

Bachelorthesis

Reconstruction of a genome-scale metabolic network of *Staphylococcus lugdunensis* N920143

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1 Background and Motivation

Staphylococcus lugdunensis is a Gram-positive and coagulase-negative human commensal [1]. Like all members of the staphylococcus genus, it is a spherical cell that forms clusters. It was first isolated and described by Freney et al. in 1988 and is named after the French city Lyon (lat. Lugdunum) [2]. *S. lugdunensis* can produce a non-ribosomally synthesized cyclic peptide antibiotic named lugdunin [3]. Lugdunin is effective against major pathogens, especially *Staphylococcus aureus*, which is known to cause aggressive infections in humans. It has been shown that nasal colonization by *S. lugdunensis* was associated with a significantly reduced *S. aureus* carriage rate. Thus, the use of *S. lugdunensis* as a probiotic to inhibit *S. aureus* in affected patients is being investigated [4]. However, this process is contraindicated by *S. lugdunensis* being an opportunistic pathogen with a high degree of virulence, unlike other Coagulase-Negative Staphylococcal species (CoNS) [5]. Although *S. lugdunensis* is a commensal and part of the normal skin flora, it has been found to cause aggressive infections similar to *S. aureus* [6]. These are often SSTIs, but the bacteria can also cause infections with high mortality, such as endocarditis [7].

The strain N920143 was isolated from a breast abscess in 1992 [5]. The genome consists of a chromosome of 2.6 Mb and no plasmids. Currently, there is no genome-scale metabolic model (GEM) for this strain, which makes the reconstruction of such a promising microbe of keen interest.

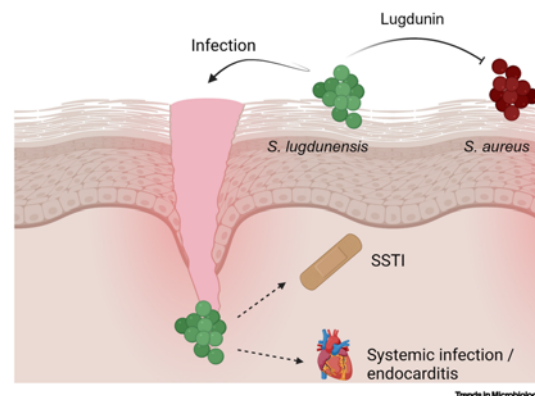


Figure 1 | *Staphylococcus lugdunensis* is found in the normal skin flora of humans. It can produce lugdunin and inhibit other bacteria like *Staphylococcus aureus*. But it also can cause skin- and soft-tissue infections (SSTIs) or more severe systemic infections (e.g., endocarditis). Image source: [https://www.cell.com/trends/microbiology/fulltext/S0966-842X\(21\)00184-0](https://www.cell.com/trends/microbiology/fulltext/S0966-842X(21)00184-0)

2 Aim and Approach

This thesis focuses on manually improving an automated draft of a genome-scale model. For the first draft, automated tools like CarveMe [8] will be used, which then will be refined using different databases and literature on *S. lugdunensis*. The aim is to create a high-quality systems biology model of *Staphylococcus lugdunensis* N920143. The quality will be assured by following the protocol for generating a high-quality genome-scale metabolic reconstruction [9]. The aim is to understand this unique bacteria's metabolic processes better and compare it to models of other strains (e.g., *Staphylococcus lugdunensis* HKU09-01). The model will be validated by testing the growth for different carbon and nitrogen sources, and testing the biomass yield.

3 Requirements

(a) Fundamental understanding of biochemistry, (b) interest in systems biology, particularly in constraint-based modeling, (c) Python programming using packages (e.g., COBRApy [10], libSBML [11]), and (d) interest in learning the usage of tools to improve the model gradually (e.g., CarveMe [8], MEMOTE [12]).

References

- [1] L. A. Heldt Manica and P. R. Cohen. “*Staphylococcus lugdunensis* infections of the skin and soft tissue: a case series and review”. In: *Dermatology and Therapy* 7.4 (2017), pp. 555–562. DOI: 10.1007/s13555-017-0202-5.
- [2] J. Freney, Y. Brun, M. Bes, H. Meugnier, F. Grimont, P. A. Grimont, C. Nervi, and J. Fleurette. “*Staphylococcus lugdunensis* sp. nov. and *Staphylococcus schleiferi* sp. nov., two species from human clinical specimens”. In: *International Journal of Systematic and Evolutionary Microbiology* 38.2 (1988), pp. 168–172. DOI: 10.1099/00207713-38-2-168.
- [3] A. Zipperer, M. C. Konnerth, C. Laux, et al. “Human commensals producing a novel antibiotic impair pathogen colonization”. In: *Nature* 535.7613 (2016), pp. 511–516. DOI: 10.1038/nature18634.
- [4] S. Heilbronner and T. J. Foster. “*Staphylococcus lugdunensis*: A skin commensal with invasive pathogenic potential”. In: *Clinical Microbiology Reviews* 34.2 (2021), e00205–20. DOI: 10.1128/CMR.00205-20.
- [5] S. Heilbronner, M. T. Holden, A. van Tonder, J. A. Geoghegan, T. J. Foster, J. Parkhill, and S. D. Bentley. “Genome sequence of *Staphylococcus lugdunensis* N920143 allows identification of putative colonization and virulence factors”. In: *FEMS Microbiology Letters* 322.1 (2011), pp. 60–67. DOI: 10.1111/j.1574-6968.2011.02339.x.
- [6] J. Lebeurre, S. Dahyot, S. Diene, A. Paulay, M. Aubourg, X. Argemi, J. Giard, I. Tournier, P. Francois, and M. Pestel-Caron. “Comparative genome analysis of *Staphylococcus lugdunensis* shows clonal complex-dependent diversity of the putative virulence factor, *ess*/type VII locus”. In: *Frontiers in Microbiology* 10 (2019), p. 2479. DOI: 10.3389/fmicb.2019.02479.
- [7] L. Bieber and G. Kahlmeter. “*Staphylococcus lugdunensis* in several niches of the normal skin flora”. In: *Clinical Microbiology and Infection* 16.4 (2010), pp. 385–388. DOI: 10.1111/j.1469-0691.2009.02813.x.
- [8] D. Machado, S. Andrejev, M. Tramontano, and K. R. Patil. “Fast automated reconstruction of genome-scale metabolic models for microbial species and communities”. In: *Nucleic Acids Research* 46.15 (2018), pp. 7542–7553. DOI: 10.1093/nar/gky537.
- [9] I. Thiele and B. Ø. Palsson. “A protocol for generating a high-quality genome-scale metabolic reconstruction”. In: *Nature Protocols* 5.1 (2010), pp. 93–121. DOI: 10.1038/nprot.2009.203.
- [10] A. Ebrahim, J. A. Lerman, B. O. Palsson, and D. R. Hyduke. “COBRApy: constraints-based reconstruction and analysis for python”. In: *BMC Systems Biology* 7.1 (2013), pp. 1–6. DOI: 10.1186/1752-0509-7-74.
- [11] B. J. Bornstein, S. M. Keating, A. Jouraku, and M. Hucka. “LibSBML: an API library for SBML”. In: *Bioinformatics* 24.6 (2008), pp. 880–881. DOI: 10.1093/bioinformatics/btn051.
- [12] C. Lieven, M. E. Beber, B. G. Olivier, et al. “MEMOTE for standardized genome-scale metabolic model testing”. In: *Nature Biotechnology* 38 (3 Mar. 2020), pp. 272–276. DOI: 10.1038/s41587-020-0446-y.