

Feature Review

Dual Neural Network Model for the Evolution of Speech and Language

Steffen R. Hage^{1,*} and Andreas Nieder^{2,*}

Explaining the evolution of speech and language poses one of the biggest challenges in biology. We propose a dual network model that posits a volitional articulatory motor network (VAMN) originating in the prefrontal cortex (PFC; including Broca's area) that cognitively controls vocal output of a phylogenetically conserved primary vocal motor network (PVMN) situated in subcortical structures. By comparing the connections between these two systems in human and nonhuman primate brains, we identify crucial biological preadaptations in monkeys for the emergence of a language system in humans. This model of language evolution explains the exclusiveness of non-verbal communication sounds (e.g., cries) in infants with an immature PFC, as well as the observed emergence of non-linguistic vocalizations in adults after frontal lobe pathologies.

The Apparent Discrepancy

Few questions in biology are as difficult and controversial as the evolution of human **speech** and **language**, and the emergence of essential speech and language brain structures such as Broca's area in the lateral frontal lobe. This is because human language vastly outperforms any primate communication system in scope and flexibility [1–3], with seemingly no counterpart in the animal kingdom, even among hominids. The vocalizations of nonhuman primates are largely innate, stereotypic, and were thought to be almost exclusively uttered affectively (Box 1) [4,5]. Humans, on the other hand, learn speech sounds, use them flexibly in combinatorial symbol systems, and can volitionally control their utterances.

This apparent discrepancy between monkey vocalization and human language is also reflected in traditionally disparate research agendas. On the one hand, neurobiologists have meticulously deciphered the neural pathways causing nonverbal vocal output in monkeys. This body of work has identified different numbers of primarily subcortical pathways that allow nonhuman primates to produce innate vocalizations. However, cortical association areas that are of paramount importance to human language production, such as Broca's area in the inferior frontal lobe, have not been assumed to play a part in it. Cognitive neuroscientists, on the other hand, usually focus on the linguistic **articulation** network and strive to understand how Broca's area allows humans to structure **semantic** verbal expressions. How the language executive associated with Broca's area becomes coupled to the necessary vocal machinery in the brainstem and how it can emerge throughout evolution has largely been neglected. Occasionally, links between Broca's area and the primate vocal pathways have been suspected but remained speculative as a consequence of lacunae in neurobiological data [5–8].

This review tries to bridge this gap by integrating novel insights about vocalizations in monkeys as well as verbal and nonverbal output in humans. We review recent behavioral, anatomical, and

Trends

A dual-network model for the evolution of language is proposed that consists of two interacting brain networks.

A phylogenetically conserved PVMN produces genetically predetermined vocalizations in nonhuman primates and non-verbal vocalizations in humans.

During the course of primate evolution, an additional VAMN in the lateral frontal lobe (including Broca's area) emerges that cognitively controls vocal output.

In humans, the VAMN gains control over articulation by modulating the output of the PVMN.

The new VAMN also plays a vital role in establishing semantics and syntax, two hallmark characteristics of symbol systems in humans.

¹Neurobiology of Vocal Communication, Werner Reichardt Centre for Integrative Neuroscience, University of Tübingen, Otfried-Müller-Strasse 25, 72076 Tübingen, Germany
²Animal Physiology Unit, Institute of Neurobiology, University of Tübingen, Auf der Morgenstelle 28, 72076 Tübingen, Germany

*Correspondence: steffen.hage@uni-tuebingen.de (S.R. Hage) and andreas.nieder@uni-tuebingen.de (A. Nieder).

Box 1. Monkey Vocalization

While human speech is above all a learned vocal pattern, the vocal motor system of non-human primates consists mainly of stereotyped and largely innate calls that are almost exclusively uttered affectively [2,5]. These assumptions are mainly bolstered by so-called ‘Kaspar Hauser’ experiments as well as by deafening and lesion studies. In these experiments, squirrel monkeys were raised without auditory feedback from conspecifics, by either hand-raising them or by muting the mother. These monkeys still produced the complete species-specific vocal repertoire and used it in the appropriate motivational and social contexts [177,178]. Deaf-born or deafened monkeys showed similar call patterns as their normal-hearing conspecifics, with only minor changes in call frequency and amplitude [178,179]. Finally, several studies in Old World and New World monkeys observed no differences in vocal pattern production after lesioning brain structures that are homologous to the brain regions that are crucial for speech production in humans [18,35–37].

However, several behavioral studies report that monkeys are able to **volitionally initiate vocal output** and instrumentalize their calls in a goal-directed (adaptive) way. Non-human primates are able to produce a vocalization or remain silent when submitted to operant conditioning tasks [65,76,84,180–183]. These studies support field studies showing that non-human primates vocalize in different ways when addressing different individuals [184], and produce or withhold alarm calls depending on the social context [185]. Therefore, calls might encode the vocalizing individual's information about the presence of a predator [185], the behavior of other individuals [186], or specific external events [187]. In a recent study we demonstrated that rhesus monkeys are able to selectively emit different call types in response to distinct visual cues [86]. Furthermore, several studies observed volitional changes of vocal parameters such as vocal duration, amplitude, and frequency within the natural constraints [33,84–86]. These results indicate that monkeys have rudimentary control over specific acoustic call parameters. In addition, recent work on highly-vocal marmoset monkeys has revealed dynamics in auditory perception of vocalizations [188], vocal flexibility [189], and some evidence of auditory comprehension learning [190,191]. Overall, these results indicate that monkeys are capable of some types of vocal learning, in other words they are able to cognitively control the onset of their vocal output and to modify their vocal patterns within the range of their natural repertoire.

physiological findings suggesting that monkeys possess the rudiments to cognitively control their vocal output so as to develop a neurobiological scaffold for language evolution in primates. Cognitive or executive control – the ability to orchestrate thought and action in accordance with internal goals [9] – is an obligatory precursor for speech and language production. Over the course of primate evolution, executive control structures residing in the PFC gradually became coupled to ancient vocal pattern-generating and **limbic networks**. By comparing and contrasting the corticocortical and corticosubcortical connections relevant for human speech and language production with those present in nonhuman primates for vocalizations, we identify crucial biological preadaptations for the emergence of a full-blown language system. Based on these findings, we suggest a dual-network model of speech and language evolution that accounts for the observation that the humble beginnings of the structural and functional evolution of a cortical speech and language system can already be observed in nonhuman primates. This model integrates seemingly disparate neurobiological findings in human and nonhuman primates and attempts to provide a conceptual framework of how speech and language might have arisen during primate evolution.

The PVMN for Innate Primate Vocalizations

All primates possess a PVMN that produces genetically predetermined vocalizations in nonhuman primates and non-verbal vocalizations in humans. This PVMN consists of two structurally and functionally distinct parts: a vocal pattern-generating system in the brainstem, and an upstream limbic vocal-initiating network driving the **pattern generator** based on affective states (Figure 1A,B) [4,5,10].

The core of the PVMN, the brainstem vocal pattern-generating system is highly conserved in all vertebrate taxa [11], and mainly consists of the interconnected areas periaqueductal grey (PAG), parabrachial nucleus (PB), and ventrolateral pontine reticular formation; the latter controls all phonatory motoneuron pools (nuclei of cranial nerves V, VII, XII; ambigular nucleus) which, in addition to controlling the musculature of the respiratory system, also innervate the muscles of the larynx, oral cavity, and tongue to produce the actual vocal sound (Figure 1A,B) [5,12–16].

Glossary

Articulation: complex movements of the cranial muscles (facial, lip, tongue, jaw) to alter a basic tone, which is produced by the vocal folds, into decodable sounds.

Grammar: rules that govern how words can be combined to form sentences.

Language: a generative symbol-system to generate infinite meanings (semantics) based on a finite set of rules (syntax) (and usually is used to communicate).

Limbic network: complex network of brain structures including ACC, hypothalamus, amygdala and several other structures that seem to be primarily responsible for emotion and that participate in memory formation and learning.

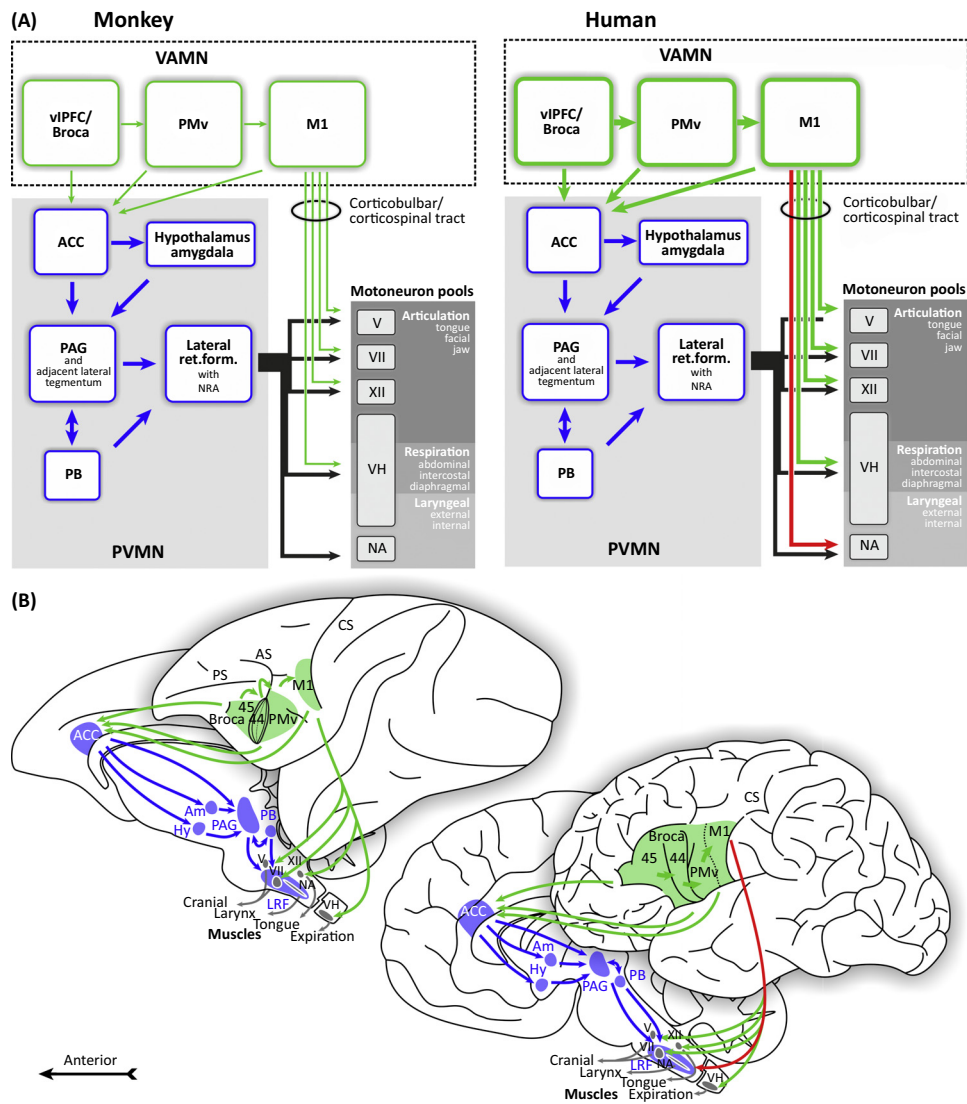
Motor/central pattern generator: a neuronal circuit that produces motor commands such as vocalizations. Pattern generators produce distinct outputs even in isolation from motor and sensory feedback.

Pyramidal tract: direct projections from neurons in the primary motor cortex to interneurons and motoneurons in the brainstem (corticobulbar tract) or the spinal cord (corticospinal tract).

Semantic: meaning of linguistic expressions.

Speech: human speech is the ability to cognitively control and produce a series of arbitrary complex articulatory movements.

Volitional vocal initiation: ability to vocalize in response to abstract, learned sensory stimuli in a goal-directed manner.



Trends in Neurosciences

Figure 1. Dual-Network Model. (A) Simplified circuit diagram summarizing the most relevant structures for vocal production in monkeys and speech in humans. Arrows indicate anatomically verified and relevant direct connections. The VAMN (indicated in green) is capable of initiating and modulating vocal production in monkeys during cognitive control of vocal onset, or the modulation of vocal patterns within natural constraints. These connections are enhanced during speech evolution in the primate lineage. (B) Anatomical locations and connections of the structures comprising the dual-network in monkeys (left) and humans (right). Lateral (front) and medial (back) views of the endbrain hemispheres are shown. Regions and arrows shaded in blue indicate the structures of the PVMN. Regions and arrows shaded in green depict cortical areas involved in the VAMN. Red arrow indicates the direct connection between the larynx area of the primary motor cortex with the ambigular nucleus as a new development in the human lineage (adapted from [4,67,200]). Abbreviations: Am, amygdala; ACC, anterior cingulate cortex; AS, arcuate sulcus; CS, central sulcus; Hy, hypothalamus; LRF, lateral reticular formation; M1, ventral primary motor cortex; NA, ambigular nucleus; NRA, retroambigular nucleus; PAG, periaqueductal grey; PB, parabrachial nucleus; PMv, ventral premotor cortex; PS, principal sulcus; PVMN, primary vocal motor network; VH, respiratory motoneuron pools in the ventral horn of the spinal tract; V, motor trigeminal nucleus; VAMN, volitional articulatory motor network; VII, facial nucleus; XII, hypoglossal nucleus.

The pivotal role of these brainstem structures has been deciphered based on electrophysiological recording, stimulation, and lesion studies in monkeys. In all of these brainstem areas, neurons show vocalization-related neuronal activity [13,15,17–19]. Electrical and chemical stimulation of the PAG reliably elicits distinct calls of the complete species-specific vocal repertoire in monkeys [20,21], whereas lesioning PAG results in mutism [22,23]. In contrast to the more general (or global) function of the PAG in producing calls, the PB is involved in the gating of vocal onset on

the basis of the momentary respiratory status [5] because electrical stimulation of the PB produces only simple vocalizations [20,24], and PB lesions uncoupled respiratory rhythms from laryngeal activity during vocalizations elicited via PAG stimulation [23].

This brainstem vocal pattern-generating system is controlled by a limbic vocal-initiating network that projects to the PAG and the entire PVMN to elicit vocalizations representing affective states. Its most important structures comprise a vocalization region in the anterior cingulate cortex (ACC) around the rostrum of the corpus callosum (including parts of areas 24, 25, and 32), the hypothalamus, other limbic diencephalic structures (such as the septum and the subcallosal gyrus), and the amygdala [23,25,26]. Electrical stimulation in all of these limbic structures including ACC elicits vocal utterances [20,27–29]. However, vocal latencies relative to stimulation onset were much longer (>1 s) compared to response latencies in PAG (<1 s) [20], suggesting that ACC and other limbic structures are coupled to, but are not part of, the pattern generator. In contrast to PAG stimulation, stimulation of most limbic structures could only elicit a subset of call types of the species-specific call repertoire. For example, vocalization with hedonistic quality can be elicited in the septum, while stimulation sites in the amygdala elicited more-aversive vocal utterances [20,30,31].

Bilateral ablations of the cingulate vocalization region have either no consistent effect on spontaneous vocal behavior [32–34], and lead to a decrease in spontaneous vocalization rate [35] or to calls weakened in amplitude and duration [36]. In all cases, however, animals were still able to spontaneously phonate after ACC lesions. In contrast to spontaneous calls, discriminatively conditioned vocal behavior was disrupted in monkeys with such bilateral lesions [35,36]. Ablations of other limbic structure such as amygdala and hypothalamus suppress distinct spontaneously uttered vocalizations, but do not abolish calls elicited by stimulating PAG [23,37].

These findings strongly suggest that the limbic vocal-initiating network is not involved in the production of vocal patterns themselves, but governs the affective (emotional or motivational) initiation of the vocal output.

The Role of the PVMN in Human Non-Verbal Vocalizations and Speech

It is important to realize that the phylogenetically conserved PVMN is still involved in vocalization in humans. One of its functions is to produce non-verbal vocal utterances such as crying, laughing, or moaning, all of which are innately predetermined and affective vocalizations considered to be directly homologous to monkey vocalizations [4,5]. Although brainstem lesions are often fatal, a clinical study revealed pathological laughter and crying due to a tumor beneath the brainstem that most likely deteriorated networks within the brainstem [38].

Several clinical studies also implicate the ACC in the initiation of non-verbal vocal utterances. In a type of frontal lobe epilepsy characterized by involuntary and stereotyped bursts of laughter ('gelastic seizures' [39]), the cingulate gyrus appears to be the most commonly disrupted site [40]. In agreement with this idea, electrical stimulation of the rostral ACC (and the hypothalamus) elicited uncontrollable, but natural-sounding laughter [39,41,42]. At the same time, ACC is important for speech. In humans, bilateral infarction of the ACC near the rostrum of the corpus callosum results in akinetic mutism [43,44], but with the potential for restoration of speech characterized by monotonous intonation, suggesting that the ACC is involved in the emotional intonation of human speech [37,45]. This idea is supported by recent imaging studies showing ACC activation during anger-expression of human speech [46]. Another important function of the PVMN is vocal output during speech production. Midbrain areas such as the PAG play a significant role in vocal production in humans, and its lesions can cause akinetic mutism [47,48]. In a case report, lesions in the PAG resulted in mutism, in other words it caused not only the absence of all non-verbal vocal utterances, but also the absence of speech [49]. Interestingly,

this patient was still able to produce the articulatory movements that are accompanied by a specific speech pattern, but the tonal component was no longer elicited [49]. However, PAG is active during the expression of non-verbal vocal utterances such as laughter in humans [50]. Moreover, a PET study showed that PAG is functionally coupled to a wide array of regions during voiced speech, but not during whispered speech [51].

Collectively, patient studies indicate that the PVMN is responsible for eliciting non-verbal affective vocalization but, in addition, plays a major role in speech production at the expense of affective vocalizations. Because the PVMN cannot subservise both affective preprogrammed vocalizations and volitional speech articulation at the same time, the evolution of speech and language may have required the taking-over and dissolution of the majority of preprogrammed vocalization patterns [52]. Potentially recapitulating such an evolutionary scenario during ontogeny, both laughter and crying constitute extremely important forms of communication in human infancy, bridging the gap between a prelinguistic stage to the later stage of speech and language acquisition. The advent of speech and language in Hominini therefore predicts the emergence of a new neocortical cognitive control network that occupies the ancient brainstem vocalization circuits through a VAMN. The neurobiological foundations for this radical reorganization are visible in nonhuman primates.

Volitional Articulatory Motor Network

Humans possess an additional VAMN consisting of cortical structures crucial for human speech control, and which is already present, although being anatomically and functionally underdeveloped, in the monkey brain. The VAMN comprises the inferior frontal gyrus (IFG), the caudally bordering ventral premotor cortex (area 6, PMv), and ventrolateral primary motor cortex (area 4, M1) including the facial and laryngeal motor cortex (Figure 1A,B). The central executive of this network is Broca's area, located in the IFG of the granular ventrolateral PFC (vIPFC). The granular PFC was added newly to the anterior pole of the frontal lobe during the evolution of primates [53,54].

Broca's area classically comprises cytoarchitectonic areas 44 (pars opercularis) and 45 (pars triangularis) in the left hemispheres, complemented by some authors by area 47 (pars orbitalis) [55]. Broca's pioneering work on brains of aphasics [56] revealed that areas 44 and 45 on the left side of the brain are instrumental for the production, or articulation, of speech and language. Studies on the cytoarchitectonics of macaque brains identified homologs of area 45 on the posterior convexity of the vIPFC, and of area 44 in the fundus of the inferior limb of the arcuate sulcus, a landmark that separates the PFC from the PMv (Figure 1B) [57,58]. No direct connections exist between the vIPFC and the primary motor cortex, but there are extensive projections to adjacent PMv that, in turn, send projections to primary motor cortex and the spinal cord. Specifically, area 44 is connected with the anterior PMv at the convexity of the inferior arcuate sulcus. The anterior PMv integrates sensorimotor signals of the posterior parietal cortex with cognitive information originating from vIPFC, pre-SMA, and cingulate area 24, the latter being in turn connected with the vIPFC [59]. Information in anterior PMv is then broadcast to the adjacent areas of the PMv and finally to ventrolateral M1, which also integrates laryngeal sensorimotor information [60] for the generation and control of face/mouth movements [61]. Neurons in both the PMv and M1 have direct access to the spinal cord, and thus can influence the generation and control of speech-related movements [62].

While the motor cortex of non-primate mammals is connected with the phonatory motor nuclei via interneurons in the reticular formation, only primates show direct connections via the corticobulbar and corticospinal tracts, respectively [63]. These connections give both New World and Old World monkeys the capability to volitionally control their articulatory muscles to respond with licking at a feeding tube in an operant conditioning task [64], and to

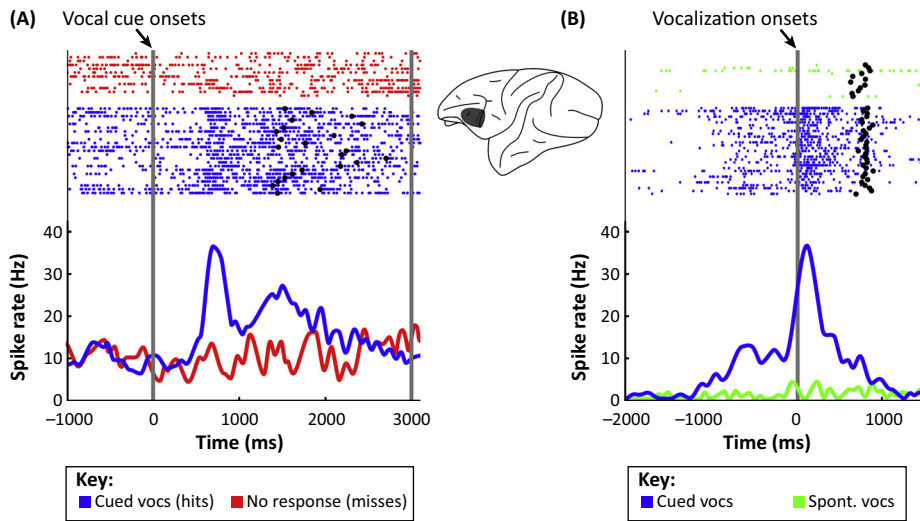
mimic articulatory vocal movements in a detection task [65], respectively. In humans only, additional direct connections of the motor cortex with the ambiguous nucleus seem to exist [66] (Figure 1A,B), and humans also possess direct projections into the anterior horn of the thoracic and upper lumbar cord (reviewed in [5]). The motor cortex therefore plays a crucial role within the VAMN in humans [67]. Lesions in the human ventral motor cortex, which controls cranial and laryngeal muscles via the corticobulbar tract, lead to severe speech impairments, while the production of innate non-verbal vocal utterances such as laughing and crying remains intact [68].

Despite the crucial role of Broca's area in human speech and language production, neither electrical stimulation nor lesions in homologous areas of monkeys have significant effects on spontaneous vocalizations [5]. For example, bilateral destruction of the 'cortical face area', including the premotor, motor, and sensory representation of the jaw muscles, lips, tongue, velum, and larynx, does not affect the acoustic structure of vocalizations in either squirrel or macaque monkeys [18,32,69]. Similarly, ablation of lateral frontal areas in macaques does not affect vocalizations [35,36]. Moreover, electrical stimulation in the motor cortex of monkeys does not elicit vocal output [70,71], and vocal utterances cannot reproducibly be elicited by stimulating the premotor and motor cortex of anesthetized chimpanzees [72–74]. These findings were taken as evidence that the lateral frontal lobe areas do not play any role in monkey vocalizations. If true, this would constitute an apparent discontinuity between the human speech system and the monkey vocalization network.

Recent experiments in nonhuman primates, however, showed that the monkey homolog of Broca's area, as well as the premotor and/or primary motor cortices, are all involved in the initiation of volitional calls that have been uttered in response to visual or auditory stimuli, respectively [65,75–77]. Vocalization-related activity that specifically predicts the preparation of instructed vocalizations in response to the detection of arbitrary visual stimuli was recorded in monkey vIPFC (areas 44 and 45) [76] and PMv [65,76] (Figure 2). Moreover, the activity of many call-related neurons before vocal output correlated with call parameters of instructed vocalizations. Furthermore, neuronal responses during conditioned vocalizations were higher than during spontaneous vocalizations, suggesting a specific involvement of the monkey homolog of Broca's area during volitional monkey vocalizations (Figure 2). At first sight, these results seem to be contradictory to earlier studies showing that bilateral ablation of ventrolateral aspects of the frontal lobe have no significant impact on discriminatively conditioned vocal behavior [36]. However, the exact positions of areas 44 and 45 were anatomically not verified until recently [57,78]. Therefore, the most likely explanation for this discrepancy is that experimental lesions in the previous studies have not included the full extent or even the bulk of the monkey vIPFC, PMv, and M1. In all lesioned monkeys, ablations left areas 6vr, 44, 45, and M1 at least partially intact in one hemisphere (see Figure 1 in [36]).

Based on recent results and neuroanatomical projections, the vIPFC might take control over the vocal motor network via ACC or alternatively via the premotor cortex that has direct input to motor cortex [79,80]. Because electrical microstimulation in area 44 and the PMv elicited orofacial and laryngeal responses (Figure 3) [65,79,81–83], the pathway leading from vIPFC via PMv to M1 to the corticobulbar and corticospinal tract to control phonatory motor neurons seems more likely.

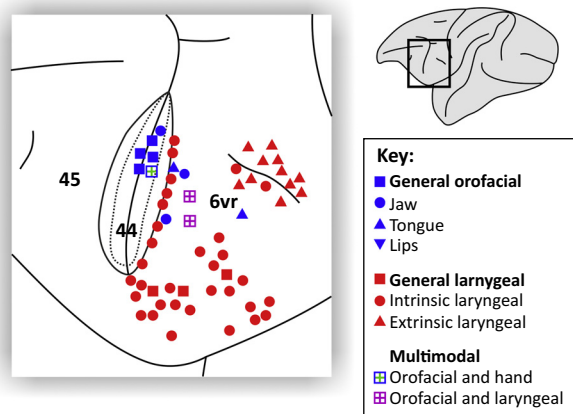
An influence via the VAMN could explain the rudimentary capability of monkeys to change vocal parameters such as vocal duration, amplitude, and frequency during cognitive vocal behavior [84–86]. Amplitude and frequency modulations can be generated by increasing the expiratory air flow passing the vocal folds in the larynx. This modulation might be solely controlled via direct



Trends in Neurosciences

Figure 2. Activity Related to the Preparation of Vocalizations of Single Neurons in the Monkey Homolog of Human Broca's Area. (Inset) Lateral view of a macaque brain indicating the position of the ventrolateral prefrontal cortex (vlPFC). (A) Example neuron recorded in the monkey homolog of human Broca's area showing a significant increase in neuronal activity during trials with cued vocalizations (hits) in comparison to no response trials (misses). (B) Example neuron recorded in the monkey homolog of human Broca's area showing significantly higher activity before vocal onset of volitional coo calls (cued vocs) compared to spontaneous coo calls (spont. vocs). (A,B) Upper panels show raster plots, black dots indicate vocal onset during hit trials in (A) and mark call offsets in (B); lower panels represent the corresponding spike-density histograms averaged and smoothed for illustration. The vertical grey lines indicate the onset and offset of the go-signal in (A) and vocal onset in (B) (adapted from [76]).

pyramidal projections from the motor cortex to the respiratory motoneuron pools within the ventral horn of the spinal tract [87]. Distinct changes in call frequency – independently of call amplitude – might be modulated by indirect projections of the laryngeal motor cortex to the ambiguus nucleus via corticofugal projections to the reticular formation of the lower brainstem [81,88,89]. For changes in call duration, modulation of the PVMN is necessary.



Trends in Neurosciences

Figure 3. Representation of Electrical Stimulation Sites in Monkey Areas 6vr and 44 Which Elicited Orofacial [General (i.e., Not Defined in Greater Detail), Jaw, Tongue, or Lip Movements], Laryngeal [General (i.e., Not Defined in Greater Detail), Intrinsic Laryngeal or Extrinsic Laryngeal] or Multimodal Responses (Orofacial and Hand Movements or Orofacial and Laryngeal Movements at the Same Penetration Site). The inferior arcuate sulcus is unfolded to show the location of area 44 within the sulcus. Stimulation sites taken from [65,79,82,83].

Brain imaging evidence suggests that the mechanism of how the lateral frontal lobe gains control over articulation is not a direct excitation of the phonatory motor neurons in the brainstem, but is instead a disinhibition of articulatory muscle activity briefly before vocal output [90]. This line of argumentation would predict that non-verbal, emotional vocalizations might emerge once the modulatory (and/or inhibitory) influence of the voluntary articulation network vanishes [91]. Indeed, non-verbal vocal utterances remain intact despite devastating impairments in speech and language production (Broca's aphasia) after damage to the posterior IFG [56,68,92,93]. Moreover, patients with a clinical diagnosis of primary progressive aphasia develop abnormal laughter-like vocalizations that increasingly replace speech in the context of progressive speech-output impairment leading to mutism, until ultimately laughter-like vocalizations are the only extended utterance produced by these patients [94]. Interestingly, some non-verbal vocal utterances occur more often during the conversation of aphasic than of nonaphasic adults [95], suggesting a competitive modulatory influence of the VAMN over the PVMN.

In agreement with such a modulatory, possibly inhibitory, effect of the VAMN is the classic finding that electrical stimulation of the Broca's area leads to speech arrest rather than to speech production [96–98]. Direct cortical recordings revealed that Broca's area is predominantly activated before the utterance of a speech signal, but is silent during the corresponding articulation [99], again indicating that Broca's area is indirectly involved in coordinating speech initiation rather than in producing speech output directly [93,100]. These findings are supported by recent findings in awake neurosurgical patients in which the activity of distinct cortical speech sites was focally decreased via cooling [101]. Cooling of Broca's area predominantly altered speech timing, again indicating an involvement of Broca's area in speech coordination rather than in direct speech production. By contrast, focal lowering of temperature in speech motor cortex led to modulation of articulation, confirming the direct role of the speech motor cortex in articulatory motor control. The recent cognitive control signals found in the monkey lateral frontal lobe, in combination with the modulatory function of the VAMN and the emergence of non-linguistic vocalizations after its damage in humans, suggest that the humble beginnings of speech control can already be witnessed in nonhuman primates. From there, they seemed to have evolved into a full-blown language-production system in humans after expansion of PFC circuitry (Box 2).

The PFC Receives and Classifies Communicative Signals

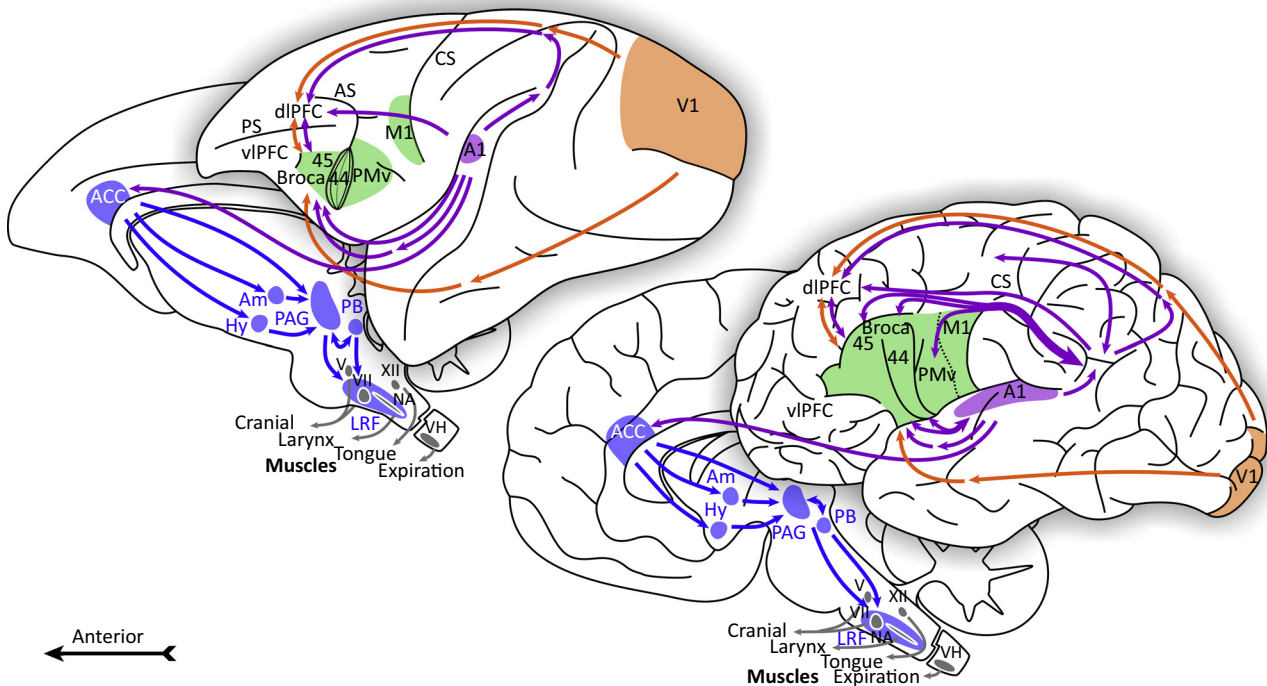
For a cortical network to control vocal output during reciprocal communication acts, sensory input from a sender is required. The vIPFC as the core of the VAMN receives highly processed information from higher-order sensory areas of all modalities (Figure 4). Neurons in the vIPFC categorize and maintain communicative signals in working memory to guide goal-directed output.

Auditory information reaches the lateral PFC via two largely anatomically and functionally segregated cortical streams: the anteroventral and the posterodorsal stream originating from the primary auditory cortex of the temporal lobe [102]. In the posterodorsal stream, the posterior auditory belt and parabelt areas project directly to the dorsolateral PFC (dlPFC; areas 8, 46, and 9) [102,103] (Figure 4). In addition, the posterior belt, parabelt, and superior temporal regions project indirectly to the posterior parietal association cortex before projecting to the dlPFC and principal sulcus (BA 46). This stream is thought to primarily encode auditory space [102] and quantity [104]. In the anteroventral stream, on the other hand, both the anterior belt and anterior parabelt regions project directly and reciprocally to vIPFC (areas 12/47, 45, 44, and 12 orbital) [103,105]. In addition to this direct projection, there is also an indirect projection to the PFC via the temporal association cortices. The anterior belt and parabelt regions are connected with the rostral (TS1, TS2) superior temporal gyrus (STG) that contains a 'voice region' in which neurons respond preferentially to monkey calls [106–108]. The STG in turn projects to relatively restricted

Box 2. The Neoteny Hypothesis of Cognitive Vocal Control for Language

How might the expansion of the VAMN have emerged in human primates over the course of evolution? One of the most powerful mechanisms evolution can use to remodel organs with very few genetic changes are delays in the timing of developmental stages [192]. A recent longitudinal behavioral study with monkeys trained to vocalize on command in response to a visual cue in a self-initiated trial suggests that retention of juvenile features, termed neoteny, might have been a key developmental event. Monkeys that laboriously master cued vocalizations during their juvenile phase gradually show more and more difficulties in producing such volitional calls with age, until they entirely discontinue controlled vocal behavior during adulthood [193]. This emerging disability was confined to volitional vocal production because the monkeys continued to vocalize spontaneously and continued to use hand movements as instructed responses during cognitive tasks in adulthood.

Because the decline in volitional call behavior correlated with the transition of the monkeys from juvenile phases to adulthood, these findings argue for a maturation process that might specifically affect the PFC. By prolonging development, greater neural plasticity early in ontogeny can be exploited to foster high-level cognition including speech and language [194,195]. In other words, linguistic capabilities may have been enabled because of an expansion of the juvenile period during the development of humans. This is supported by the finding that the PFC which contains Broca's language area experiences extraordinary long phases of developmental reorganization of neuronal circuits [196]. Genes related to the development of the PFC show excessive neotenic expression in humans relative to chimpanzees and rhesus macaques [197]. As a consequence, excessive synaptic connections and dendritic spines are initially over-produced to about twice the adult number before being pruned during puberty to reach the adult level at the onset of adolescence [196,198,199]. This pruning process might decouple the juvenile connection between the PFC and the PVMN during adolescence in monkeys. This points to non-activity-related developmental reorganization in the brain of maturing monkeys, rendering adult monkeys unable to bring vocalizations under cognitive control. This hypothesis suggests that expansion of the juvenile period during ontogeny may be one of the key events in the evolution of speech and language.



Trends in Neurosciences

Figure 4. Anatomical Locations and Connections of Sensory Structures That Provide Input to the Vocalization/Speech Systems in Monkeys and Humans. Regions and arrows shaded in blue indicate the structures of the PVMN. Regions shaded in green depict cortical areas comprising the VAMN. Regions and arrows shaded in orange show the interaction with auditory structures. Regions and arrows shaded in orange indicate the connections from the visual system (adapted from [4,67,102,200]). Abbreviations: A1, primary auditory cortex; ACC, anterior cingulate cortex; Am, amygdala; AS, arcuate sulcus; CS, central sulcus; dIPFC, dorsolateral prefrontal cortex; Hy, hypothalamus; LRF, lateral reticular formation; M1, ventral primary motor cortex; NA, ambigular nucleus; PAG, periaqueductal grey; PB, parabrachial nucleus; PMv, ventral premotor cortex; PS, principal sulcus; PVMN, primary vocal motor network; V1, primary visual cortex; VH, respiratory motoneuron pools in the ventral horn of the spinal tract; viPFC, ventrolateral prefrontal cortex; V, motor trigeminal nucleus; VAMN, volitional articulatory motor network; VII, facial nucleus; XII, hypoglossal nucleus.

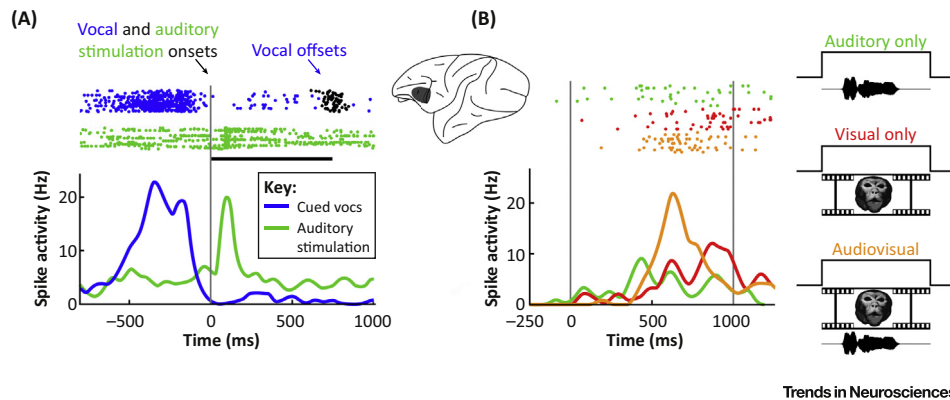


Figure 5. Audio-Vocal and Multisensory Activity in Single Neurons of the Monkey Ventrolateral Prefrontal Cortex (vIPFC). (Inset) Lateral view of a macaque brain indicating the position of the vIPFC. (A) Example neuron recorded in the monkey homolog of human Broca's area showing a phasic response during auditory stimulation and increased neuronal activity before volitional calls (cued vocs) (from [111]). (B) Example neuron showing nonlinear activity in response to auditory (vocalization), visual (silent movie), and audiovisual stimuli (congruent face-vocalization movie) demonstrating a significant interaction between auditory and visual stimuli, and exhibiting multisensory enhancement (adapted from [126]). Upper panels show raster plots (A,B); black dots indicate vocal onset during hit trials in (A), and lower panels represent the corresponding spike-density histograms averaged and smoothed for illustration. The vertical grey lines indicate the onset of the go-signal in (A), and the onset and offset of the vocalization, the silent movie, and the face-vocalization movies (B).

clusters of the vIPFC. The anteroventral stream is thought to encode auditory identity. Patches of single neurons in vIPFC respond robustly to complex sounds such as species-specific vocalizations or human vocalizations [109–111] (Figure 5A), and to some categories of vocalization calls [112]. In addition, dense auditory projections also exist from the rostral STG to the medial PFC, specifically to areas 32 and 25 of the ACC [113].

These separate projection streams in the auditory cortex are reminiscent of the temporal 'what' and dorsal 'where' visual pathways [114], or 'perception–action' pathways, respectively [115]. In the temporal visual pathway, higher-order representations of objects are found in the termination zone of the inferior temporal cortex where neurons encode specific object categories such as faces [116,117]. The downstream projections from IT cortex give rise to patches of face-selective neurons in the IPFC [118,119].

The largely segregated visual and auditory pathways converge in the vIPFC to give rise to neurons that represent higher-order multisensory and categorical representations of communicative signals. In line with evidence that vIPFC neurons represent perceptual and abstract categories [120,121], neuronal responses in this region correlate with monkeys' choices in an auditory same–different task [122] and during categorization of human speech sounds [123]. Supporting the integration of modal communication channels, neurons in vIPFC also respond to particular face–voice combinations [124–126] (Figure 5B). In addition, recent recording and inactivation studies in monkeys showed that the vIPFC is particularly important for processing behaviorally-relevant stimuli during auditory and audiovisual working memory [127,128]. Such multisensory PFC activity might enable human and nonhuman primates to recognize the correspondence between vocalizations and the facial postures associated with them [126–129].

Basic auditory tasks also activate the human vIPFC [130–132]. More-dorsal PFC regions (area 46/9) are utilized when verbal working memory is required, whereas vIPFC regions (47/12; 45) are recruited during active retrieval of verbal and nonverbal auditory information [133,134]. During the selective retrieval of information from auditory memory, the vIPFC interacts both with

the auditory temporal region (object information) and the inferior parietal lobule (spatial information) [134]. The broad involvement of the vIPFC in both verbal and nonverbal auditory tasks suggests that this area may be an interface in the auditory–vocal cycle. Collectively, these data suggest that vIPFC is specialized for processing and integrating social communication information in monkeys [135], in the same way as the human IFG is specialized for processing and integrating speech and gestures [136,137].

PFC Precursors of Semantics in Primate Referential Systems

The VAMN is much more than merely a high-order motor network. It also plays a vital role in establishing semantics and syntax, two hallmark characteristics of the two symbol systems – language and number theory. In symbolic reference, relations are established between spoken/written words or numbers, respectively, on the basis of compositional rules (i.e., syntax) [138,139]. However, simpler and both phylogenetically and ontogenetically earlier referential associations are ‘indices’ – signs that are characterized by spatial or temporal association between sign and object (reference based on contiguity or correlation) [140]. Animal communication typically is indexical, for example vervet monkey alarm calls indicate the presence of specific predator categories that have to be learned by juvenile monkeys [141]. Moreover, conditioned sign–object associations established in animals by reward contingencies are typically indexical, and can be investigated as a precursor for human symbolic reference [142].

Neurons in the vIPFC establish semantic associations: after training monkeys to associate the number of items in a set with arbitrary visual signs, many of the same IPFC neurons represent the abstract numerical meaning associated with such signs [143]. Similarly, fMRI studies show that the PFC is much more active in children learning semantic associations compared to proficient adults [144,145]. Damage to the human lateral frontal cortex results in severe impairment in tasks that require learning of arbitrary associations [146]. Networks within the lateral frontal cortex may thus fulfill the requirements for high-order associations between signs, ultimately giving rise to the cultural invention of linguistic and number symbols. Symbolic reference may thus emerge as a function of a largely expanded IPFC in humans [140].

Semantic associations require interactions of the PFC with the medial temporal lobe, which traditionally has been linked to declarative long-term memories [147]. Recordings in behaving monkeys show that, during learning and retrieval of long-term associations, the temporal lobe interacts with the PFC to store memories or reactivate information about past experiences [148,149]. With its link to the medial temporal lobe structures, the PFC is ideally suited to cognitively control memories, permitting an organism to establish sign reference and to strategically plan communicative acts in a flexible and goal-directed way.

PFC Precursors of Grammar in Primate Referential Systems

To establish a fully fledged symbol/language system, meaningful associations (semantics) are not sufficient. Sign sequences must be hierarchically structured according to action plans or rules guiding the structuring of signs – ‘syntax’. Syntax refers to the rules governing structure in natural language sentences or mathematical systems. Syntax establishes relations between signs that determine the meaning of an expression. Therefore, syntax and semantics are inextricably linked in symbolic systems.

Simple syntactical rules can be mastered by monkeys [150]. As a putative correlate for simple syntactical processing, single neurons in the primate IPFC encode sequence plans [151], the start and end states of behavioral sequences [152], and changing abstract rules [153,154]. Moreover, precursors of a code for abstract temporal structures of sounds and hand movements have been revealed by single-unit and fMRI studies in monkey PFC [155,156]. In humans, fMRI data suggest that Broca's area processes the start- and end-points of higher-order motor

segments and controls the nesting of functional segments, thereby forming the hierarchical structure of action plans [157]. By contrast, other speech-related areas, such as the SMA complex and the insula, are more specifically involved in the precise timing of ongoing motor acts underlying the execution of motor sequences needed for articulation [157–159].

Although neuronal circuits representing abstract sequences and rules are present in the monkey vIPFC, a grasp of high-level recursive tree structures seems to be unique to human language and mathematics [160–162]. Complex syntactical representations may have impinged on inferior frontal cortex networks [163]: while simple, non-recursive **grammar** (finite-state grammar) activates the phylogenetically-older frontal operculum (i.e., premotor cortex), the computation of recursive hierarchical sequences (phrase/structure grammar) that is characteristic of human language additionally recruits the phylogenetically-younger Broca's area (areas 44 and 45). Human fMRI studies suggest purely syntax-related activation in Broca's area, either in BA 45 and 47 [164], in BA 45 [165], or in BA 44 [166]. Using meta-analytic connectivity-based parcellation, Clos *et al.* [167] identified five functionally distinct clusters within left area 44 (associated with action processing, sequencing, linguistic working memory, meaning, and task-switching/cognitive control). While these functions are highly relevant in the context of language production and comprehension, they will obviously also be recruited by other domains including action and social cognition, therefore pointing to putative pre-adaptive functions in nonhuman primates.

Conclusions and Future Directions

In this review we suggest that the incipient linking of the prefrontal central executive of the brain with the vocalization system is a key neurobiological event and pre-adaptation for the evolution of speech and language. Crucial evidence for this hypothesis stems from comparative investigations of the cytoarchitecture of the human and the monkey vIPFC. These studies demonstrated that the basic architectonic plans are similar in these two primate brains, despite considerable development of the vIPFC areas in the human brain [57,78,168]. In addition to anatomical similarities, recent neurophysiological experiments in nonhuman primates provide the necessary functional evidence. Studies in behaving macaque monkeys showed that neuronal activity in vIPFC is correlated with volitional call initiation [76]. Although these correlative measures have led to novel insights concerning the role of inferior frontal lobe structures in controlling vocal output, the causal efficacy of vocalization-related activity needs to be addressed in the future. This could be achieved by probing the direct impact of experimental neural perturbations (stimulation or inhibition) on vocal behavior [169]. If physiologically characterized vocalization-related neurons are causally involved in eliciting cued vocalizations, lower vocalization thresholds (i.e., higher vocalization rates) can be expected while electrically stimulating such neurons [170], whereas their transient chemical inactivation (i.e., by increasing synaptic inhibition) is predicted to temporarily impair volitional call initiation. Such coarse hypotheses are of course subject to adjustments and refinements with increasing knowledge about the cortical vocal network.

As indicated by the (simplified) connections in our model, the vIPFC is the central executive of the VAMN. At the same time, however, it likely interacts with other areas of a larger frontal lobe network to encode cognitively controlled vocalizations. Medial frontal lobe areas, such as the anterior cingulate cortex (ACC) and the pre-supplementary motor area (pre-SMA), might play additional roles not only in affective vocal output but also in controlling volitional vocalizations [5,29,171,172]. Data from the ACC and pre-SMA, among other areas, will be necessary to complete the emerging picture of a frontal vocal network.

A third important aspect is to clarify the level of specialization of frontal lobe circuitry in initiating goal-directed vocalizations. Instead of encoding any volitional motor act, neurons of the VAMN are expected to show some specificity towards controlling vocal output. To decipher the

Outstanding Questions

How does activation or inactivation of distinct areas of the monkey homolog of the human Broca's area deteriorate volitional vocal output?

Do preSMA, SMA, and the insula play a significant role in cognitive control of vocal output in non-human primates?

How is the VAMN interconnected, and how is it linked to the PMVN, during development from prelinguistic infants to linguistic children?

What role do auditory feedback mechanisms play in shaping vocal patterns in non-human primates? How does the PFC participate in potential vocal learning mechanisms?

vocalization-specific aspects of the VAMN, neurophysiological investigations in monkeys trained to perform volitional acts using not only the vocal apparatus but also other effector organs (e.g., hand movements) would be appropriate. This would allow the dedicated role of the VAMN *vis-à-vis* other volitional behaviors to be investigated. However, that neurons might respond exclusively to volitional vocal output does not appear to be realistic. After all, the primate vIPFC operates at the apex of the cortical hierarchy and is involved in a variety of executive functions in different domains [9, 173–175]. Despite its reputation as classical language area, even Broca's area in humans is part of a larger cognitive control network and plays important roles in structuring a variety of hierarchically organized behaviors [157, 176]. Neurophysiological studies in nonhuman primates will remain indispensable for identifying prefrontal cognitive control functions as well as specific aspects of human speech and language functions.

Acknowledgements

We thank Barbara Peysakhovich for proofreading. This work was supported by the Werner Reichardt Centre for Integrative Neuroscience (CIN) at the Eberhard Karls University of Tübingen (CIN is an Excellence Cluster funded by the Deutsche Forschungsgemeinschaft within the frame-work of the Excellence Initiative EXC 307).

References

- Balter, M. (2010) Animal communication helps reveal roots of language. *Science* 328, 969–971
- Hammerschmidt, K. and Fischer, J. (2008) Constraints in primate vocal production. In *Evolution of Communicative Flexibility: Complexity, Creativity, and Adaptability in Human and Animal Communication* (Oller, D.K. and Griebel, U., eds), pp. 93–121, MIT Press
- Ghazanfar, A.A. (2008) Language evolution: neural differences that make the difference. *Nat. Neurosci.* 11, 382–384
- Ackermann, H. et al. (2014) Brain mechanisms of acoustic communication in humans and nonhuman primates: an evolutionary perspective. *Behav. Brain Sci.* 37, 529–546
- Jürgens, U. (2002) Neural pathways underlying vocal control. *Neurosci. Biobehav. Rev.* 26, 235–258
- Deacon, T.W. (1992) The neural circuitry underlying primate calls and human language. In *Language Origin: A Multidisciplinary Approach* (Wind, J. et al., eds), pp. 121–162, Springer
- Owren, M.J. et al. (2011) Two organizing principles of vocal production: implications for nonhuman and human primates. *Am. J. Primatol.* 73, 530–544
- Holstege, G. and Subramanian, H.H. (2016) Two different motor systems are needed to generate human speech. *J. Comp. Neurol.* 524, 1558–1577
- Miller, E.K. and Cohen, J.D. (2001) An integrative theory of prefrontal cortex function. *Annu. Rev. Neurosci.* 24, 167–202
- Jürgens, U. (2009) The neural control of vocalization in mammals: a review. *J. Voice* 23, 1–10
- Bass, A.H. and Chagnaud, B.P. (2012) Shared developmental and evolutionary origins for neural basis of vocal-acoustic and pectoral-gestural signaling. *Proc. Natl. Acad. Sci. U. S. A.* 109 (Suppl. 1), 10677–10684
- Jürgens, U. (2000) Localization of a pontine vocalization-controlling area. *J. Acoust. Soc. Am.* 108, 1393–1396
- Lüthe, L. et al. (2000) Neuronal activity in the medulla oblongata during vocalization. A single-unit recording study in the squirrel monkey. *Behav. Brain Res.* 116, 197–210
- Hannig, S. and Jürgens, U. (2006) Projections of the ventrolateral pontine vocalization area in the squirrel monkey. *Exp. Brain Res.* 169, 92–105
- Hage, S.R. and Jürgens, U. (2006) On the role of the pontine brainstem in vocal pattern generation. A telemetric single-unit recording study in the squirrel monkey. *J. Neurosci.* 26, 7105–7115
- Hage, S.R. and Jürgens, U. (2006) Localization of a vocal pattern generator in the pontine brainstem of the squirrel monkey. *Eur. J. Neurosci.* 23, 840–844
- Larson, C.R. and Kistler, M.K. (1984) Periaqueductal gray neuronal activity associated with laryngeal EMG and vocalization in the awake monkey. *Neurosci. Lett.* 46, 261–266
- Kirzinger, A. and Jürgens, U. (1991) Vocalization-correlated single-unit activity in the brain stem of the squirrel monkey. *Exp. Brain Res.* 84, 545–560
- Düsterhöft, F. et al. (2004) Neuronal activity in the periaqueductal gray and bordering structures during vocal communication in the squirrel monkey. *Neuroscience* 123, 53–60
- Jürgens, U. and Ploog, D. (1970) Cerebral representation of vocalization in the squirrel monkey. *Exp. Brain Res.* 10, 532–554
- Lu, C.L. and Jürgens, U. (1993) Effects of chemical stimulation in the periaqueductal gray on vocalization in the squirrel monkey. *Brain Res. Bull.* 32, 143–151
- Adamez, J. and O'Leary, J.L. (1959) Experimental mutism resulting from periaqueductal lesions in cats. *Neurology* 9, 636–642
- Jürgens, U. and Pratt, R. (1979) Role of the periaqueductal grey in vocal expression of emotion. *Brain Res.* 167, 367–378
- Jürgens, U. and Müller-Preuss, P. (1977) Convergent projections of different limbic vocalization areas in the squirrel monkey. *Brain Res.* 29, 75–83
- Dujardin, E. and Jürgens, U. (2005) Afferents of vocalization-controlling periaqueductal regions in the squirrel monkey. *Brain Res.* 1034, 114–131
- Dujardin, E. and Jürgens, U. (2006) Call type-specific differences in vocalization-related afferents to the periaqueductal gray of squirrel monkeys (*Saimiri sciureus*). *Behav. Brain Res.* 168, 23–36
- Smith, W.K. (1945) The functional significance of the rostral cingulate cortex as revealed by its responses to electrical excitation. *J. Neurophysiol.* 8, 241–255
- Apfelbach, R. (1972) Electrically elicited vocalizations in the gibbon *Hylobates lar* (Hylobatidae), and their behavioral significance. *Z. Tierpsychol.* 30, 420–430
- Vogt, B.A. and Barbas, H. (1988) Structure and connections of the cingulate vocalization region in the rhesus monkey. In *The Physiological Control of Mammalian Vocalization* (Newman, J.D., ed.), pp. 203–225, Plenum Press
- Jürgens, U. (1979) Vocalization as an emotional indicator. A neuroethological study in the squirrel monkey. *Behaviour* 69, 88–117
- Jürgens, U. (1982) Amygdalar vocalization pathways in the squirrel monkey. *Brain Res.* 241, 189–196
- Kirzinger, A. and Jürgens, U. (1982) Cortical lesion effects and vocalization in the squirrel monkey. *Brain Res.* 233, 299–315

33. Trachy, R.E. *et al.* (1981) Primate phonation: anterior cingulate lesion effects on response rate and acoustical structure. *Am. J. Primatol.* 1, 43–55
34. Franzen, E.A. and Myers, R.E. (1973) Neural control of social behavior: prefrontal and anterior temporal cortex. *Neuropsychology* 11, 141–157
35. Aitken, P.G. (1981) Cortical control of conditioned and spontaneous vocal behaviour in rhesus monkeys. *Brain Lang.* 13, 171–184
36. Sutton, D. *et al.* (1974) Neocortical and limbic lesion effects on primate phonation. *Brain Res.* 71, 61–75
37. Jürgens, U. *et al.* (1982) The effects of deep-reaching lesions in the cortical face area on phonation: a combined case report and experimental monkey study. *Cortex* 18, 125–139
38. Cantu, R.C. (1966) Importance of pathological laughing and/or crying as a sign of occurrence or recurrence of a tumor lying beneath the brainstem. *J. Nerv. Ment. Dis.* 143, 508–512
39. Wild, B. *et al.* (2003) Neural correlates of laughter and humour. *Brain* 126, 2121–2138
40. Kovac, S. *et al.* (2009) Gelastic seizures: a case of lateral frontal lobe epilepsy and review of the literature. *Epilepsy Behav.* 15, 249–253
41. Kuzniecky, R. *et al.* (1997) Intrinsic epileptogenesis of hypothalamic hamartomas in gelastic epilepsy. *Ann. Neurol.* 42, 60–67
42. Sperli, F. *et al.* (2006) Contralateral smile and laughter, but no mirth, induced by electrical stimulation of the cingulate cortex. *Epilepsia* 47, 440–443
43. Nielsen, J.M. and Jacobs, L.L. (1951) Bilateral lesions of the anterior cingulate gyri; report of case. *Bull. Los Angel. Neuro. Soc.* 16, 231–234
44. Barris, R.W. and Schuman, H.R. (1953) Bilateral anterior cingulate gyrus lesions; syndrome of the anterior cingulate gyri. *Neurology* 3, 44–52
45. Paus, T. (2001) Primate anterior cingulate cortex: where motor control, drive and cognition interface. *Nat. Rev. Neurosci.* 2, 417–424
46. Frühholz, S. *et al.* (2015) Talking in fury: the cortico-subcortical network underlying angry vocalizations. *Cereb. Cortex* 25, 2752–2762
47. Nagaratnam, N. *et al.* (1999) Akinetic mutism and mixed transcortical aphasia following left thalamo-mesencephalic infarction. *J. Neurol. Sci.* 163, 70–73
48. Nagaratnam, N. *et al.* (2004) Akinetic mutism following stroke. *J. Clin. Neurosci* 11, 25–30
49. Esposito, A. *et al.* (1999) Complete mutism after midbrain periaqueductal gray lesion. *Neuroreport* 10, 681–685
50. Wattendorf, E. *et al.* (2013) Exploration of the neural correlates of ticklish laughter by functional magnetic resonance imaging. *Cereb. Cortex* 23, 1280–1289
51. Schulz, G.M. *et al.* (2005) Functional neuroanatomy of human vocalization: an H₂¹⁵O PET study. *Cereb. Cortex* 15, 1835–1847
52. Deacon, T.W. (1992) The neural circuitry underlying primate calls and human language. In *Language Origin: A Multidisciplinary Approach* (Wind, J. *et al.*, eds), pp. 121–162, Kluwer Academic Publishers
53. Preuss, T.M. (2006) Evolutionary specializations of primate brain systems. In *Primate Origins and Adaptations* (Ravoso, M.J. and Dagosto, M., eds), pp. 625–675, Kluwer Academic/Plenum Press
54. Wise, S.P. (2008) Forward frontal fields: phylogeny and fundamental function. *Trends Neurosci.* 31, 599–608
55. Brodmann, K. ed. (1909) *Vergleichende Lokalisationslehre der Grosshirnrinde in ihren Prinzipien dargestellt auf Grund des Zellenbaues*, Barth JA
56. Broca, P. (1861) Remarques sur le siege de la faculté du langage articulé, suivies d'une observation d'aphémie (perte de la parole) [Remarks on the seat of the faculty of articulated language, following an observation of aphemia (loss of speech)]. *Bull. Mem. Soc. Anat. Paris* 36, 330–357
57. Petrides, M. and Pandya, D.N. (2002) Comparative cytoarchitectonic analysis of the human and the macaque ventrolateral prefrontal cortex and corticocortical connection patterns in the monkey. *Eur. J. Neurosci.* 16, 291–310
58. Preuss, T.M. (2000) What's human about the human brain? In *The New Cognitive Neurosciences* (Gazzaniga, M.S., ed.), pp. 1219–1234, MIT Press
59. Gerbella, M. *et al.* (2011) Cortical connections of the anterior (F5a) subdivision of the macaque ventral premotor area F5. *Brain Struct. Funct.* 216, 43–65
60. Kumar, V. *et al.* (2016) Structural organization of the laryngeal motor cortical network and its implication for evolution of speech and language. *J. Neurosci.* 36, 4170–4181
61. Rizzolatti, G. *et al.* (1981) Response properties and behavioural modulation of 'mouth' neurons of the postarcuate. *Brain Res.* 255, 421–424
62. Dum, R.P. and Strick, P.L. (1991) The origin of corticospinal projections from the premotor areas in the frontal lobe. *J. Neurosci.* 11, 667–689
63. Simonyan, K. and Horwitz, B. (2011) Laryngeal motor cortex and control of speech in humans. *Neuroscientist* 17, 197–208
64. Song, X. *et al.* (2016) Complex pitch perception mechanisms are shared by humans and a New World monkey. *Proc. Natl. Acad. Sci. U. S. A.* 113, 781–786
65. Coude, G. *et al.* (2011) Neurons controlling voluntary vocalization in the macaque ventral premotor cortex. *PLoS One* 6, e26822
66. Simonyan, K. (2014) The laryngeal motor cortex: its organization and connectivity. *Curr. Opin. Neurobiol.* 28, 15–21
67. Fuertinger, S. *et al.* (2015) The functional connectome of speech control. *PLoS Biol.* e1002209
68. Groswasser, Z. *et al.* (1988) Mutism associated with buccofacial apraxia and bihemispheric lesions. *Brain Lang.* 34, 157–168
69. Green, H.D. and Walker, A.E. (1938) The effects of ablation of the cortical motor face area in monkeys. *J. Neurophysiol.* 1, 262–280
70. Kaada, B.R. (1951) Somato-motor, autonomic and electro-corticographic responses to electrical stimulation of 'rhinencephalic' and other structures in primates, cat and dog. *Acta Physiol. Scand.* 24 (Suppl. 83), 1–285
71. Robinson, B.W. (1967) Vocalization evoked from forebrain in *Macaca mulatta*. *Physiol Behav* 2, 345–354
72. Leyton, A.S.F. and Sherrington, C.S. (1917) Observations on the excitable cortex of the chimpanzee, orangutan, and gorilla. *Quart. J. Exp. Biol.* 11, 135–222
73. Dusser de Barenne, J. *et al.* (1941) The 'motor' cortex of the chimpanzee. *J. Neurophysiol.* 4, 287–303
74. Hines, M. (1940) Movements elicited from precentral gyrus of adult chimpanzees by stimulation with sine wave currents. *J. Neurophysiol.* 3, 442–466
75. Gemba, H. *et al.* (1999) Cortical field potentials associated with audio-initiated vocalization in monkeys. *Neurosci. Lett.* 272, 49–52
76. Hage, S.R. and Nieder, A. (2013) Single neurons in monkey prefrontal cortex encode volitional initiation of vocalizations. *Nat. Commun.* 4, 2409
77. Fukushima, M. *et al.* (2014) Modeling vocalization with ECoG cortical activity recorded during vocal production in the macaque monkey. *Conf. Proc. IEEE Eng. Med. Biol. Soc.* 2014, 6794–6797
78. Petrides, M. and Pandya, D.N. (1999) Dorsolateral prefrontal cortex: comparative cytoarchitectonic analysis in the human and the macaque brain and corticocortical connection patterns. *Eur. J. Neurosci.* 11, 1011–1036
79. Simonyan, K. and Jürgens, U. (2002) Cortico-cortical projections of the motorcortical larynx area in the rhesus monkey. *Brain Res.* 949, 23–31
80. Yeterian, E.H. *et al.* (2012) The cortical connectivity of the prefrontal cortex in the monkey brain. *Cortex* 48, 58–81
81. Simonyan, K. and Jürgens, U. (2003) Efferent subcortical projections of the laryngeal motorcortex in the rhesus monkey. *Brain Res.* 974, 43–59
82. Petrides, M. *et al.* (2005) Orofacial somatomotor responses in the macaque monkey homologue of Broca's area. *Nature* 435, 1235–1238

83. Hast, M.H. *et al.* (1974) Cortical motor representation of the laryngeal muscles in *Macaca mulatta*. *Brain Res.* 73, 229–240
84. Sutton, D. *et al.* (1973) Vocalization in rhesus monkeys: conditionability. *Brain Res.* 52, 225–231
85. Larson, C.R. *et al.* (1973) Sound spectral properties of conditioned vocalization in monkeys. *Phonetica* 27, 100–110
86. Hage, S.R. *et al.* (2013) Cognitive control of distinct vocalizations in rhesus monkeys. *J. Cogn. Neurosci.* 25, 1692–1701
87. Lemon, R.N. (2008) Descending pathways in motor control. *Ann. Rev. Neurosci.* 31, 195–218
88. Jürgens, U. and Zwirner, P. (1996) The role of the periaqueductal grey in limbic and neocortical vocal fold control. *Neuroreport* 7, 2921–2923
89. Jürgens, U. and Ehrenreich, L. (2007) The descending motor-cortical pathway to the laryngeal motoneurons in the squirrel monkey. *Brain Res.* 1148, 90–95
90. McCairn, K.W. *et al.* (2016) A primary role for nucleus accumbens and related limbic network in vocal tics. *Neuron* 89, 300–307
91. Lauterbach, E.C. *et al.* (2013) Toward a more precise, clinically-informed pathophysiology of pathological laughing and crying. *Neurosci. Biobehav. Rev.* 37, 1893–1916
92. Caplan, D. *et al.* (1996) Location of lesions in stroke patients with deficits in syntactic processing in sentence comprehension. *Brain* 119, 933–994
93. Lazar, R.M. and Mohr, J.P. (2011) Revisiting the contributions of Paul Broca to the study of aphasia. *Neuropsychol. Rev.* 21, 236–239
94. Rohrer, J.D. *et al.* (2009) Abnormal laughter-like vocalisations replacing speech in primary progressive aphasia. *J. Neurol. Sci.* 284, 120–123
95. Norris, M.R. and Drummond, S.S. (1998) Communicative functions of laughter in aphasia. *J. Neuroling.* 11, 391–402
96. Penfield, W. and Roberts, L. (1959) *Speech and Brain Mechanisms*, Princeton University Press
97. Epstein, C.M. *et al.* (1999) Localization and characterization of speech arrest during transcranial magnetic stimulation. *Clin. Neurophysiol.* 110, 1073–1079
98. Axelson, H.W. *et al.* (2009) Successful localization of the Broca area with short-train pulses instead of 'Penfield' stimulation. *Seizure* 18, 374–375
99. Flinker, A. *et al.* (2015) Redefining the role of Broca's area in speech. *Proc. Natl. Acad. Sci. U. S. A.* 112, 2871–2875
100. Dronkers, N.F. and Baldo, J.V. (2010) Broca's area. In *The Cambridge Encyclopedia of the Language Sciences* (Hogan, P.C., ed.), pp. 139–142, Cambridge University Press
101. Long, M.A. *et al.* (2016) Functional segregation of cortical regions underlying speech timing and articulation. *Neuron* 89, 1187–1193
102. Rauschecker, J.P. and Scott, S.K. (2009) Maps and streams in the auditory cortex: nonhuman primates illuminate human speech processing. *Nat. Neurosci.* 12, 718–724
103. Romanski, L.M. *et al.* (1999) Dual streams of auditory afferents target multiple domains in the primate prefrontal cortex. *Nat. Neurosci.* 2, 1131–1136
104. Nieder, A. (2012) Supramodal numerosity selectivity of neurons in primate prefrontal and posterior parietal cortices. *Proc. Natl. Acad. Sci. U. S. A.* 109, 11860–11865
105. Hackett, T.A. *et al.* (1998) Thalamocortical connections of the parabelt auditory cortex in macaque monkeys. *J. Comp. Neurol.* 400, 271–286
106. Poremba, A. *et al.* (2004) Species-specific calls evoke asymmetric activity in the monkey's temporal poles. *Nature* 427, 448–451
107. Gil-da-Costa, R. *et al.* (2006) Species-specific calls activate homologs of Broca's and Wernicke's areas in the macaque. *Nat. Neurosci.* 9, 1064–1070
108. Perrodin, C. *et al.* (2011) Voice cells in the primate temporal lobe. *Curr. Biol.* 21, 1408–1415
109. Romanski, L.M. and Goldman-Rakic, P.S. (2002) An auditory domain in primate prefrontal cortex. *Nat. Neurosci.* 5, 15–16
110. Romanski, L.M. *et al.* (2005) Neural representation of vocalizations in the primate ventrolateral prefrontal cortex. *J. Neurophysiol.* 93, 734–747
111. Hage, S.R. and Nieder, A. (2015) Audio-vocal interaction in single neurons of the monkey ventrolateral prefrontal cortex. *J. Neurosci.* 35, 7030–7040
112. Averbeck, B.B. and Romanski, L.M. (2006) Probabilistic encoding of vocalizations in macaque ventral lateral prefrontal cortex. *J. Neurosci.* 26, 11023–11033
113. Medalla, M. and Barbas, H. (2014) Specialized prefrontal 'auditory fields': organization of primate prefrontal-temporal pathways. *Front. Neurosci.* 8, 77
114. Mishkin, M. *et al.* (1983) Object vision and spatial vision: two cortical pathways. *Trends Neurosci.* 6, 414–417
115. Goodale, M.A. and Milner, A.D. (1992) Separate visual pathways for perception and action. *Trends Neurosci.* 15, 20–25
116. Gross, C.G. *et al.* (1972) Visual properties of neurons in inferotemporal cortex of the macaque. *J. Neurophysiol.* 35, 96–111
117. Desimone, R. *et al.* (1984) Stimulus-selective properties of inferior temporal neurons in the macaque. *J. Neurosci.* 4, 2051–2062
118. O'Scalaidhe, S.P. *et al.* (1997) Areal segregation of face-processing neurons in prefrontal cortex. *Science* 278, 1135–1138
119. Tsao, D.Y. *et al.* (2008) Patches of face-selective cortex in the macaque frontal lobe. *Nat. Neurosci.* 11, 877–879
120. Freedman, D.J. *et al.* (2001) Categorical representation of visual stimuli in the primate prefrontal cortex. *Science* 291, 312–316
121. Nieder, A. (2016) The neuronal code for number. *Nat. Rev. Neurosci.* 17, 366–382
122. Russ, B.E. *et al.* (2008) Prefrontal neurons predict choices during an auditory samedifferent task. *Curr. Biol.* 18, 1483–1488
123. Lee, J.H. *et al.* (2009) Prefrontal activity predicts monkeys' decisions during an auditory category task. *Front. Integr. Neurosci.* 3, 16
124. Sugihara, T. *et al.* (2006) Integration of auditory and visual communication information in the primate ventrolateral prefrontal cortex. *J. Neurosci.* 26, 11138–11147
125. Romanski, L.M. and Hwang, J. (2012) Timing of audiovisual inputs to the prefrontal cortex and multisensory integration. *Neuroscience* 214, 36–48
126. Diehl, M.M. and Romanski, L.M. (2014) Responses of prefrontal multisensory neurons to mismatching faces and vocalizations. *J. Neurosci.* 34, 11233–11243
127. Hwang, J. and Romanski, L.M. (2015) Prefrontal neuronal responses during audiovisual mnemonic processing. *J. Neurosci.* 35, 960–971
128. Plakke, B. *et al.* (2015) Inactivation of primate prefrontal cortex impairs auditory and audiovisual working memory. *J. Neurosci.* 35, 9666–9675
129. Ghazanfar, A.A. and Takahashi, D.Y. (2014) The evolution of speech: vision, rhythm, cooperation. *Trends Cogn. Sci.* 18, 543–553
130. Zatorre, R.J. *et al.* (1994) Neural mechanisms underlying melodic perception and memory for pitch. *J. Neurosci.* 14, 1908–1919
131. Grady, C.L. *et al.* (2008) Age-related differences in brain activity underlying working memory for spatial and nonspatial auditory information. *Cereb. Cortex* 18, 189–199
132. Protzner, A.B. and McIntosh, A.R. (2009) Modulation of ventral prefrontal cortex functional connections reflects the interplay of cognitive processes and stimulus characteristics. *Cereb. Cortex* 19, 1042–1054
133. Petrides, M. (1996) Specialized systems for the processing of mnemonic information within the primate frontal cortex. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 351, 1455–1461
134. Kostopoulos, P. and Petrides, M. (2016) Selective memory retrieval of auditory what and auditory where involves the ventrolateral prefrontal cortex. *Proc. Natl. Acad. Sci. U. S. A.* 113, 1919–1924
135. Romanski, L.M. (2012) Integration of faces and vocalizations in ventral prefrontal cortex: implications for the evolution of audiovisual speech. *Proc. Natl. Acad. Sci. U. S. A.* 109 (Suppl. 1), 10717–10724

136. Homae, F. *et al.* (2002) From perception to sentence comprehension: the convergence of auditory and visual information of language in the left inferior frontal cortex. *Neuroimage* 16, 883–900
137. Xu, J. *et al.* (2009) Symbolic gestures and spoken language are processed by a common neural system. *Proc. Natl. Acad. Sci. U. S. A.* 106, 20664–20669
138. Deacon, T.W. (1996) Prefrontal cortex and symbol learning: why a brain capable of language evolved only once. In *Communicating Meaning. The Evolution and Development of Language* (Velichkovsky, B.M. and Rumbaugh, D.M., eds), pp. 103–138, Lawrence Erlbaum Associates
139. Wiese, H. (2003) *Numbers, Language, and the Human Mind*, Cambridge University Press
140. Deacon, T.W. (1997) *The Symbolic Species: The Co-evolution of Language and the Human Brain*, W. W. Norton & Company
141. Cheney, D.L. and Seyfarth, R.M. (1992) *How Monkeys See The World: Inside the Mind of Another Species*, University of Chicago Press
142. Nieder, A. (2009) Prefrontal cortex and the evolution of symbolic reference. *Curr. Opin. Neurobiol.* 19, 99–108
143. Diester, I. and Nieder, A. (2007) Semantic associations between signs and numerical categories in the prefrontal cortex. *PLoS Biol.* 5, e294
144. Ansari, D. *et al.* (2005) Neural correlates of symbolic number processing in children and adults. *Neuroreport* 16, 1769–1773
145. Cantlon, J.F. *et al.* (2009) The neural development of an abstract concept of number. *J. Cogn. Neurosci.* 21, 2217–2229
146. Petrides, M. (1985) Deficits on conditional associative-learning tasks after frontal- and temporal-lobe lesions in man. *Neuropsychologia* 23, 601–614
147. Simons, J.S. and Spiers, H.J. (2003) Prefrontal and medial temporal lobe interactions in long-term memory. *Nat. Rev. Neurosci.* 4, 637–648
148. Tomita, H. *et al.* (1999) Top-down signal from prefrontal cortex in executive control of memory retrieval. *Nature* 401, 699–703
149. Brincat, S.L. and Miller, E.K. (2015) Frequency-specific hippocampal-prefrontal interactions during associative learning. *Nat. Neurosci.* 18, 576–581
150. Wilson, B. *et al.* (2013) Auditory artificial grammar learning in macaque and marmoset monkeys. *J. Neurosci.* 33, 18825–18835
151. Mushiake, H. *et al.* (2006) Activity in the lateral prefrontal cortex reflects multiple steps of future events in action plans. *Neuron* 50, 631–641
152. Fujii, N. and Graybiel, A.M. (2003) Representation of action sequence boundaries by macaque prefrontal cortical neurons. *Science* 301, 1246–1249
153. Wallis, J.D. *et al.* (2001) Single neurons in prefrontal cortex encode abstract rules. *Nature* 411, 953–956
154. Bongard, S. and Nieder, A. (2010) Basic mathematical rules are encoded by primate prefrontal cortex neurons. *Proc. Natl. Acad. Sci. U. S. A.* 107, 2277–2282
155. Shima, K. *et al.* (2007) Categorization of behavioural sequences in the prefrontal cortex. *Nature* 445, 315–318
156. Wang, L. *et al.* (2015) Representation of numerical and sequential patterns in macaque and human brains. *Curr. Biol.* 25, 1966–1974
157. Koehlin, E. and Jubault, T. (2006) Broca's area and the hierarchical organization of human behavior. *Neuron* 50, 963–974
158. Dronkers, N.F. (1996) A new brain region for coordinating speech articulation. *Nature* 384, 159–161
159. Ackermann, H. and Riecker, A. (2004) The contribution of the insula to motor aspects of speech production: a review and a hypothesis. *Brain Lang.* 89, 320–328
160. Chomsky, N. (1956) Three models for the description of language. *IEEE Trans. Inf. Theory* 2, 113–124
161. Penn, D.C. *et al.* (2008) Darwin's mistake: explaining the discontinuity between human and nonhuman minds. *Behav. Brain Sci.* 31, 109–130
162. Dehaene, S. *et al.* (2015) The neural representation of sequences: from transition probabilities to algebraic patterns and linguistic trees. *Neuron* 88, 2–19
163. Friederici, A.D. *et al.* (2006) The brain differentiates human and non-human grammars: functional localization and structural connectivity. *Proc. Natl. Acad. Sci. U. S. A.* 103, 2458–2463
164. Pallier, C. *et al.* (2011) Cortical representation of the constituent structure of sentences. *Proc. Natl. Acad. Sci. U. S. A.* 108, 2522–2527
165. Shetreet, E. *et al.* (2009) An fMRI study of syntactic layers: sentential and lexical aspects of embedding. *Neuroimage* 48, 707–716
166. Goucha, T. and Friederici, A.D. (2015) The language skeleton after dissecting meaning: A functional segregation within Broca's Area. *Neuroimage* 114, 294–302
167. Cios, M. *et al.* (2013) Tackling the multifunctional nature of Broca's region meta-analytically: co-activationbased parcellation of area 44. *Neuroimage* 83, 174–188
168. Petrides, M. and Pandya, D.N. (2009) Distinct parietal and temporal pathways to the homologues of Broca's area in the monkey. *PLoS Biol.* 7, e1000170
169. Wurtz, R.H. (2015) Using perturbations to identify the brain circuits underlying active vision. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 370, 20140205
170. Cohen, M.R. and Newsome, W.T. (2004) What electrical microstimulation has revealed about the neural basis of cognition. *Curr. Opin. Neurobiol.* 14, 169–177
171. West, R.A. and Larson, C.R. (1995) Neurons of the anterior mesial cortex related to faciavocal activity in the awake monkey. *J. Neurophysiol.* 74, 1856–1869
172. Alario, F.X. *et al.* (2006) The role of the supplementary motor area (SMA) in word production. *Brain Res.* 1076, 129–143
173. Merten, K. and Nieder, A. (2012) Active encoding of decisions about stimulus absence in primate prefrontal cortex neurons. *Proc. Natl. Acad. Sci. U. S. A.* 109, 6289–6294
174. Vallentin, D. *et al.* (2012) Numerical rule coding in the prefrontal, premotor, and posterior parietal cortices of macaques. *J. Neurosci.* 32, 6621–6630
175. Jacob, S.N. and Nieder, A. (2014) Complementary roles for primate frontal and parietal cortex in guarding working memory from distractor stimuli. *Neuron* 83, 226–237
176. Fazio, P. *et al.* (2009) Encoding of human action in Broca's area. *Brain* 132, 1980–1988
177. Winter, P. *et al.* (1973) Ontogeny of squirrel monkey calls under normal conditions and under acoustic isolation. *Behaviour* 47, 230–239
178. Hammerschmidt, K. *et al.* (2001) Vocal development in squirrel monkeys. *Behaviour* 138, 1179–1204
179. Talmage-Riggs, G. *et al.* (1972) Effect of deafening on the vocal behavior of the squirrel monkey (*Saimiri sciureus*). *Folia Primatol.* 17, 404–420
180. Aitken, P.G. and Wilson, D.A. (1979) Discriminative vocal conditioning in Rhesus monkeys: evidence for volitional control? *Brain Lang.* 8, 227–240
181. Hihara, S. *et al.* (2003) Spontaneous vocal differentiation of co-calls for tools and food in Japanese monkeys. *Neurosci. Res.* 45, 383–389
182. Koda, H. *et al.* (2007) Experimental evidence for the volitional control of vocal production in an immature gibbon. *Behaviour* 144, 681–692
183. Sutton, D. *et al.* (1985) Discriminative phonation in macaques: effects of anterior mesial cortex damage. *Exp. Brain Res.* 59, 410–413
184. Cheney, D.L. and Seyfarth, R.M., eds (2007) *Baboon Metaphysics: The Evolution of a Social Mind*, University of Chicago Press
185. Seyfarth, R.M. *et al.* (1980) Monkey responses to three different alarm calls: evidence for predator classification and semantic communication. *Science* 210, 801–803
186. Wich, S.A. and de Vries, H. (2006) Male monkeys remember which group members have given alarm calls. *Proc. Biol. Sci.* 273, 735–740

187. Ouattara, K. *et al.* (2009) Campbell's monkeys concatenate vocalizations into context-specific call sequences. *Proc. Natl. Acad. Sci. U. S. A.* 106, 22026–22031
188. Eliades, S.J. and Wang, X. (2005) Dynamics of auditory–vocal interaction in monkey auditory cortex. *Cereb. Cortex* 15, 1510–1523
189. Choi, J.Y. *et al.* (2015) Cooperative vocal control in marmoset monkeys via vocal feedback. *J. Neurophysiol.* 114, 274–283
190. Takahashi, D.Y. *et al.* (2015) The developmental dynamics of marmoset monkey vocal production. *Science* 349, 734–738
191. Takahashi, D.Y. *et al.* (2016) Early development of turn-taking with parents shapes vocal acoustics in infant marmoset monkeys. *Phil. Trans. R. Soc. B* 371, 20150370
192. Gould, S.J. ed. (1977) *Ontogeny and Phylogeny*, Harvard University Press
193. Hage, S.R. *et al.* (2016) Developmental changes of cognitive vocal control in monkeys. *J. Exp. Biol.* 219, 1744–1749
194. Carroll, S.B. (2003) Genetics and the making of *Homo sapiens*. *Nature* 422, 849–857
195. Oller, D.K. ed. (2000) *The Emergence of the Speech Capacity*, Psychology Press
196. Petanjek, Z. *et al.* (2011) Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc. Natl. Acad. Sci. U. S. A.* 108, 13281–13286
197. Somel, M. *et al.* (2013) Human brain evolution: transcripts, metabolites and their regulators. *Nat. Rev. Neurosci.* 14, 112–127
198. Bourgeois, J.P. *et al.* (1994) Synaptogenesis in the prefrontal cortex of rhesus monkeys. *Cereb. Cortex* 4, 78–96
199. Huttenlocher, P.R. and Dabholkar, A.S. (1997) Regional differences in synaptogenesis in human cerebral cortex. *J. Comp. Neurol.* 387, 167–178
200. Hickok, G. and Poeppel, D. (2007) The cortical organization of speech processing. *Nat. Rev. Neurosci.* 8, 393–402