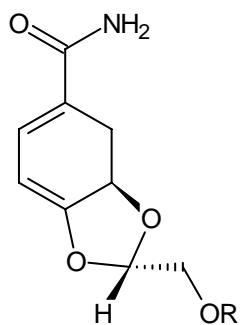


Habitat: terrestrial; Alice Springs, Australia

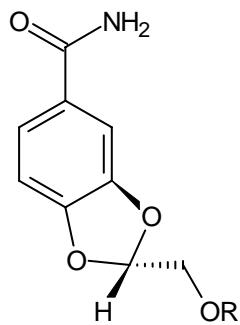
Screening method: HPLC-DAD

Biological activity: none

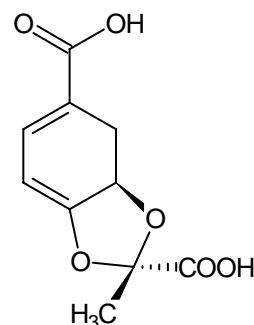
Structures:



Dioxolide A R = H
Dioxolide B R = COCH₃



Dehydrodioxolide A R = H
Dehydrodioxolide B R = COCH₃



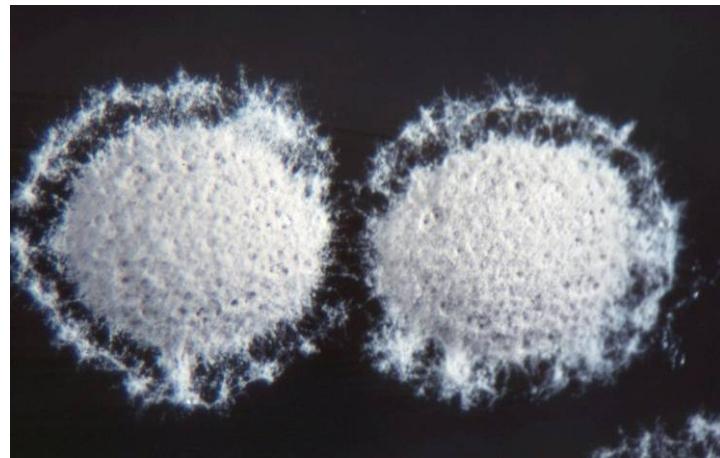
Dioxolide D

Reference:

Blum, S.; I. Groth, J. Rohr & H.-P. Fiedler. J. Basic Microbiol. 36: 19-25, 1996

Echinosserine

Producing organism: *Streptomyces tendae* Tü 4031

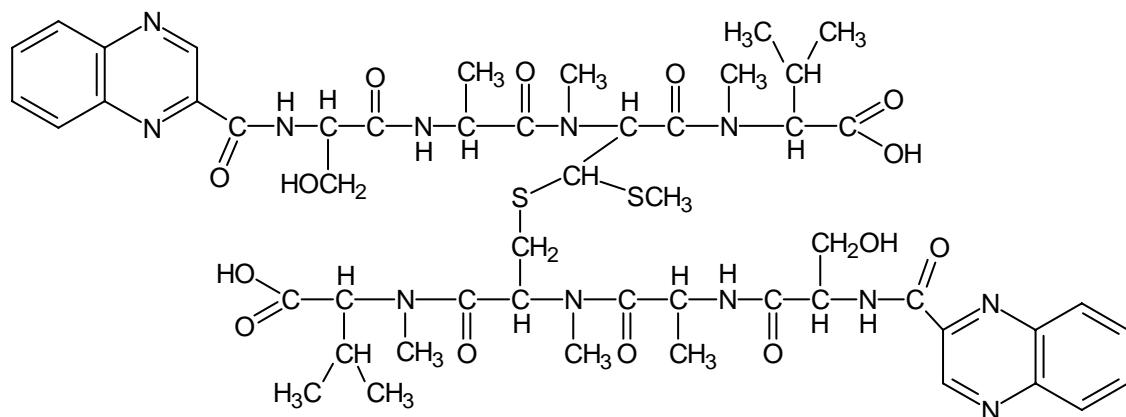


Habitat: terrestrial; Alice Springs, Australia

Screening method: HPLC-DAD

Biological activity: antibacterial

Structure:



Reference:

Blum, S.; H.-P. Fiedler, I. Groth, C. Kempfer, H. Stephan, G. Nicholson, J.W. Metzger & G. Jung. *J. Antibiotics* 48: 619-625, 1995

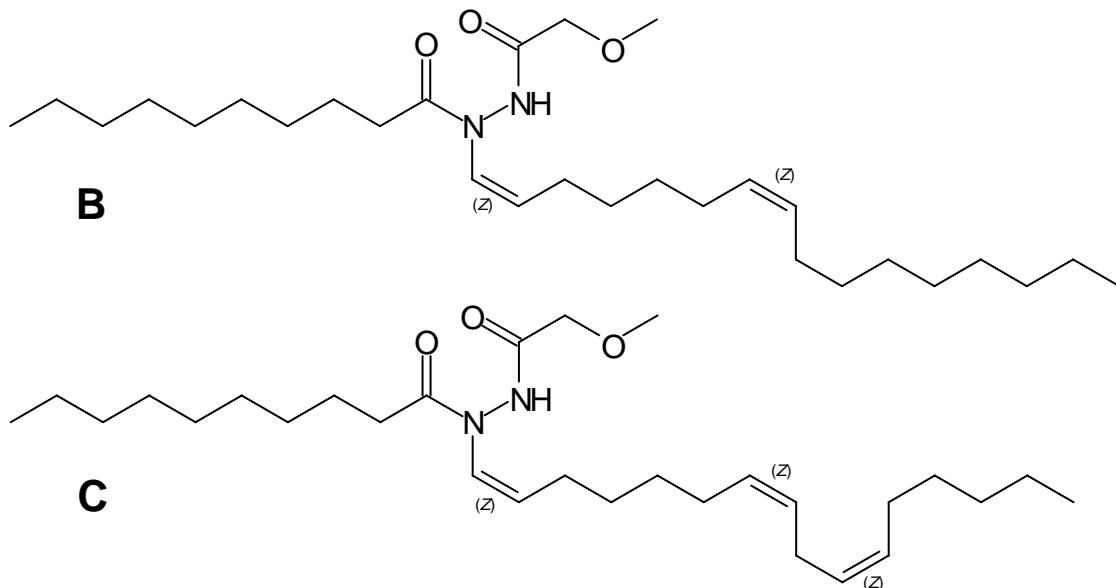
Elaiomycins B and C

Producing organism: *Streptomyces* sp. BK 190



Habitat: terrestrial; hay meadow soil, Cockle Park, Northumberland, UK
Screening method: HPLC-DAD
Biological activity: antibacterial, inhibitors of phosphodiesterase and acetylcholinesterase

Structures:



References:

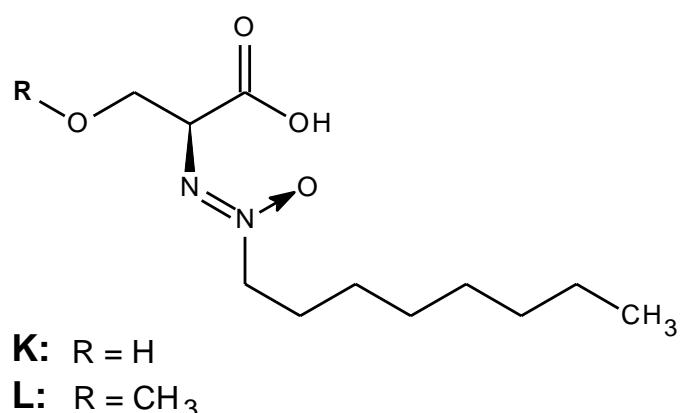
- Kim, B.-Y.; S. Willbold, A. Kulik, S.E. Helaly, H. Zinecker, J. Wiese, J.F. Imhoff, M. Goodfellow, R.D. Süssmuth & H.-P. Fiedler. *J. Antibiotics* 64: 595-597, 2011
Helaly, S.E.; A. Pesic, H.-P. Fiedler & R.D. Süssmuth. *Org. Lett.* 13: 1052-1054, 2011

Elaiomycins K and L

Producing organism: *Streptomyces* sp. Tü 6399



Habitat: terrestrial; rhizosphere of Norway spruce, Rammert Forest, Germany
Screening method: HPLC-DAD
Biological activity: antibacterial
Structures:



References:

Manderscheid, N.; S.E. Helaly, A. Kulik, J. Wiese, J.F. Imhoff, H.-P. Fiedler & R.D. Süssmuth. *J. Antibiotics* 66: 85-88, 2013

Elloxazinones A and B

Producing organism: *Streptomyces griseus* Acta 2871

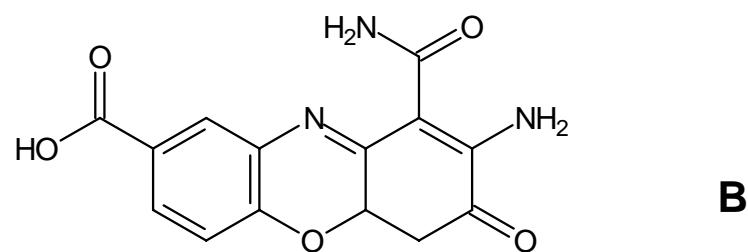
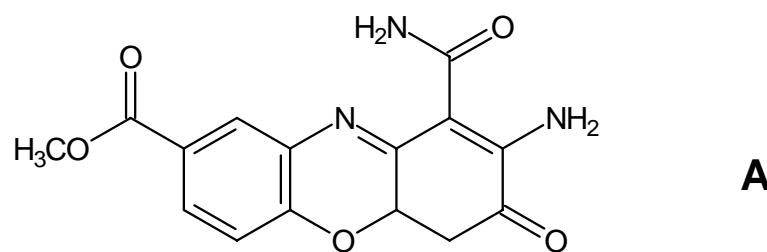


Habitat: terrestrial; steel waste tip soil, Consett, UK

Screening method: HPLC-DAD

Biological activity: antitumor

Structures:

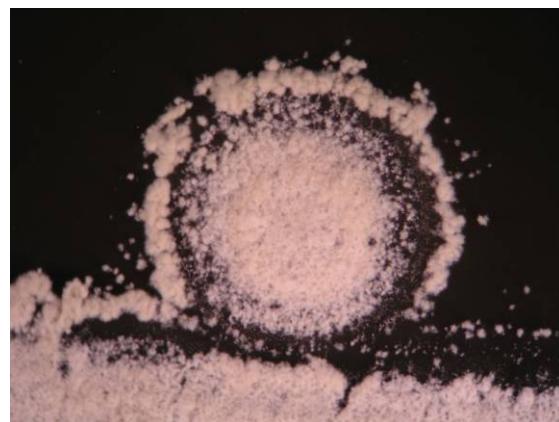


Reference:

Graf, E.; K. Schneider, G. Nicholson, M. Ströbele, A.L. Jones, M. Goodfellow, W. Beil, R.D. Süssmuth & H.-P. Fiedler. J. Antibiotics 60: 277-284, 2007

Endophenazines A–D

Producing organism: *Streptomyces anulatus* LU 9663, LU 9843, LU 9958, LU 10099

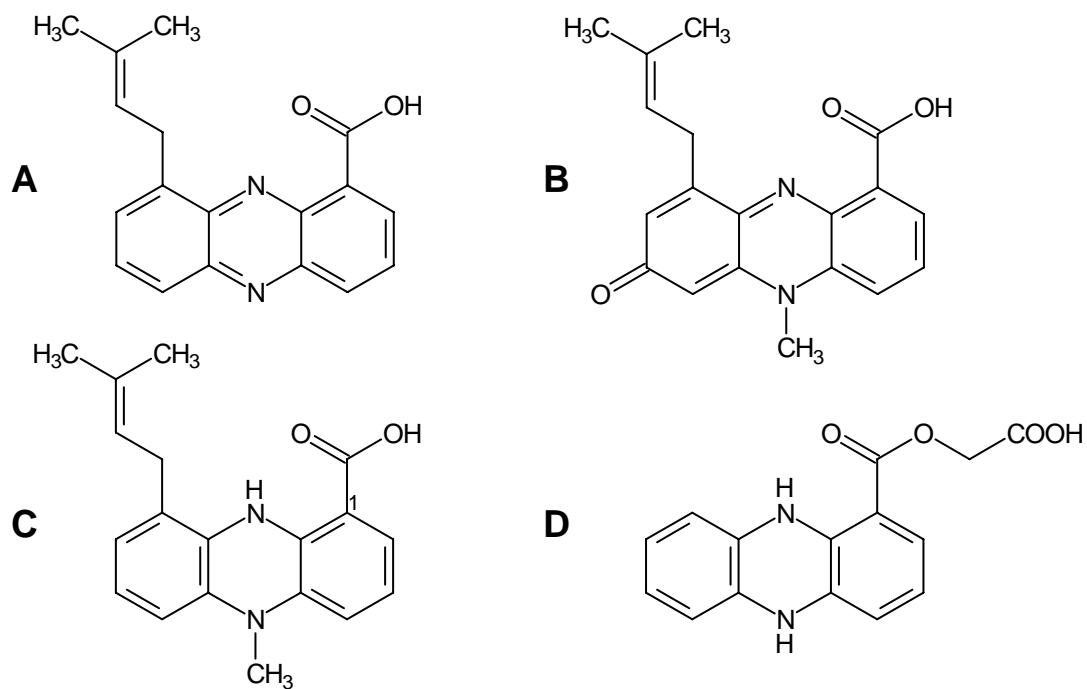


Habitat: endosymbiotic; gut of wood-louse Gen. spec. (LU 9663), gut of leaf-beetle *Exosoma lusitanica* (LU 9843), gut of millipede *Glomeris* spec. (LU 9958), gut of wood-louse *Armadillidium* sp. (LU 10099)

Screening method: HPLC-DAD

Biological activity: antibacterial, antifungal, herbicidal

Structures:



References:

Gebhardt, K.; J. Schimana, P. Krastel, K. Dettner, J. Rheinheimer, A. Zeeck & H.-P. Fiedler: *J. Antibiotics* 55: 794-800, 2002.

Krastel, P.; A. Zeeck, K. Gebhardt, H.-P. Fiedler & J. Rheinheimer. *J. Antibiotics* 55: 801-806, 2002

Fluostatins C–E

Producing organism: *Streptomyces lavendulae* Acta 1383

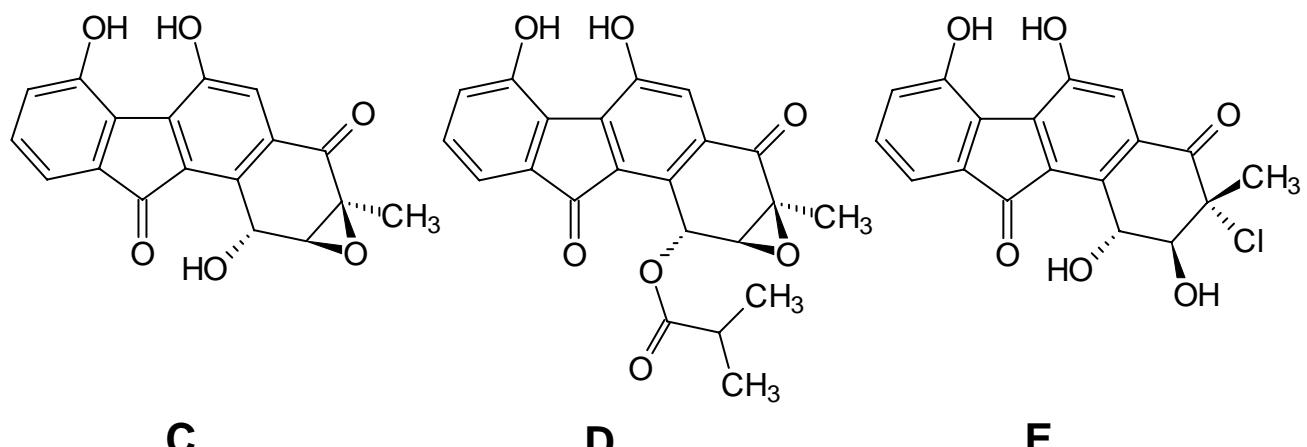


Habitat: terrestrial, rhizosphere; Kaisariani area, Greece

Screening method: HPLC-DAD

Biological activity: antitumor

Structures:



References:

Baur, S.; J. Niehaus, A.D. Karagouni, E.A. Katsifas, K. Chalkou, C. Meintanis, A. Jones, M. Goodfellow, A.C. Ward, W. Beil, K. Schneider, R.D. Süßmuth & H.-P. Fiedler. *J. Antibiotics* 59: 293-297, 2006

Schneider, K.; G. Nicholson, M. Ströbele, S. Baur, J. Niehaus, H.-P. Fiedler & R.D. Süßmuth. *J. Antibiotics* 59: 105-109, 2006

Fogacin

Producing organism: *Streptomyces* sp. Tü 6319

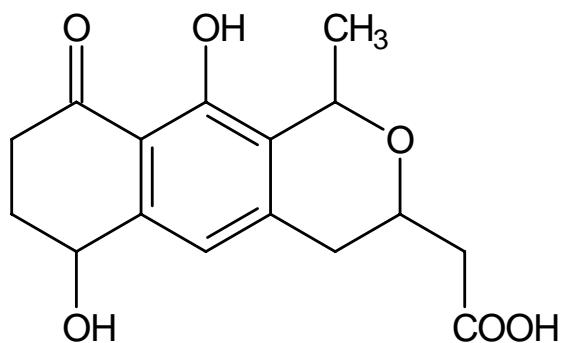


Habitat: terrestrial; Fogaras, Romania

Screening method: HPLC-DAD

Biological activity: antitumor

Structure:



Reference:

Radzom, M.; A. Zeeck, N. Antal & H.-P. Fiedler: J. Antibiotics 59: 315-317, 2006

Fomannoxin Acids DFA, MFA-1 and MFA-2

Producing organism: *Streptomyces* sp. AcH 505

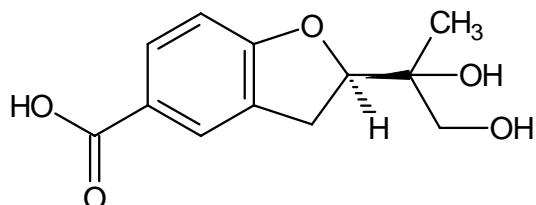


Habitat: terrestrial; rhizospheric soil, Schönbuch Forest, Germany

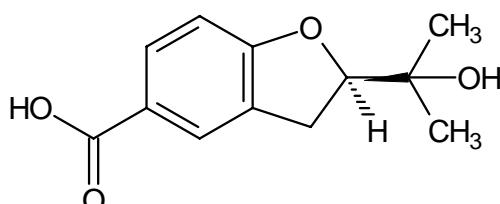
Screening method: HPLC-DAD

Biological activity: inactive conversion products of the fungal phytotoxin fomannoxin from *Heterobasidion* sp.

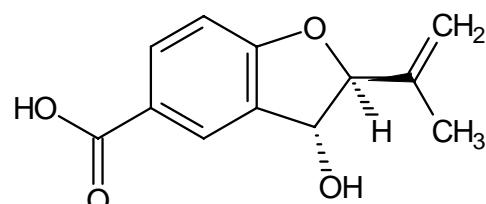
Structures:



DFA
(Dihydroxy-Fomannoxin Acid)



MFA-1
(Monohydroxy-Fomannoxin Acid-1)



MFA-2
(Monohydroxy-Fomannoxin Acid-2)

Reference:

Horlacher, N.; J. Nachtigall, D. Schulz, R.D. Süssmuth, R. Hampp, H.-P. Fiedler & S. Schrey. J. Chem. Ecol. DOI: 10.1007/s10886-013-0290-3, 2013

Frigocyclinone

Producing organism: *Streptomyces griseus* NTK 97

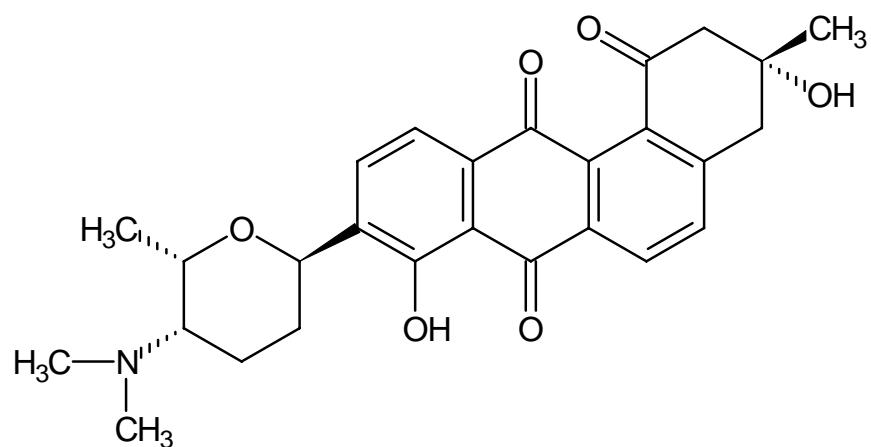


Habitat: terrestrial; Terra Nova Bay, Antarctica

Screening method: HPLC-DAD

Biological activity: antibacterial

Structure:



Reference:

Bruntner,C.; T. Binder, W. Pathom-aree, M. Goodfellow, A.T. Bull, O. Potterat, C. Puder, S. Höller, A. Schmid, W. Bolek, K. Wagner, G. Mihm & H.-P. Fiedler. J. Antibiotics 58: 346-349, 2005

Galtamycin B

Producing organism: *Micromonospora* sp. Tü 6368

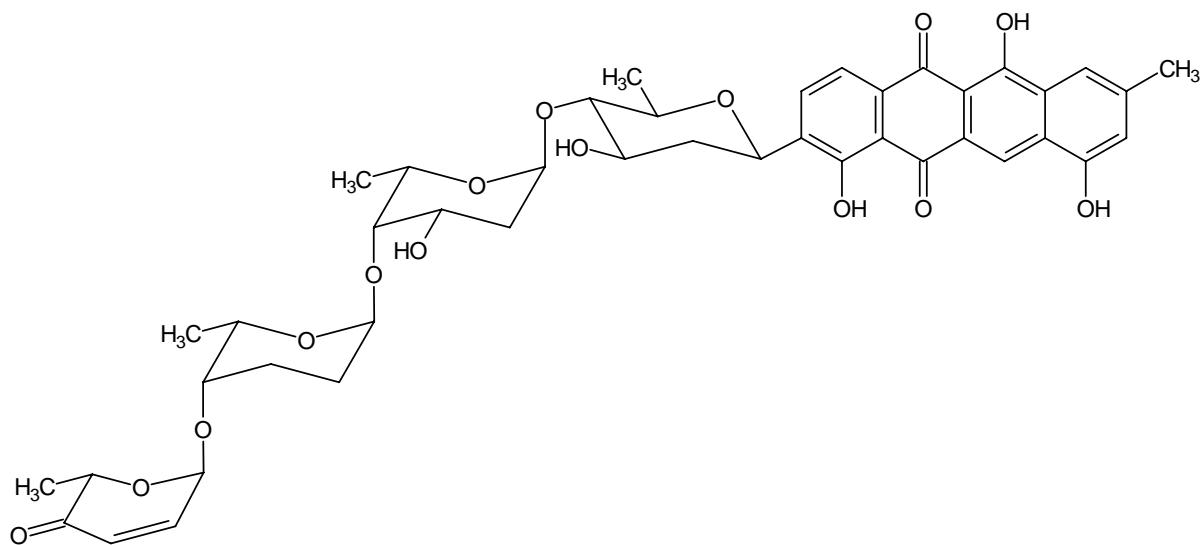


Habitat: terrestrial, Rety, Romania

Screening method: HPLC-DAD

Biological activity: antitumor

Structure:



References:

Antal, N.; H.-P. Fiedler, E. Stackebrandt, W. Beil, K. Ströch & A. Zeeck. J. Antibiotics 58: 95-102, 2005

Ströch, K.; A. Zeeck, N. Antal & H.-P. Fiedler. J. Antibiotics 58: 103-110, 2005

Genoketides A1 and A2, Prechrysophanol-Glucuronide and Chrsophanol-Glucuronide

Producing organism: *Streptomyces* sp. AK 671

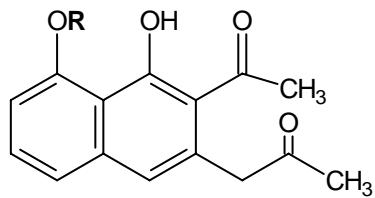


Habitat: terrestrial; Hamsterley Forest, Northumberland, UK

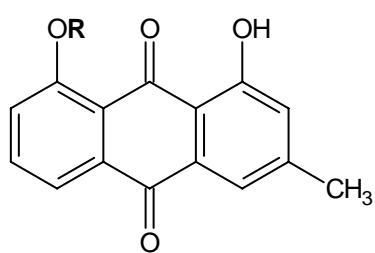
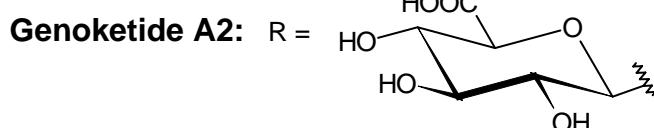
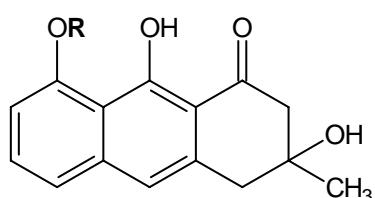
Screening method: HPLC-DAD

Biological activity: antitumor (genoketetide B2)

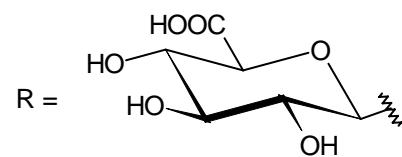
Structures:



Genoketide A1: R = H



Prechrysophanol-Glucuronide



Chrysophanol-Glucuronide

Reference:

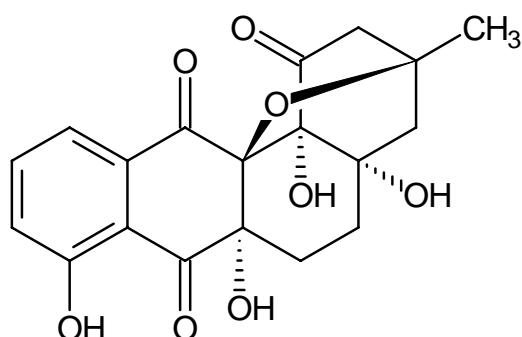
Fiedler, H.-P.; A. Dieter, T.A.M. Gulder, I. Kajahn, A. Hamm, R. Brown, A.L. Jones, M. Goodfellow, W.E.G. Müller & G. Bringmann. J. Antibiotics 61: 464-473, 2008

Gephyromycin

Producing organism: *Streptomyces griseus* NTK 14



Habitat:	terrestrial; Terra Nova Bay, Antarctica
Screening method:	HPLC-DAD
Biological activity:	glutaminergic activity towards neuronal cells
Structure:	

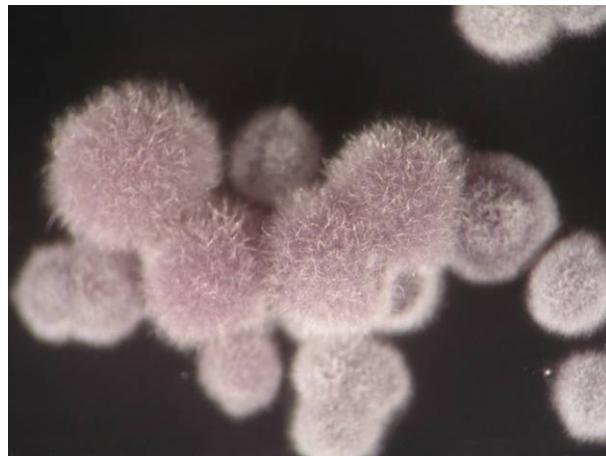


Reference:

Bringmann, G.; G. Lang, K. Maksimenka, A. Hamm, T.A.M. Gulder, A. Dieter, A.T. Bull, J.E.M. Stach, N. Kocher, W.E.G. Müller & H.-P. Fiedler. Phytochemistry 66: 1365-1372, 2005

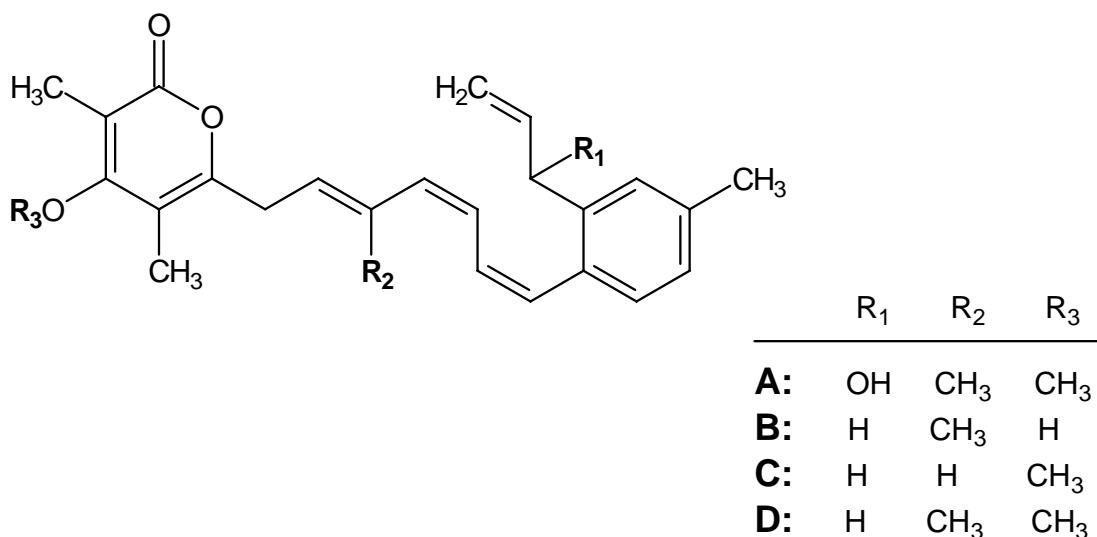
Gombapyrones A–D

Producing organism: *Streptomyces griseoruber* Acta 3662



- Habitat:** terrestrial; rhizosphere of bamboo trees, Gombak, Malaysia
- Screening method:** HPLC-DAD
- Biological activity:** inhibitors of protein tyrosine phosphatase 1B and glycogen synthase kinase 3 β

Structures:



Reference:

Helaly, S.; K. Schneider, J. Nachtingall, S. Vikineswary, G.Y.A. Tan, H. Zinecker, J.F. Imhoff, R.D. Süssmuth & H.-P. Fiedler. *J. Antibiotics* 62: 445-452, 2009

Grecocyclines A and B

Producing organism: *Streptomyces* sp. Acta 1362

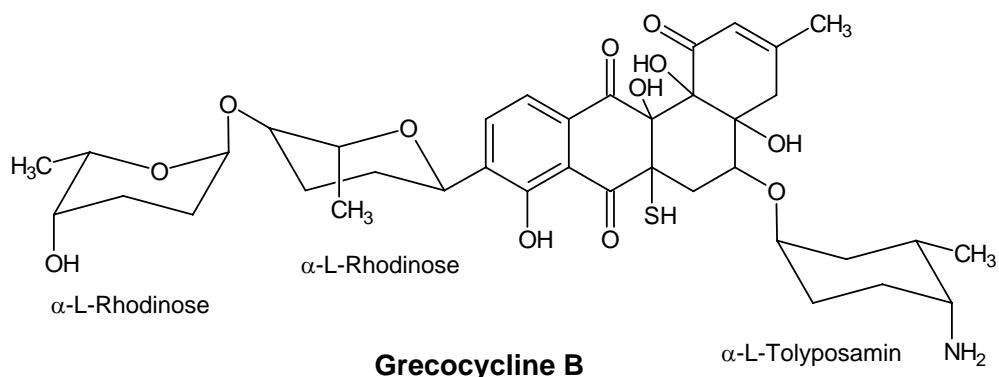
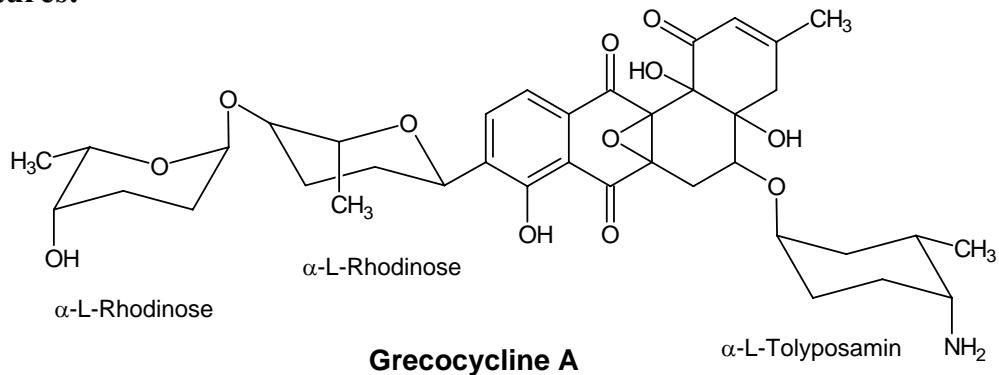


Habitat: terrestrial, rhizosphere; Crete, Greece

Screening method: HPLC-DAD

Biological activity: cytotoxic (A), protein tyrosine phosphatase 1B inhibitor (B)

Structures:



Reference:

Paululat, T.; A. Kulik, H. Hausmann, A.D. Karagouni, H. Zinecker, J.F. Imhoff & H.-P. Fiedler. Eur. J. Org. Chem. 2010: 2344-2350, 2010

Grecoketides A and B

Producing organism: *Streptomyces* sp. Acta 1362

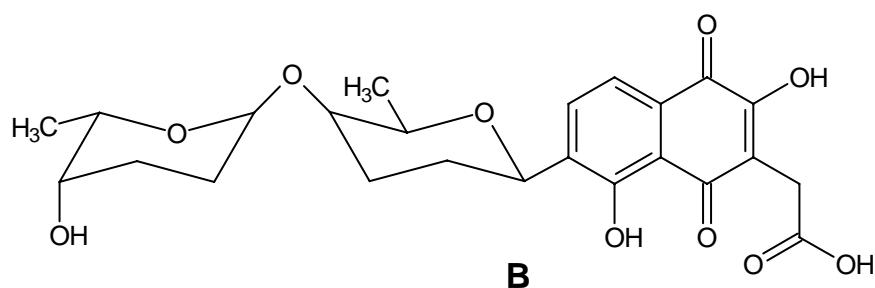
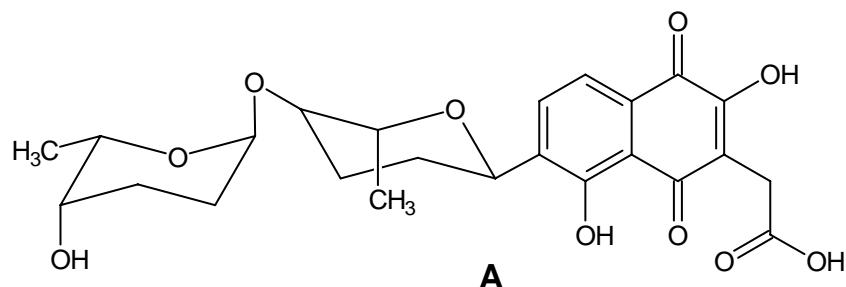


Habitat: terrestrial, rhizosphere; Crete, Greece

Screening method: HPLC-DAD

Biological activity: none

Structures:

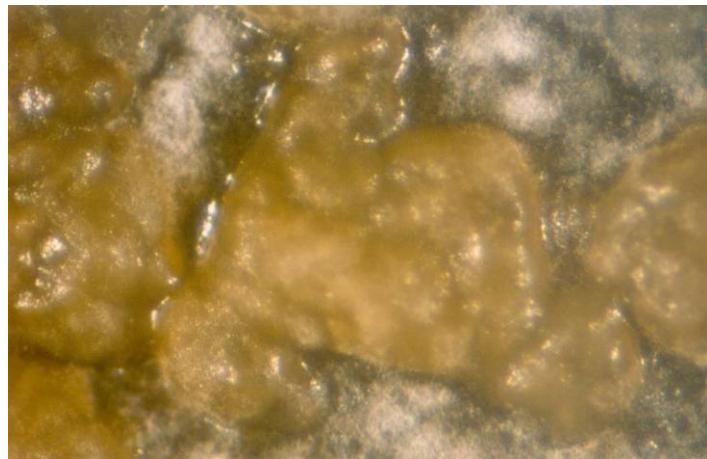


Reference:

Paululat, T.; E.A. Katsifas, A.D. Karagouni & H.-P. Fiedler. Eur. J. Org. Chem. 2008: 5283-5288, 2008

1-Hydroxy-4-Methoxy-2-Naphthoic Acid

Producing organism: *Streptosporangium cinnabarinum* NNO 29536

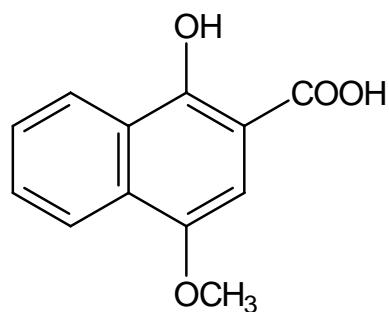


Habitat: terrestrial (ATCC 31213)

Screening method: HPLC-DAD

Biological activity: herbicidal

Structure:



Reference:

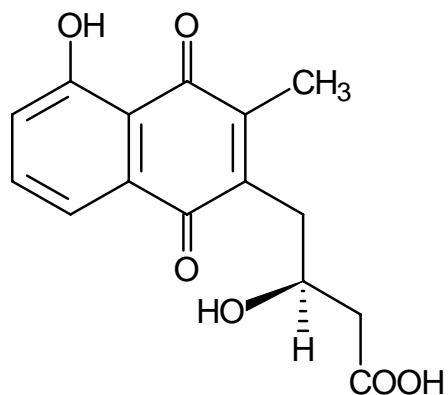
Pfefferle, C.; J. Breinholt, H. Gürtler & H.-P. Fiedler. J. Antibiotics 50: 1067-1068, 1997

Juglomycin Z

Producing organism: *Streptomyces tendae* Tü 901



Habitat: terrestrial; Nikko, Japan
Screening method: HPLC-DAD
Biological activity: antibacterial, antifungal
Structure:

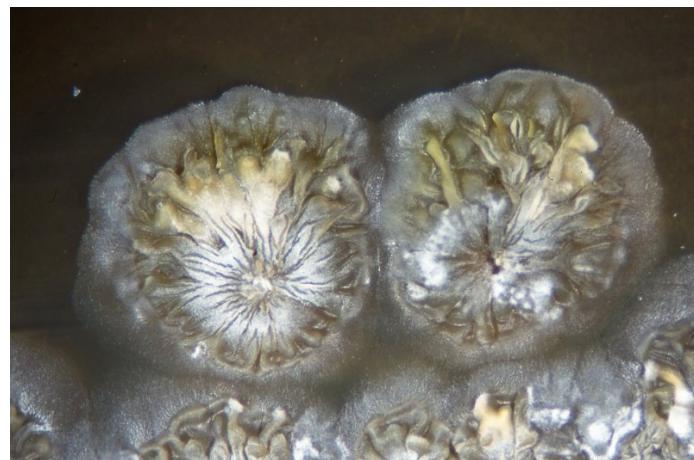


Reference:

Fiedler, H.-P.; A. Kulik, T.C. Schüz, C. Volkmann & A. Zeeck. J. Antibiotics 47: 1116-1122, 1994

Kanchanamycins A, C, D

Producing organism: *Streptomyces olivaceus* Tü 4018

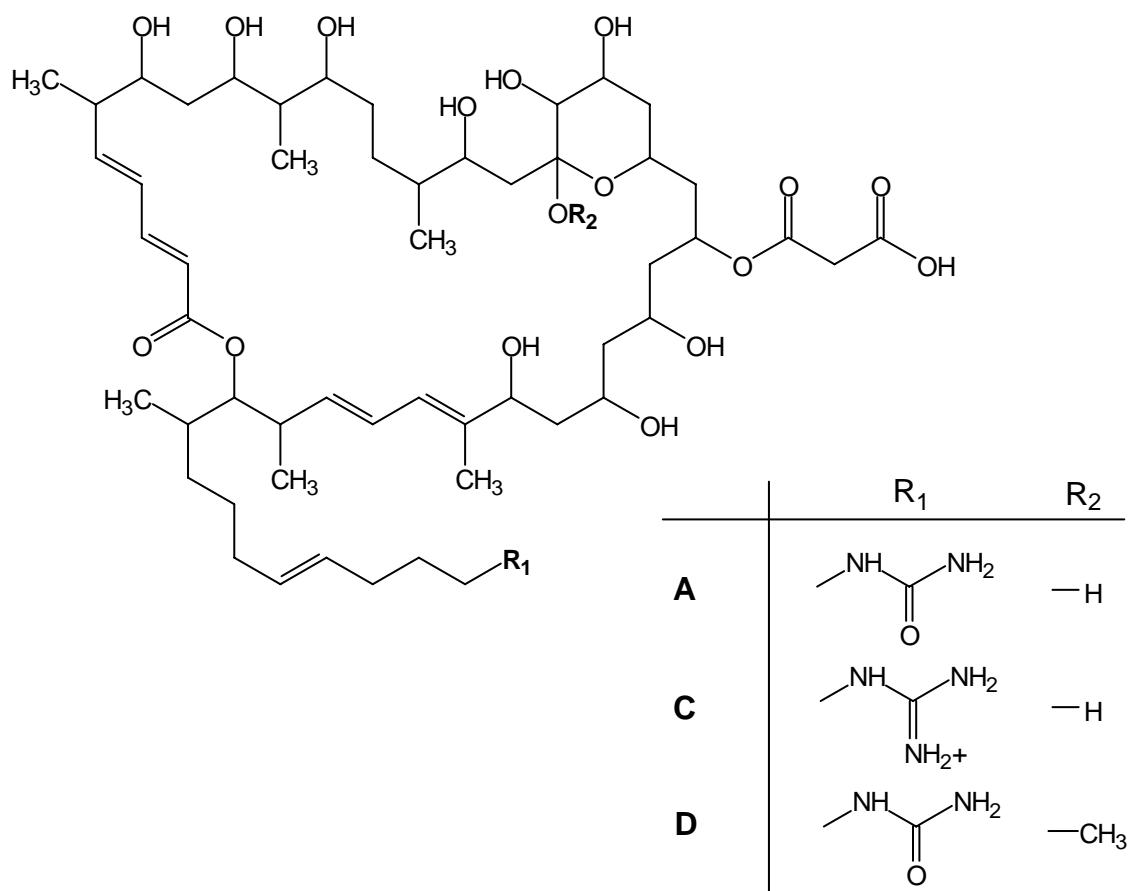


Habitat: terrestrial; Kanchana Buri, Thailand

Screening method: HPLC-DAD

Biological activity: antibacterial, antifungal

Structures:



Kanchanamycins A, C, D

References:

Fiedler, H.-P.; M. Nega, C. Pfefferle, I. Groth, C. Kempfer, H. Stephan & J.W. Metzger. *J. Antibiotics* 49: 101-107, 1996

Stephan, H.; C. Kempfer, J.W. Metzger, G. Jung, O. Potterat, C. Pfefferle & H.-P. Fiedler. *J. Antibiotics* 49: 109-113, 1996

Kyanomycin

Producing organism: *Nonomuria* sp. NN0 22303

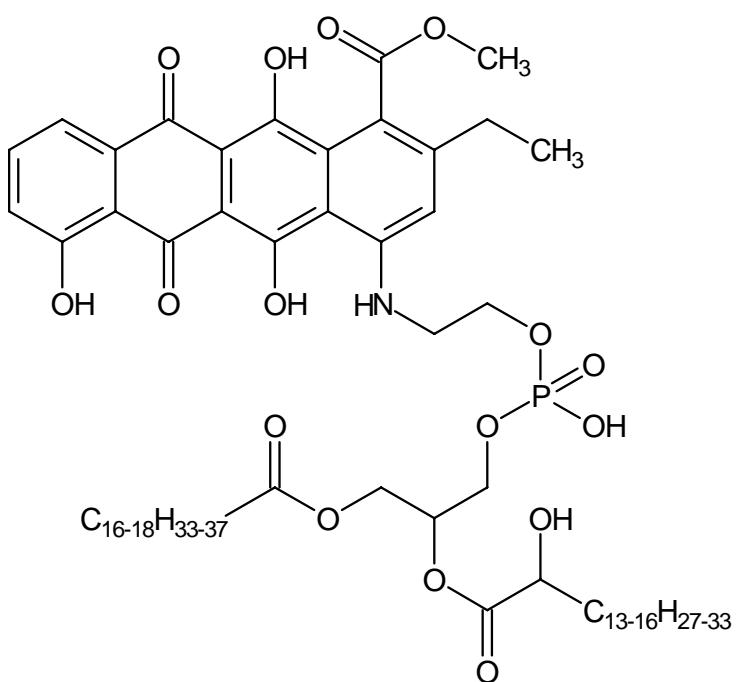


Habitat: terrestrial; India

Screening method: HPLC-DAD

Biological activity: antibacterial

Structure:



Reference:

Pfefferle, C.; J. Breinholt, C.E. Olsen, R.M. Kroppenstedt, E.M.H. Wellington, H. Gürtler & H.-P. Fiedler. *J. Nat. Prod.* 63: 295-298, 2000

Lactonamycin Z

Producing organism: *Streptomyces sanglieri* AK 623

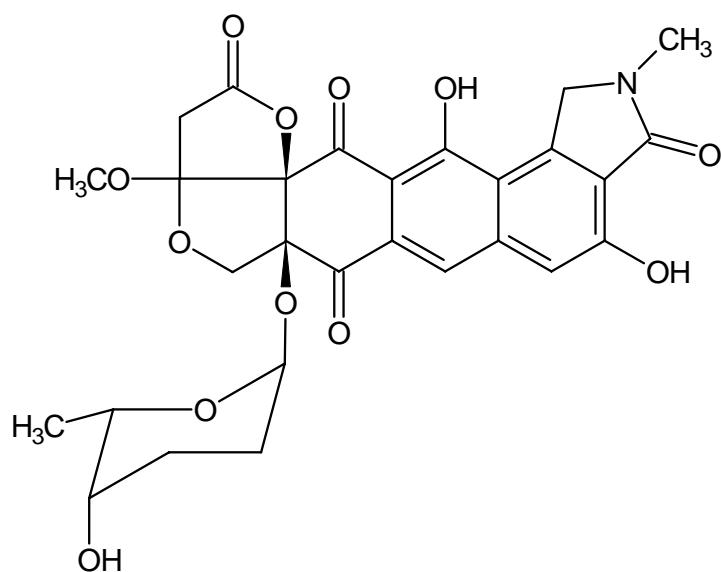


Habitat: terrestrial; Hamsterley Forest, UK

Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor

Structure:



Reference:

Höltzel, A.; A. Dieter, D.G. Schmid, R. Brown, M. Goodfellow, W. Beil, G. Jung & H.-P. Fiedler. J. Antibiotics 56: 1058-1061, 2003

Langkocyclines A1, A2, A3, B1 and B2

Producing organism: *Streptomyces* sp. Acta 3034

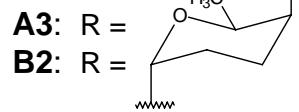
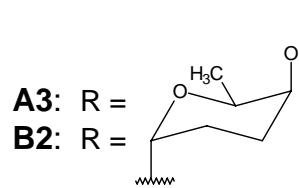
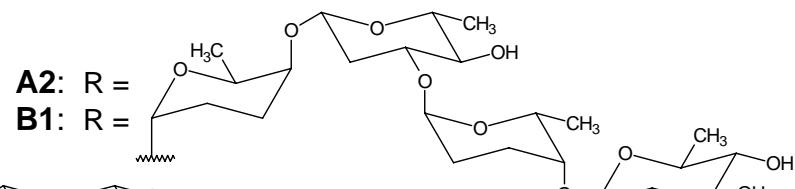
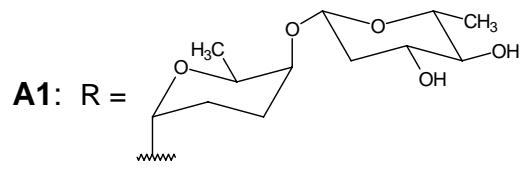
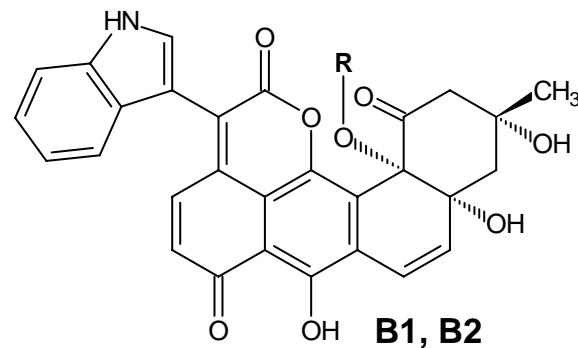
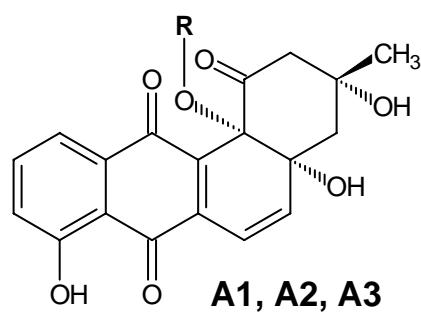


Habitat: terrestrial; rhizospheric soil, Langkawi, Malaysia

Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor

Structures:



Reference:

Kalyon, B.; G.Y.A. Tan, J.M. Pinto, C.Y. Foo, J. Wiese, J.F. Imhoff, R.D. Süßmuth, V. Sabaratnam & H.-P. Fiedler. *J. Antibiotics*, DOI:10.1038/ja.2013.53, 2013

Langkolid

Producing organism: *Streptomyces sp.* Acta 3062

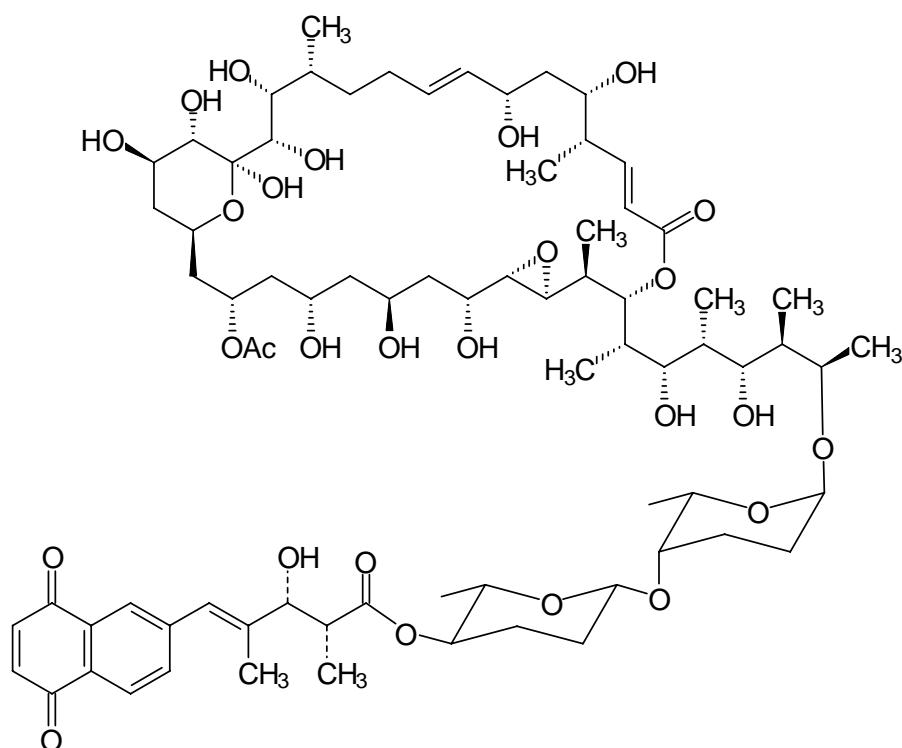


Habitat: terrestrial; rhizospheric soil, Langkawi, Malaysia

Screening method: HPLC-DAD

Biological activity: antifungal, antitumor, inhibitor of phosphodiesterase and glycogen synthase kinase

Structure:



Reference:

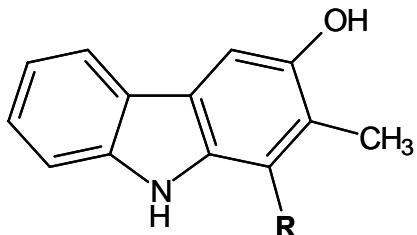
Helaly, S.; A. Kulik, H. Zinecker, K. Ramachandaran, G.Y.A. Tan, J.F. Imhoff, R.D. Süssmuth, H.-P. Fiedler & V. Sabaratnam. *J. Nat. Prod.* 75: 1018-1024, 2012

Lipocarbazoles A1–A4

Producing organism: *Tsukamurella pseudospumae* Acta 1857



Habitat: activated sludge foam, Stoke Bardolph, UK
Screening method: HPLC-DAD
Biological activity: free radical scavengers, antioxidant
Structures:



A1: R = $(\text{CH}_2)_7\text{CH}=\text{CHCH}_2\text{CH}=\text{CHCH}_2\text{CH}=\text{CHCH}_2\text{CH}_3$

A2: R = $(\text{CH}_2)_7\text{CH}=\text{CHCH}_2\text{CH}=\text{CH}(\text{CH}_2)_4\text{CH}_3$

A3: R = $(\text{CH}_2)_7\text{CH}=\text{CH}(\text{CH}_2)_7\text{CH}_3$

A4: R = $(\text{CH}_2)_{16}\text{CH}_3$

Reference:

Schneider, K.; J. Nightigall, A. Hänenchen, G. Nicholson, M. Goodfellow, R.D. Süssmuth & H.-P. Fiedler. *J. Nat. Prod.* 72: 1768-1772, 2009

Naphthgeranine F

Producing organism: *Streptomyces violaceus* Tü 3556

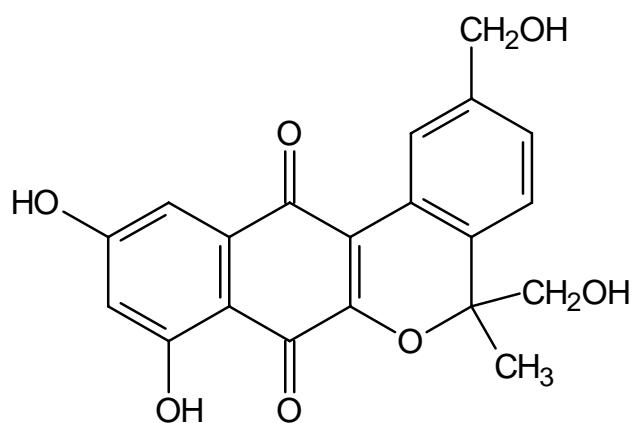


Habitat: terrestrial; Dakshinkali, Nepal

Screening method: HPLC-DAD

Biological activity: antibacterial, antifungal

Structure:



Reference:

Volkmann, C.; U. Hartjen, A. Zeeck & H.-P. Fiedler. J. Antibiotics 48: 522-524, 1995

Nataxazole

Producing organism: *Streptomyces* sp. Tü 6176

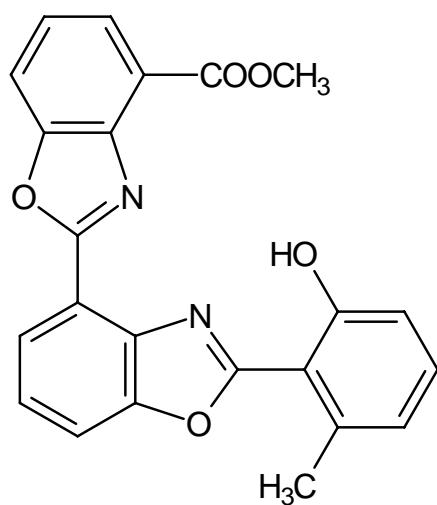


Habitat: terrestrial; Natal, Brazil

Screening method: HPLC-DAD

Biological activity: antitumor

Structure:



Reference:

Sommer, P.S.M.; R.C. Almeida, K. Schneider, W. Beil, R.D. Süssmuth & H.-P. Fiedler: J. Antibiotics 61: 683-686, 2008

Nocardichelins A and B

Producing organism: *Nocardia* sp. Acta 3026

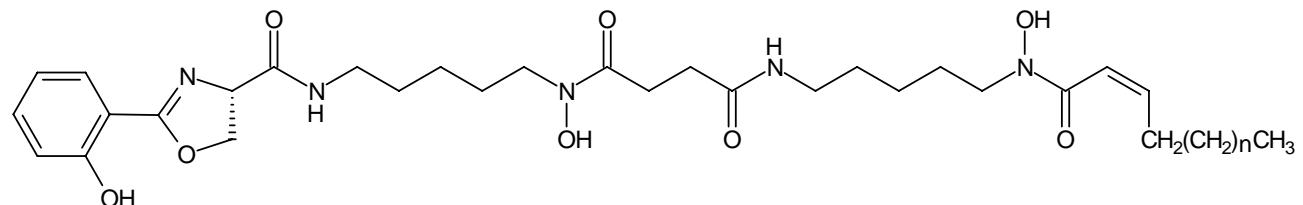


Habitat: terrestrial, rhizosphere; Morib, Selangor, Malaysia

Screening method: HPLC-DAD

Biological activity: siderophor

Structures:



Nocardichelin A: n = 11

Nocardichelin B: n = 9

Reference:

Schneider, K.; I. Rose, S. Vikineswary, A.L. Jones, M. Goodfellow, G. Nicholson, W. Beil, R. Süßmuth & H.-P. Fiedler. J. Nat. Prod. 70: 932-935, 2007

(E)-4-Oxonon-2-Enoic Acid

Producing organism: *Streptomyces olivaceus* Tü 4018

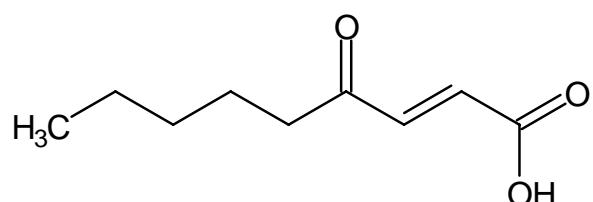


Habitat: terrestrial; Kanchana Buri, Thailand

Screening method: HPLC-DAD

Biological activity: antibacterial

Structure:



Reference:

Pfefferle, C.; C. Kempfer, J.W. Metzger & H.-P. Fiedler. J. Antibiotics 49: 826-828, 1996

Phenalinolactones A–D

Producing organism: *Streptomyces* sp. Tü 6071

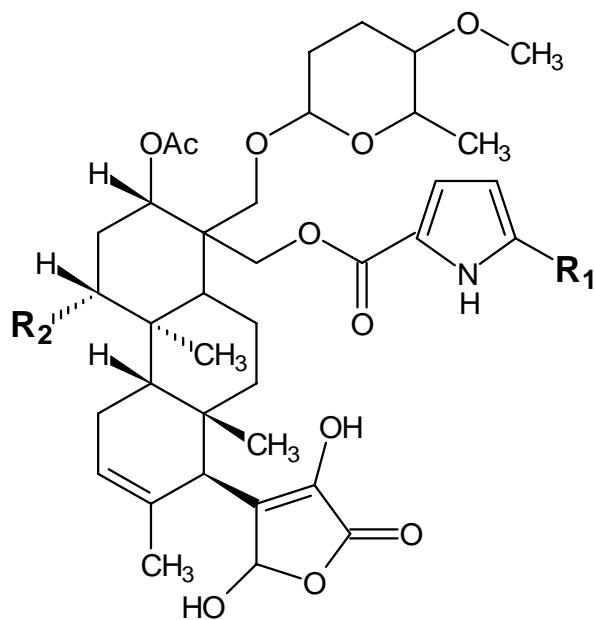


Habitat: terrestrial; Cape Coast, Ghana

Screening method: HPLC-DAD

Biological activity: antibacterial against Gram-positive bacteria

Structures:



Phenalinolactones

A : $R_1 = CH_3$; $R_2 = OH$

B : $R_1 = H$; $R_2 = OH$

C : $R_1 = CH_2OCH_3$; $R_2 = OH$

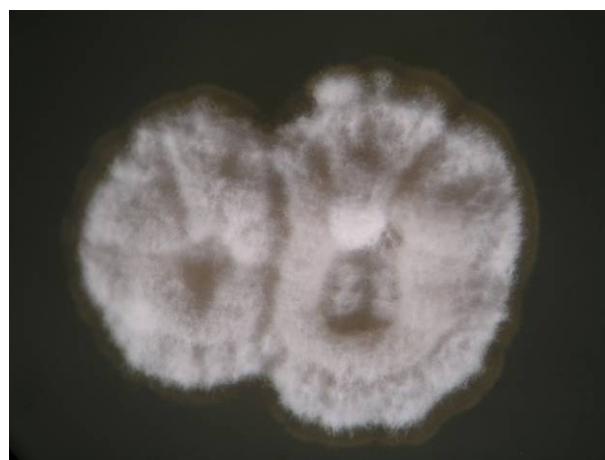
D : $R_1 = CH_3$; $R_2 = H$

Reference:

Gebhardt, K.; S.W. Meyer, J. Schinko, G. Bringmann, A. Zeeck & H.-P. Fiedler. J. Antibiotics 64: 229-232, 2011

Phenelfamycins G and H

Producing organism: *Streptomyces albospinus* Acta 3619

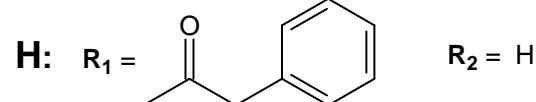
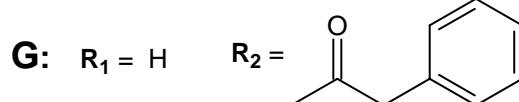
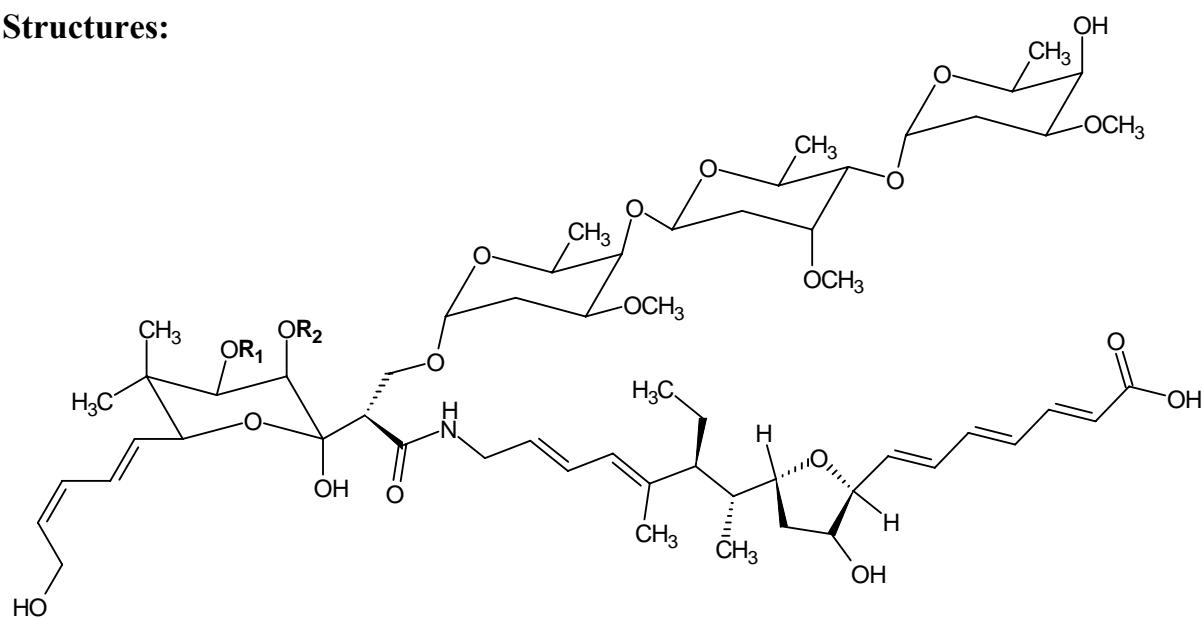


Habitat: terrestrial; rhizosphere soil from Gombak, Malaysia

Screening method: HPLC-DAD

Biological activity: narrow antibacterial spectrum

Structures:



Reference:

Brötz, E.; A. Kulik, S. Vikineswary, C.-T. Lim, G.Y.A. Tan, H. Zinecker, J.F. Imhoff, T. Paululat & H.-P. Fiedler. *J. Antibiotics* 64: 257-266, 2011

Piceamycin

Producing organism: *Streptomyces* sp. GB 4-2

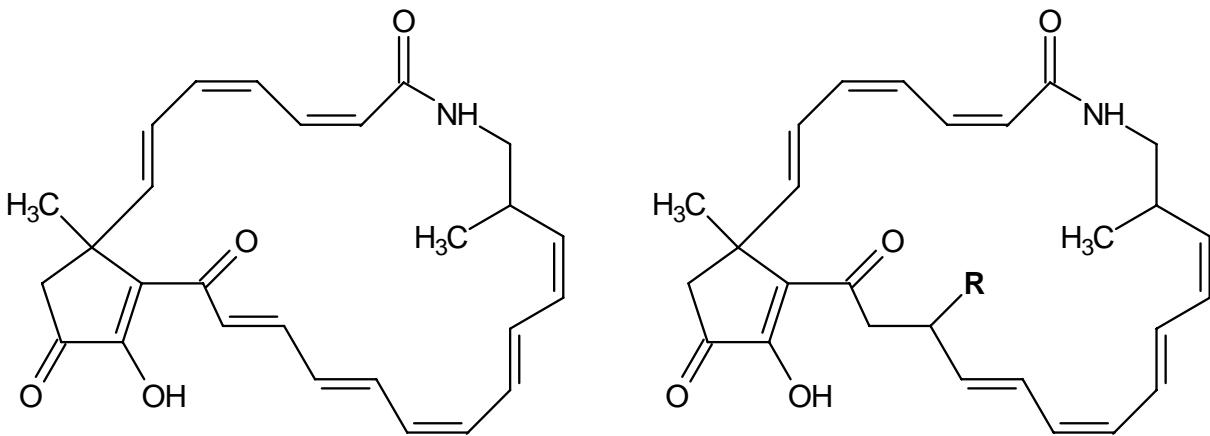


Habitat: terrestrial; rhizosphere of Norway spruce, Schönbuch , Germany

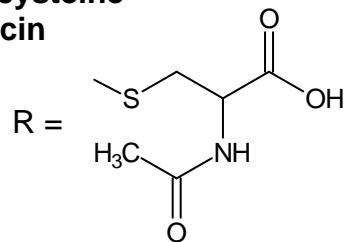
Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor, inhibitor of protein tyrosine phosphatase 1B

Structures:



***N*-acetylcysteine
piceamycin**



Reference:

Schulz, D.; J. Nachtigall, J. Riedlinger, K. Schneider, K. Poralla, J.F. Imhoff, W. Beil, G. Nicholson, H.-P. Fiedler & R.D. Süßmuth. *J. Antibiotics* 62, 513-518, 2009

Polyketomycin

Producing organism: *Streptomyces diastatochromogenes*. Tü 6028

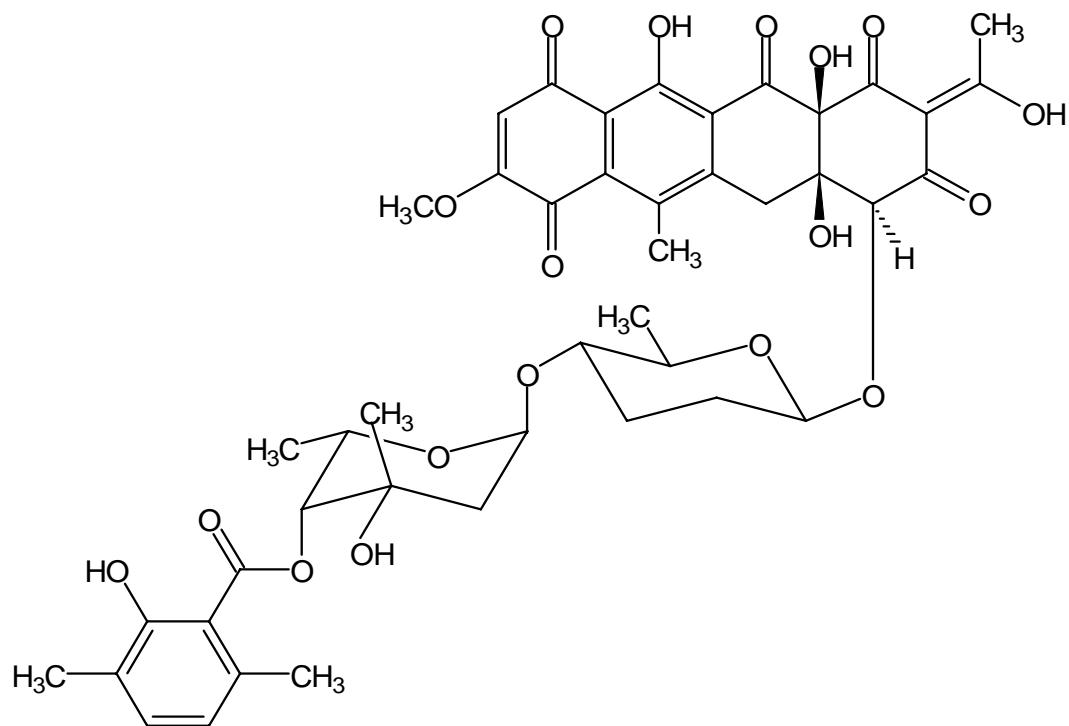


Habitat: terrestrial; Iguaçu, Argentina

Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor

Structure:



Reference:

Paululat, T.; A. Zeeck, J.M. Gutterer & H.-P. Fiedler. J. Antibiotics 52: 96-101, 1999

Proximicins A–C

Producing organism: *Verrucospora fiedleri* MG-37

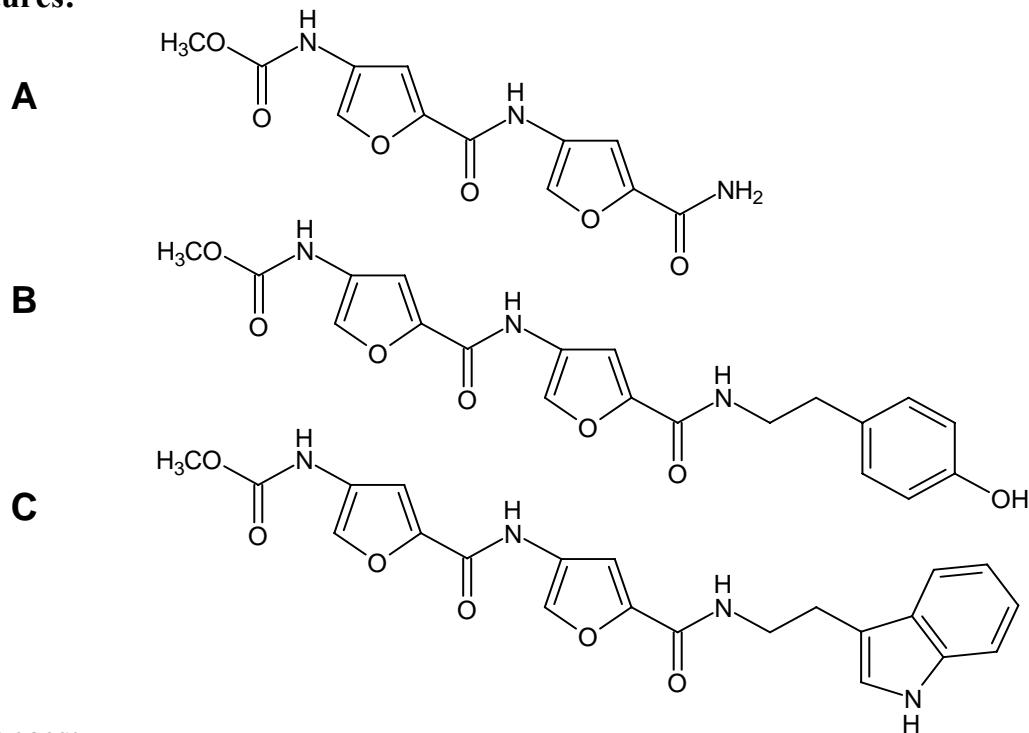


Habitat: marine; sediment –250 m, Raune Fjord, Norway

Screening method: HPLC-DAD

Biological activity: antitumor

Structures:



References:

Fiedler, H.-P.; C. Bruntner, J. Riedlinger, A.T. Bull, G. Knutsen, A.L. Jones, M. Goodfellow, W. Beil, K. Schneider, S. Keller & R.D. Süßmuth. *J. Antibiotics* 61: 158-163, 2008

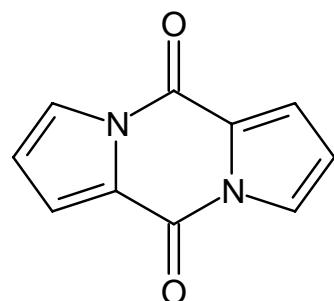
Schneider, K.; S. Keller, F.E. Wolter, L. Röglin, W. Beil, O. Seitz, G. Nicholson, C. Bruntner, J. Riedlinger, H.-P. Fiedler & R.D. Süßmuth: *Angew. Chem. Int. Ed.* 47: 3258-3261, 2008

Pyrocoll

Producing organism: *Streptomyces* sp. AK 409



Habitat: terrestrial; steel waste tip soil, Consett, UK
Screening method: HPLC-DAD
Biological activity: antibacterial, antiparasitic, antitumor
Structure:

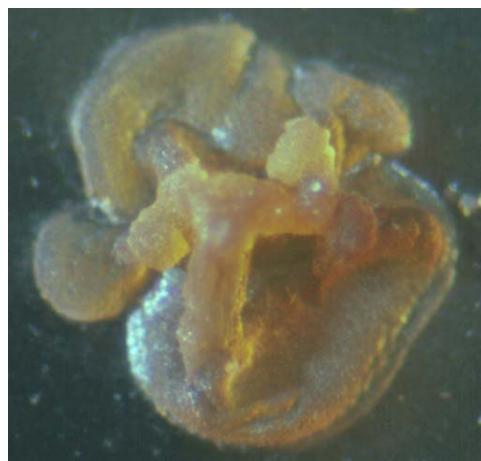


Reference:

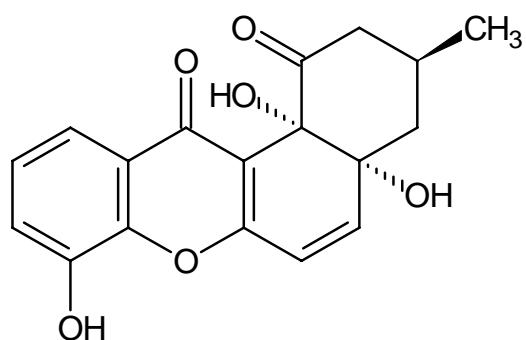
Dieter, A.; A. Hamm, H.-P. Fiedler, M. Goodfellow, W.E.G. Müller, R. Brun, W. Beil & G. Bringmann. J. Antibiotics 56: 639-646, 2003

Retymicin

Producing organism: *Micromonospora* sp. Tü 6368



Habitat: terrestrial, Rety, Romania
Screening method: HPLC-DAD
Biological activity: antitumor
Structure:

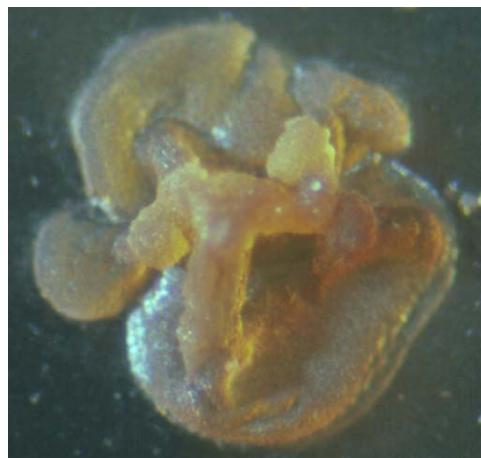


References:

- Antal, N.; H.-P. Fiedler, E. Stackebrandt, W. Beil, K. Ströch & A. Zeeck. J. Antibiotics 58: 95-102, 2005
Ströch, K.; A. Zeeck, N. Antal & H.-P. Fiedler. J. Antibiotics 58: 103-110, 2005

1-(α -Ribofuranosyl)-lumichrome

Producing organism: *Micromonospora* sp. Tü 6368

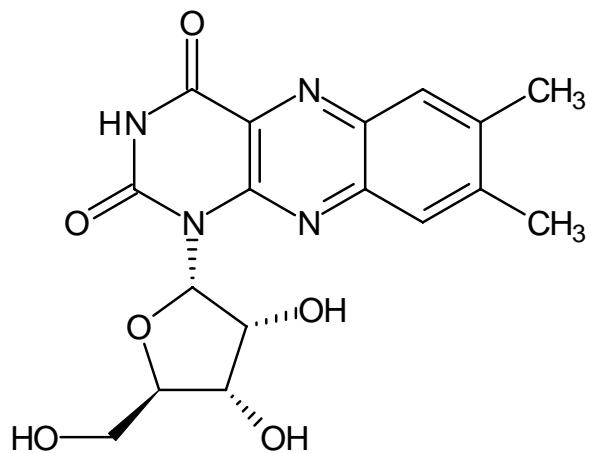


Habitat: terrestrial, Rety, Romania

Screening method: HPLC-DAD

Biological activity: none

Structure:



References:

Antal, N.; H.-P. Fiedler, E. Stackebrandt, W. Beil, K. Ströch & A. Zeeck. J. Antibiotics 58: 95-102, 2005

Ströch, K.; A. Zeeck, N. Antal & H.-P. Fiedler. J. Antibiotics 58: 103-110, 2005

Ripromycin

Producing organism: *Streptomyces* sp. Tü 6239

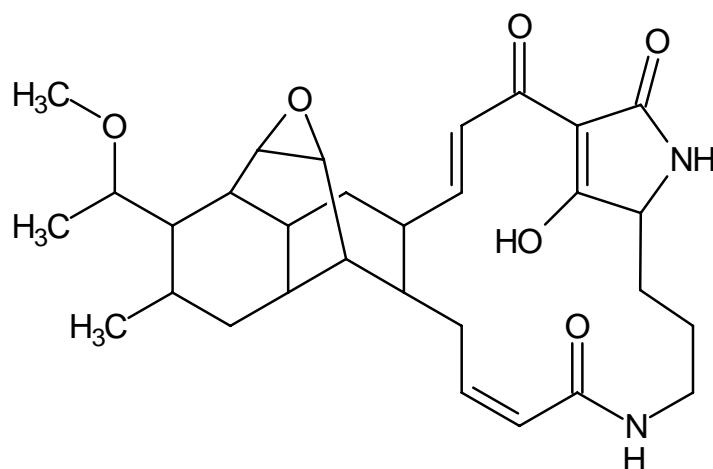


Habitat: terrestrial, São José do Rio Preto, Brazil

Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor

Structure:

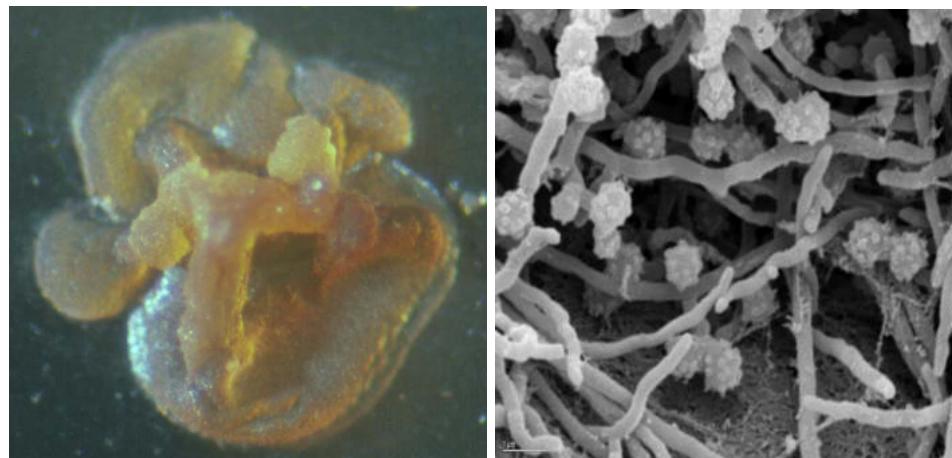


Reference:

Bertasso, M.; M. Holzenkämpfer, A. Zeeck, E. Stackebrandt, W. Beil & H.-P. Fiedler. J. Antibiotics 56: 364-371, 2003

Saquayamycin Z

Producing organism: *Micromonospora* sp. Tü 6368

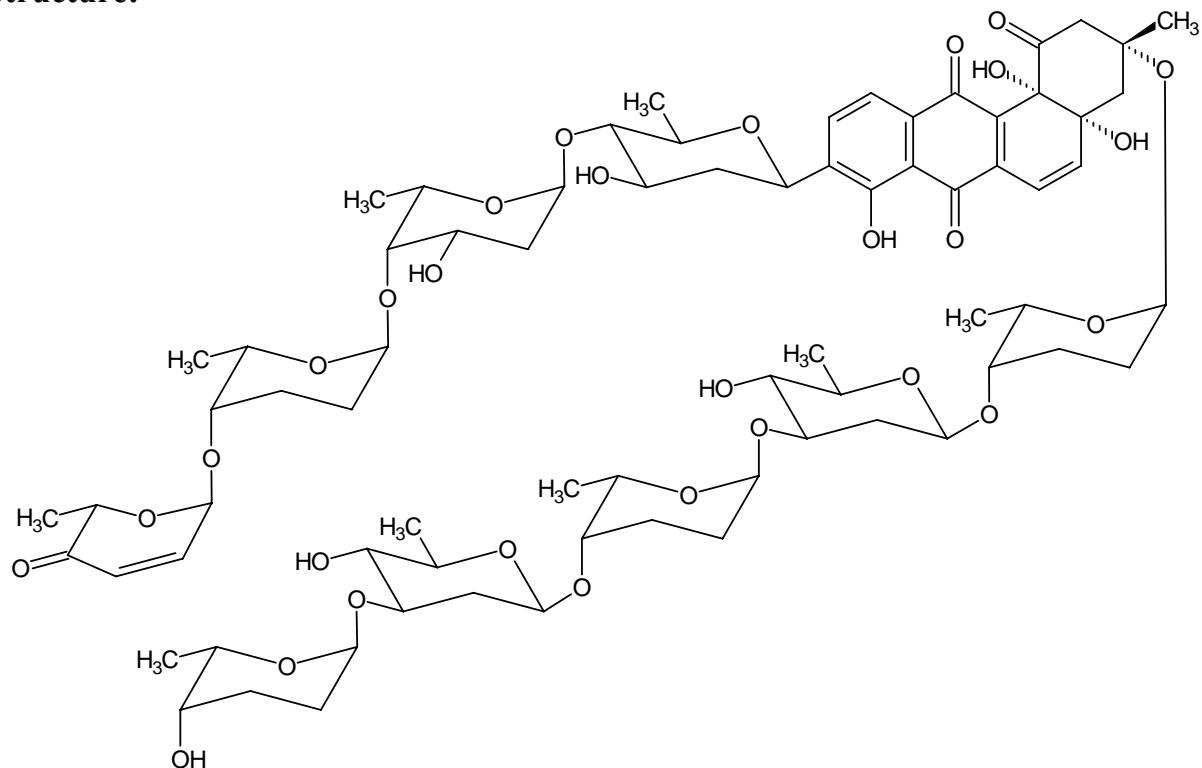


Habitat: terrestrial, Rety, Romania

Screening method: HPLC-DAD

Biological activity: antibacterial, antitumor

Structure:



References:

Antal, N.; H.-P. Fiedler, E. Stackebrandt, W. Beil, K. Ströch & A. Zeeck. J. Antibiotics 58: 95-102, 2005

Ströch, K.; A. Zeeck, N. Antal & H.-P. Fiedler. J. Antibiotics 58: 103-110, 2005

Silvalactam

Producing organism: *Streptomyces* sp. Tü 6392

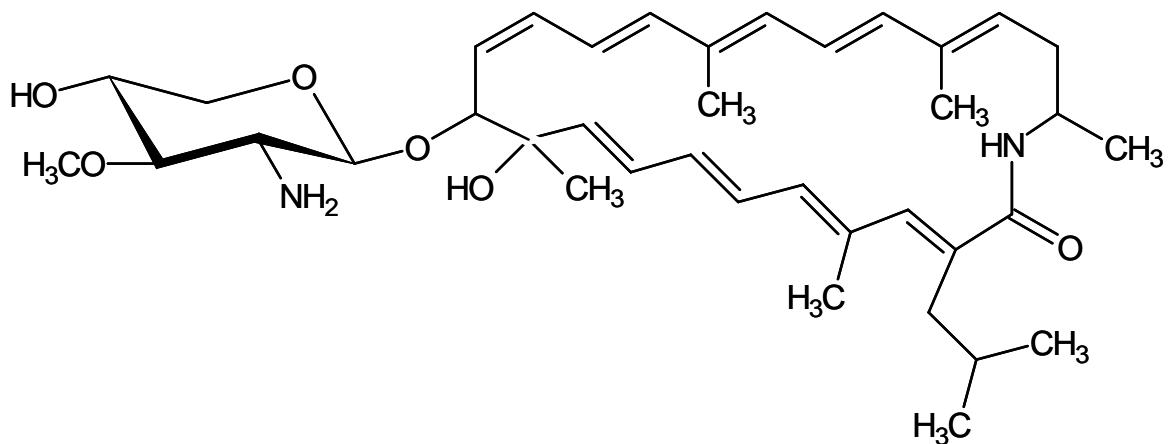


Habitat: terrestrial; rhizosphere of Norway spruce, Rammert Forest, Tübingen, Germany

Screening method: HPLC-DAD

Biological activity: antitumor

Structure:



Reference:

Schulz, D.; J. Nachtingall, U. Geisen, H. Kalthoff, J.F. Imhoff, H.-P. Fiedler & R.D. Süssmuth. *J. Antibiotics* 65: 369-372, 2012

Simocyclinones A–D

Producing organism: *Streptomyces antibioticus* Tü 6040



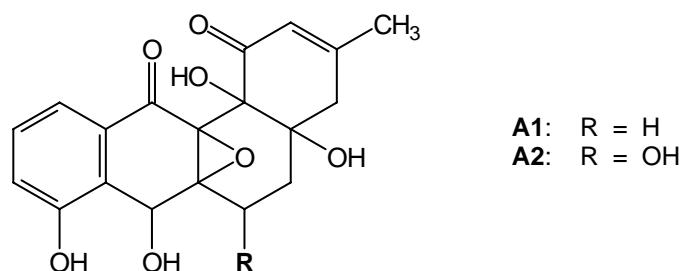
Habitat: terrestrial; Iguaçu Falls, Argentina

Screening method: HPLC-DAD

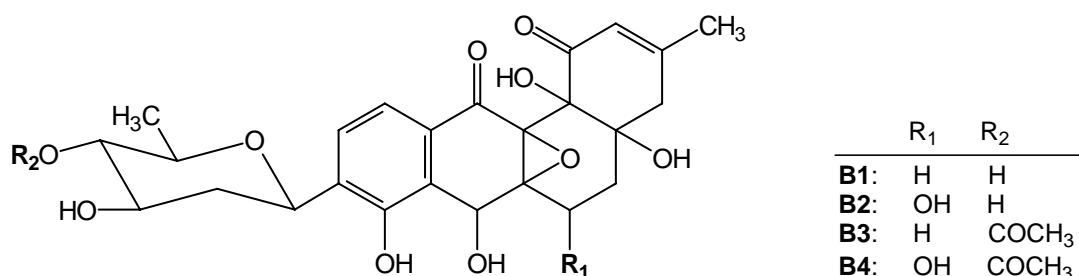
Biological activity: antibacterial, inhibitors of bacterial gyrase (simocyclinones D); antitumor, inhibitors of protein kinase C (simocyclinones C and D)

Structures:

Simocyclinones A

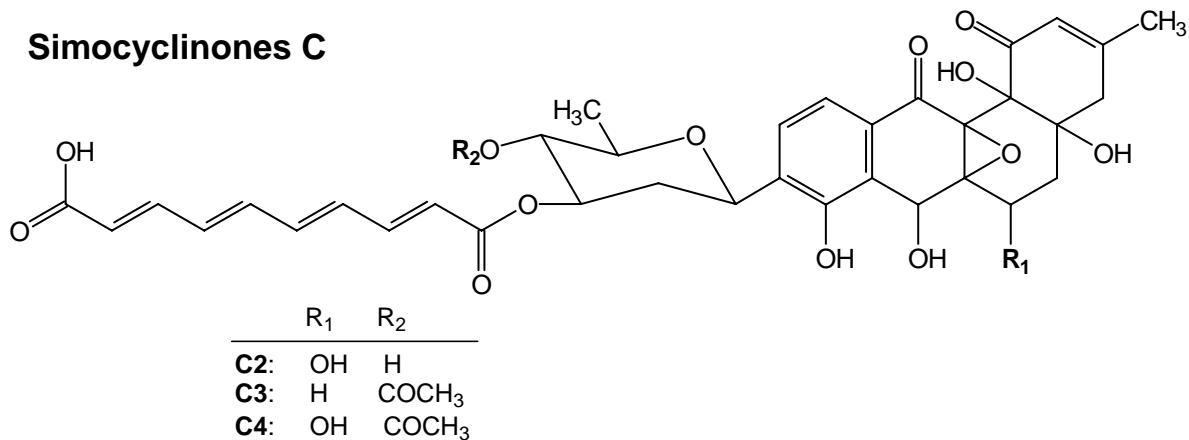


Simocyclinones B

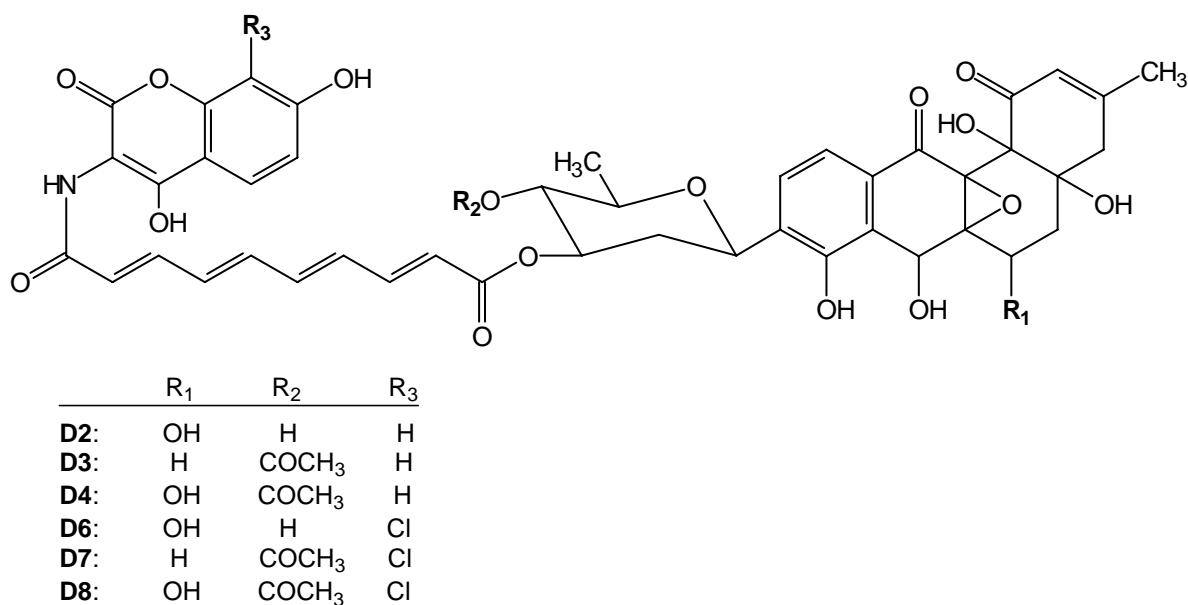


Simocyclinones A–D

Simocyclinones C



Simocyclinones D



Simocyclinones A–D

References:

- Schimana, J.; H.-P. Fiedler, I. Groth, R. Süssmuth, W. Beil, M. Walker & A. Zeeck. *J. Antibiotics* 53: 779-787, 2000
- Theobald, U.; J. Schimana & H.-P. Fiedler. *Antonie van Leeuwenhoek* 78: 307-313, 2000
- Schimana, J.; M. Walker, A. Zeeck & H.-P. Fiedler. *J. Ind. Microbiol. Biotechnol.* 27: 144-148, 2001
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- Oppgaard, L.M.; B.L. Hamann, K.R. Streck, K.C. Ellis, H.-P. Fiedler, A.B. Khodursky & H. Hiasa. *Antimicrob. Agents Chemother.* 53:2110-2119, 2009
- Le, T.B.K.; H.-P. Fiedler, C.D. den Hengst, S.K. Ahn, A. Maxwell & M.J. Buttner. *Molec. Microbiol.* 72: 1462-1474, 2009
- Edwards, M.J.; R.H. Flatman, L.A. Mitchenall, C.E.M. Stevenson, T.B.K. Le, T.A. Clarke, A.R. McKay, H.-P. Fiedler, M.J. Buttner, D.M. Lawson & A. Maxwell. *Science* 326: 1415-1417, 2009

Skyllamycins A and B

Producing organism: *Streptomyces* sp. Acta 2897

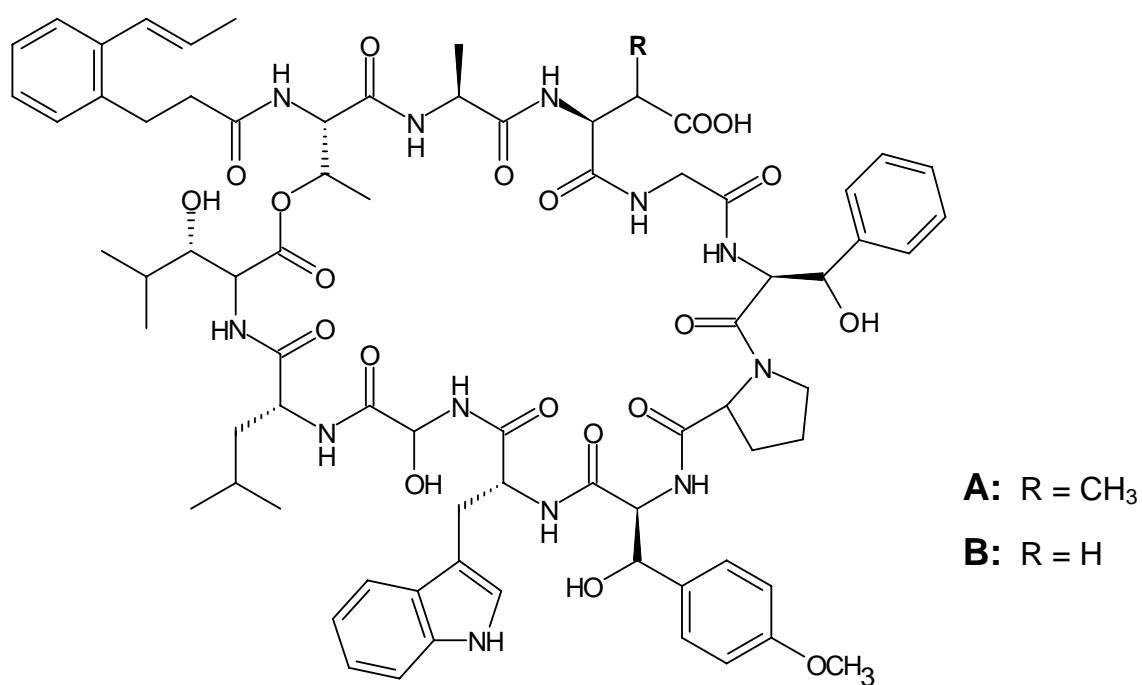


Habitat: terrestrial; Dune Slack, Warkworth, UK

Screening method: HPLC-DAD

Biological activity: antibacterial

Structures:



Reference:

Pohle, S.; C. Appelt, M. Roux, H.-P. Fiedler & R.D. Süssmuth. J. Am. Chem. Soc. 133: 6194-6205, 2011

Spirodionic Acid Tü 6077

Producing organism: *Streptomyces* sp. Tü 6077

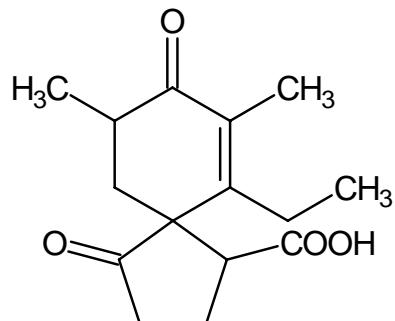


Habitat: terrestrial; Cape Coast, Ghana

Screening method: HPLC-DAD

Biological activity: none

Structure:

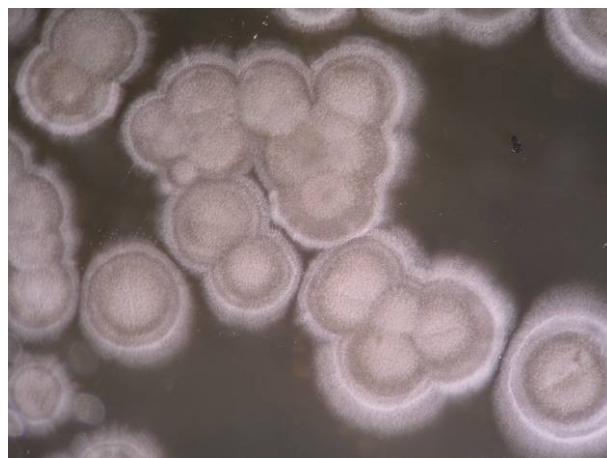


Reference:

Textor, A.; I. Papastavrou, J. Siewert, J. Magull, A. Kulik, H.-P. Fiedler, P. von Zezschwitz & S. Grond. Chem. Eur. J. 13: 7416-7423, 2007

Spirofungin

Producing organism: *Streptomyces violaceusniger* Tü 4113

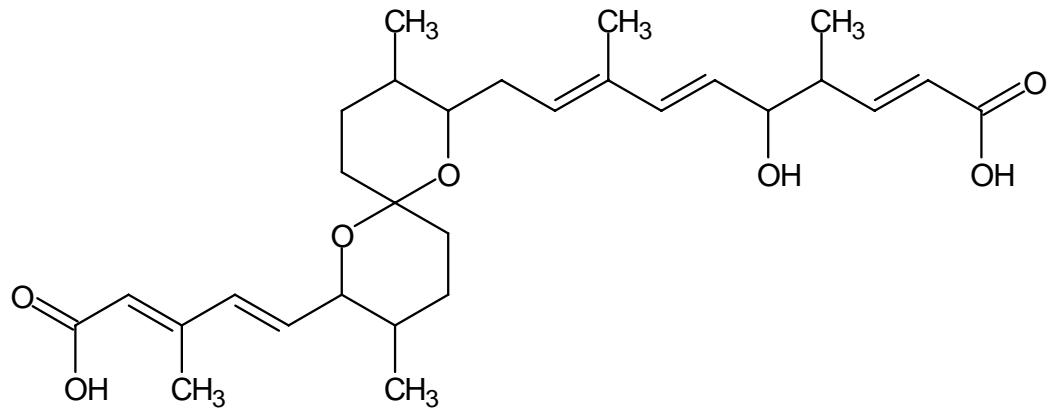


Habitat: terrestrial; Otway National Park, Australia

Screening method: HPLC-DAD

Biological activity: antifungal

Structure:



Reference:

Höltzel, A.; C. Kempfer, J.W. Metzger, G. Jung, I. Groth, T. Fritz & H.-P. Fiedler. J. Antibiotics 51: 699-707, 1998

Streptocidins A–D

Producing organism: *Streptomyces* sp. Tü 6071

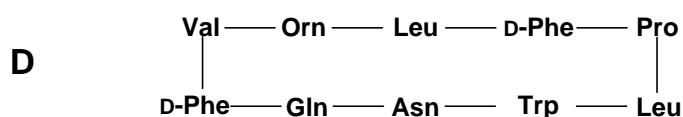
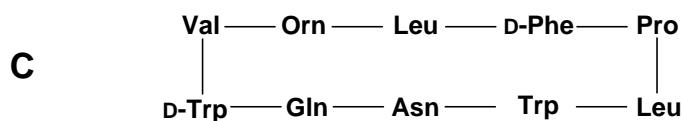
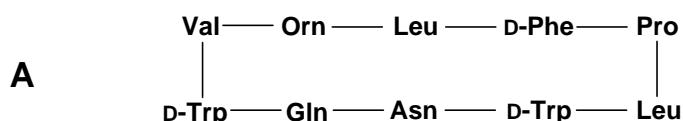


Habitat: terrestrial; Cape Coast, Ghana

Screening method: HPLC-DAD

Biological activity: antibacterial

Structures:



References:

Gebhardt, K.; R. Pukall & H.-P. Fiedler. J. Antibiotics 54: 428-433, 2001

Höltzel, A.; R.D. Süssmuth, R.W. Jack, G.J. Nicholson, K. Gebhardt, H.-P. Fiedler & G. Jung. J. Antibiotics 54: 434-440, 2001

Streptofactin

Producing organism: *Streptomyces tendae* Tü 901/8c

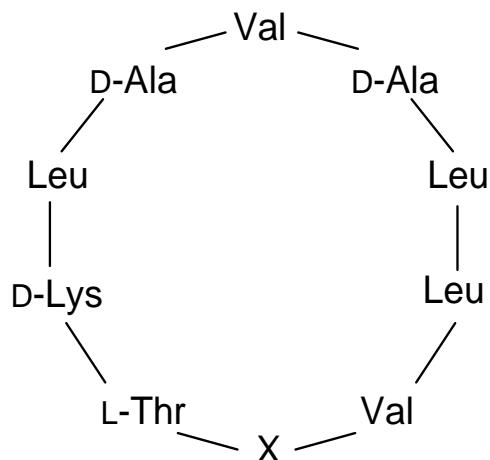


Habitat: terrestrial; Nikko, Japan

Screening method: complementation of bald mutants

Biological activity: induction of aerial mycelium

Structure:



Reference:

Richter, M.; J.M. Willey, R. Süssmuth, G. Jung & H.-P. Fiedler. FEMS Microbiol. Lett. 163: 165-171, 1998

Tetracenomycins B3, D3

Producing organism: *Streptomyces olivaceus* Tü 2353-R

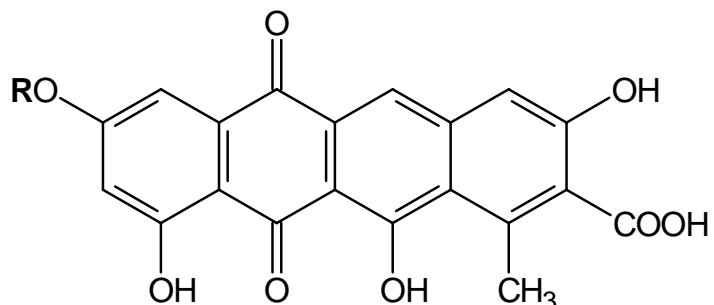


Habitat: terrestrial; Ellora, India

Screening method: HPLC-DAD

Biological activity: antibacterial (tetracenomycin D3)

Structures:



B₃ : R = CH₃

D₃ : R = H

Reference:

Rohr, J.; S. Eick, A. Zeeck, P. Reuschenbach, H. Zähner & H.-P. Fiedler. J. Antibiotics 41: 1066-1073, 1988

Tigloside

Producing organism: *Amycolatopsis* sp. NNO 21702

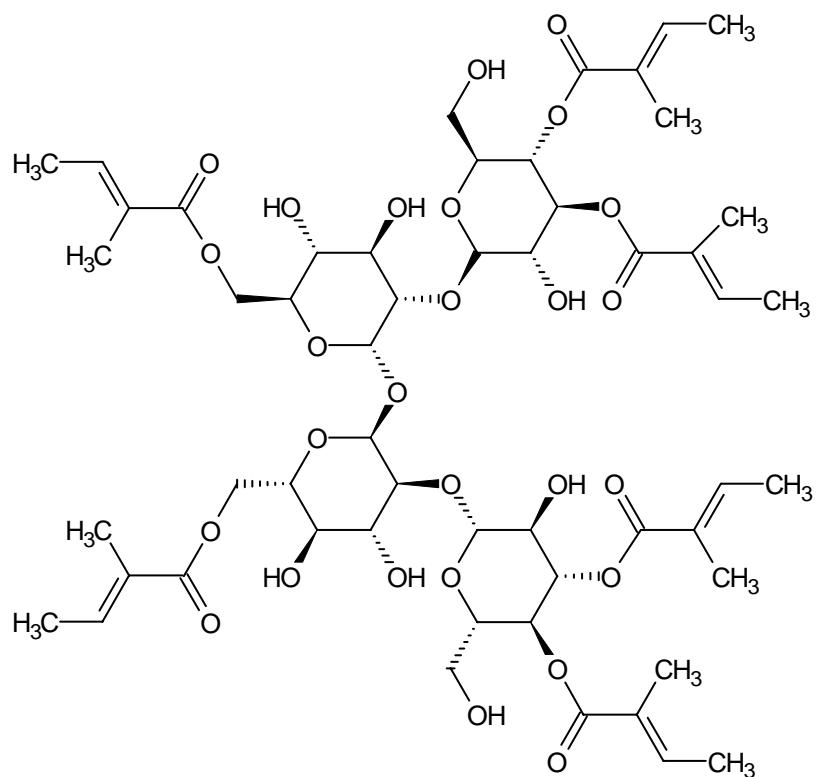


Habitat: terrestrial, Australia

Screening method: HPLC-DAD

Biological activity: none

Structure:



Reference:

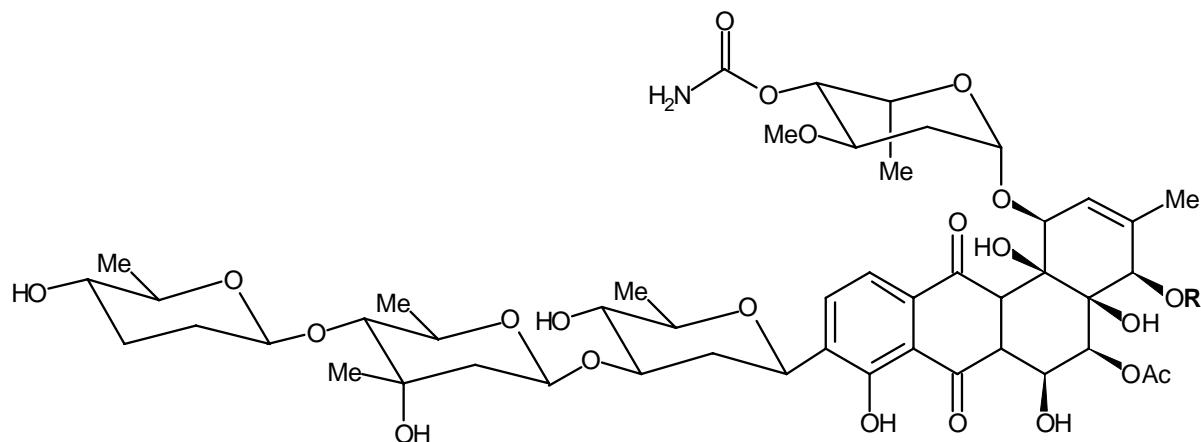
Breinholt, J.; A. Kulik, H. Gürtler & H.-P. Fiedler. Acta Chem. Scand. 52: 1239-1242, 1998

Warkmycin

Producing organism: *Streptomyces* sp. Acta 2930



- Habitat:** terrestrial; alkaline soil from sand dune, Warkworth, Northumberland, UK
- Screening method:** HPLC-DAD
- Biological activity:** antibacterial, antitumor
- Structures:**



Warkmycin: R = COMe

Deacetyl-Warkmycin: R = H

Reference:

Helaly, S.E.; M. Goodfellow, H. Zinecker, J.F. Imhoff, R.D. Süssmuth & H.-P. Fiedler. *J. Antibiotics*, in press 2013

III. Coworkers Involved in Metabolite Screening

My special thanks are due to my doctoral students who were involved in the screening for novel secondary metabolites and fermentation studies. Their names are in chronological order:

Willi Kuhn (1993)
Uwe Hartjen (1993)
Uwe Pfefferle (1994)
Sabine Blum (1995)
Christoph Pfefferle (1997)
Monika Richter (1997)
Judith Schimana (1999)
Markus Kempf (1999)
Klaus Gebhardt (2001)
Anke Dieter (2002)
Marcelo Bertasso (2002)
Noëmi Antal (2004)
Julia Riedlinger (2006)
Dirk Schulz (2009)
Nadine Horlacher (2012)
Marcell Wagner (2012)

Besides the doctoral students the following diploma students contributed to the progress in screening of secondary metabolites:

Thomas Fritz, Jan Mirko Gutterer, Nicole Zwicker, Armin Maier, Antje Seidel, Miriam Langbecker, Elisabeth Zürn, Andreas Reicke, Tatiana Binder, Stefanie Baur, Grit Hanschel, Bettina Wieser, Andreas Gierth, Claudia Hohmann, Sonja Gern, Philipp Jetter, Nicole Klüglein, Niko Manderscheid and Rebekka Jacek.

Without the support by my Post-Docs as the responsible persons, the various projects could not be realized. I appreciate in particular the work of Dr. Uwe Theobald, Dr. Johannes Müller, Dr. Judith Schimana and Dr. Christina Bruntner.

Last but not least, the assistance of the technical staff contributed greatly to the success of the research: Dip. Ing. Georg Grewe, Dipl. Ing. Andreas Kulik, Dipl. Ing. Jochen Schinko, Dipl. Biol. Barbara Holz, Mrs. Jutta Wachter, Mrs. Brigitte Maurer, Mr. Mulugeta Nega, Mr. Mete Bayraktar and Mr. Christian Tänzer.

IV. Collaborations and Fundings

National and international collaborations with colleagues from various universities and chemical-pharmaceutical companies were essential in the screening of microbial strains and determining the biological activities and structures of the isolated metabolites.

Professor Dr. Michael Goodfellow and Professor Dr. Alan C. Ward from the University of Newcastle, Newcastle upon Tyne, UK, provided us with alkaliphilic, acidophilic and thermophilic actinomycetes strains from terrestrial soils and fresh water sediments collected at various sites in the UK. Halophilic actinomycetes from Egypt and marine actinomycetes isolated from sediments of deep sea trenches and various regions of the North Atlantic completed the spectrum of extreme habitats. Full taxonomic characterization was done in Newcastle in the case of antibiotic producing strains.

Professor Dr. Alan T. Bull from University of Kent, Canterbury, UK, provided us with marine actinomycetes from sediments of the Pacific and Atlantic Ocean including deep sea trenches, with terrestrial actinomycetes from Antarctica and with actinomycetes isolated from the Atacama Desert in Chile.

Professor Dr. Vikineswary from the University of Malaya, Kuala Lumpur, Malaysia, provided us with a huge collection of new isolated actinomycetes strains from pristine tropical habitats in Malaysia.

Dr. Patricia S.M. Sommer from University of Rio Grande do Norte, Natal, Brasil, provided us with actinomycetes strains which were isolated from Brazilian medicinal plants.

Professor Dr. Konrad Dettner from University of Bayreuth, Germany, provided us with endosymbiotic actinomycetes and fungi, which were isolated from the guts of a huge diversity of arthropods, especially insects and millipedes.

Professor Dr. Erko Stackebrandt and Professor Dr. Rainer-Michael Kroppenstedt supported the taxonomic characterization of new actinomycete isolates and in the specialist training of some of my doctoral students in taxonomical techniques.

With Dr. Ingrid Groth from Hans-Knöll-Institut für Naturstoff-Forschung, Jena, we had a fruitful long-term collaboration in taxonomic characterization of new isolated actinomycetes, resulting in various common publications.

Professor Dr. Rüdiger Hampp from the Department of Physiological Ecology of Plants and Professor Dr. Karl Poralla from the Institute of Microbiology of the University of Tübingen provided us with streptomycetes acting as mycorrhiza helper bacteria.

Without chemical characterization of the isolated metabolites the whole research in screening for new biologically active secondary metabolites would be only patchwork. Therefore, my deepest thanks are due to the groups of Professor Dr. Axel Zeeck, Dr. Jürgen Rohr and Dr. Stephanie Grond and their teams from the Organic Chemistry

Department of the University of Göttingen (they moved recently to the Organic Chemistry Department of the University of Tübingen), Professor Dr. Günther Jung and his team from the Organic Chemistry Department of the University of Tübingen, Professor Dr. Gerhard Bringmann and his team from the Organic Chemistry Department of the University of Würzburg, Dr. Thomas Paululat from the Organic Chemistry Department of the University of Siegen, and last but not least my friend Professor Dr. Roderich Süßmuth from the Chemical Institute of the Technical University of Berlin, who started his career in Tübingen and moved in 2004 to Berlin. Roderich and his team of diploma and doctoral students were mainly involved during the last decade in structural elucidation of our isolated secondary metabolites. Without their enthusiasm for natural products our research could not be realized. But not only chemists from the various universities, also chemists from chemical-pharmaceutical companies contributed to the success of my group, as Dr. Jens Breinholt from Novo Nordisk, Denmark, Dr. Olivier Potterat and Dr. Carsten Puder from Boehringer-Ingelheim Pharma, Biberach, Germany.

Due to the limited facilities in determining the biological activities of the isolated new secondary metabolites, the collaboration in biological assays has been an essential part in my work. Professor Dr. Winfried Beil from the Medizinische Hochschule Hannover, Germany, performed antitumor assays to investigate activities against various human tumor cell lines. Professor Dr. Reto Brun from the Swiss Tropical Institute, Basel, took the part in screening for antiparasitic activities against various protozoan pathogens. Professor Dr. Johannes Imhoff from GEOMAR Helmholtz-Zentrum für Ozeanforschung Kiel, participated with testing our compounds in extended antibacterial, antifungal, antitumor and enzymatic assays.

Without financial support no reaserch would be possible. Therefore, my warmest thanks are due to following organisations and companies:

The Deutsche Forschungsgemeinschaft contributed with a basic support for fermentation and HPLC equipment within SFB 323 during the years 1988-1999.

The Bundesministerium für Forschung und Technologie (BMFT) Germany, supported our research in the optimisation of the biotechnological manufacturing of secondary metabolites within the ‘Zentrales Schwerpunktprojekt Bioverfahrenstechnik’ from 1988-98.

Novo Nordisk A/S, Bagsværd, Denmark, supported from 1995-1998 our research in secondary metabolites from the families *Streptosporangiaceae* and *Micromonosporaceae*.

The Bundesministerium für Bildung, Wissenschaft, Forschung und Technologie (BMBF) Germany together with BASF AG, Ludwigshafen, Germany, supported our research in secondary metabolites from endosymbiotic actinomycetes over a 6-year period from 1996-2001.

The European Commission supported our research within the 5th framework, project ACTAPHARM in the screening for new bioactive secondary metabolites from various actinomycetes genera from 2001-2004.

Boehringer Ingelheim Pharma GmbH, Biberach, Germany, supported our research in secondary metabolites from marine actinomycetes from 2002-2004.

The Deutsche Forschungsgemeinschaft contributed from 2002-2010 within 'Graduiertenkolleg Infektionsbiologie' with the support of doctoral theses to our research on the interaction of soil streptomycetes, mycorrhization and plants.

V. List of Publications

- Fiedler, E.; H.-P. Fiedler, A. Gerhard, W. Keller-Schierlein, W.A. König & H. Zähner: Synthese und Biosynthese substituierter Tryptanthrine. Arch. Microbiol. 107: 249-256, 1976.
- Fiedler, H.-P.: Isolierung von mikrobiellen Stoffwechselprodukten mit Hilfe von Adsorberharzen. Chem. Rundschau 30: 2-3, 1977.
- Fiedler, H.-P. & J. Sauerbier: Isolation and quantitative determination of siderochromes. Europ. J. Appl. Microbiol. Biotechnol. 5: 51-57, 1978.
- Hartmann, A.; H.-P. Fiedler & V. Braun: Uptake and conversion of the antibiotic albomycin by *Escherichia coli* K112. Europ. J. Biochem. 99: 517-524, 1979.
- Hagenmaier, H.; W.A. König, H. Zähner, H.-P. Fiedler, W. Dehler, A. Keckiesen, H. Holst & G. Zobelein: Neue Nikkomycine, ein Verfahren zu ihrer Herstellung, sowie ihre Verwendung als Schädlingsbekämpfungsmittel (1979). Europ. Patentschrift 0022 964 vom 9.2.1983, Anmeldetag: 4.7.1980, Offenlegungstag: 28.1.1981.
- König, W.A.; W. Hass, W. Dehler, H.-P. Fiedler & H. Zähner: Strukturaufklärung und Partialsynthese des Nucleosidantibiotikums Nikkomycin B. Liebigs Ann. Chem. 1980: 622-628, 1980.
- Hagenmaier, H.; A. Keckiesen, W. Dehler, H.-P. Fiedler, H. Zähner & W.A. König: Konstitutionsaufklärung der Nikkomycine I, J, N und M. Liebigs Ann. Chem. 1981: 1018-1024, 1981.
- Fiedler, H.-P.: Quantitation of nikkomycins in biological fluids by ion-pair reversed-phase high-performance liquid chromatography. J. Chromatogr. 204: 313-318, 1981.
- Fiedler, H.-P.: Preparative scale HPLC of ferricrocin, a microbial product. J. Chromatogr. 209: 103-106, 1981.
- Fiedler, H.-P.; W. Wörner, H. Zähner, H.P. Kaiser, A. Müller & W. Keller-Schierlein: Isolation and characterization of niphithricins A, B, and elaiophylin, antibiotics produced by *Streptomyces violaceoniger*. J. Antibiot. 34: 1107-1118, 1981.
- Fiedler, H.-P.; R. Kurth & W. Dehler: Nikkomycin quantitation in fermentation process by high-performance liquid chromatography. Biotechnol. Lett. 3: 303-308, 1981.
- Fiedler, H.-P.; R. Kurth, J. Langhärig, J. Delzer & H. Zähner: Nikkomycins, microbial inhibitors of the chitin synthase. J. Chem. Technol. Biotechnol. 32: 271-280, 1982.
- Delzer, J.; H.-P. Fiedler, H. Müller, H. Zähner, R. Rathmann, K. Ernst & W.A. König: New nikkomycins by mutasynthesis and directed fermentation. J. Antibiot. 37: 80-82, 1984.
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